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13 NOV 2000

EXPRESS MAIL CERTIFICATE

I hereby certify that this paper or fee is being deposited with the United States Postal Service "Express Mail Post Office to Address" service under 37 C.F.R. § 1.10 on the date indicated above and is addressed to the Assistant Commissioner for Patents, Washington, D.C. 20231.

09/700354

Typed or Printed Name	Mathew GHS	Express Mail No.	EL 563 650 845 US
Signature	<i>Mathew GHS</i>	Date	November 13, 2000

Form PTO-1390
(REV 10-94)

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER
IRVN-007CIP2

TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371

U.S. APPLICATION NO.

INTERNATIONAL APPLICATION NO.
PCT US99/10793INTERNATIONAL FILING DATE
May 14, 1999PRIORITY DATE CLAIMED
May 14, 1998

TITLE OF INVENTION: Factors Affecting Tumor Necrosis Factor Receptor Releasing Enzyme Activity

APPLICANT(S) FOR DO/EO/US (The Regents of the University of California)

GATANAGA Tetsuya

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This express request to begin examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and the PCT Articles 22 and 39(1).
4. ☒ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2)):
 - a. ☐ is transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☒ has been transmitted by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☐ A translation of the International Application into English (35 U.S.C. 371(c)(2)).
7. ☒ Amendments to the claims of the International Application under PCT Article 19 (37 U.S.C. 371(c)(3)):
 - a. ☒ are transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ have been transmitted by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☐ have not been made and will not be made.
8. ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)). **(SIGNED)**
10. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11 to 16 below concern other document(s) or information included:

11. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
12. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. ☒ A **FIRST** preliminary amendment.
☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
14. ☐ A substitute specification.
15. ☐ A change of power of attorney and/or address letter.
16. ☐ Other items or information:

17. ☒ The following fees are submitted:**Basic National Fee (37 CFR 1.492(a)(1)-(5)):**

Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and international search report not prepared by the EPO or JPO \$970.00

International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search report prepared by the EPO or JPO \$840.00

International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$690.00

International preliminary examination fee (37 CFR 1.482) paid to USPTO but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$670.00

International preliminary examination fee (37 CFR 1.482) paid to USPTO and all claims satisfied provisions of PCT Article 33(1)-(4) \$ 96.00

ENTER APPROPRIATE BASIC FEE AMOUNT = \$840.00Surcharge of **\$130.00** for furnishing the oath or declaration later than ___ 20 ___ 30 months from the earliest claimed priority date (37 CFR 1.492(e)).

\$

CLAIMS

NUMBER FILED

NUMBER EXTRA

RATE

Total Claims

20 - 20 =

00

X \$18.00

\$

Independent Claims

06 - 03 =

03

X \$78.00

\$234.00

MULTIPLE DEPENDENT CLAIM(S) (if applicable)

+ \$260.00

\$

TOTAL OF ABOVE CALCULATIONS =

\$

Reduction of ½ for filing by small entity, if applicable. (Note 37 CFR 1.9, 1.27, 1.28)

\$537.00

SUBTOTAL =

\$537.00

Processing Fee of **\$130.00** for furnishing the English translation later than ___ 20 ___ 30 months from the earliest claimed priority date (37 CFR 1.492(f)).

\$

TOTAL NATIONAL FEE =

\$537.00

Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property.

\$

TOTAL FEES ENCLOSED =

\$537.00

Amount to be:
refunded

\$

charged

\$537.00

a. ___ A check in the amount of \$ ___ to cover the above fees is included.

b. ☒ Please charge my Deposit Account No. 50-0815 in the amount of \$ 537.00 to cover the above fees.c. ☒ The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 50-0815.**NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.**

SEND ALL CORRESPONDENCE TO

BOZICEVIC, FIELD & FRANCIS LLP
200 Middlefield Road, Suite 200
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(650) 327-3400 Telephone
(650) 327-3231 Facsimile

SIGNATURE

NAME: Carol L. Francis

36,513

REGISTRATION NUMBER

09/700354

529 Rec'd PCT/PTC 13 NOV 2000
Atty Dkt. No.: IRVN-007CIP2

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Typed or Printed Name	Matthew OTT5	Express Mail No.	EL 563 650 845 US
Signature	<i>Matthew Ott</i>	Date	November 13, 2000

<p align="center">PRELIMINARY AMENDMENT</p> <p>Address to: Commissioner for Patents Washington, D.C. 20231</p>	Attorney Docket	IRVN-007CIP2 (UC 96-367-4)
	First Named Inventor	Gatanaga, et al.
	Application Number	N/A
	Filing Date	Herewith
	Group Art Unit	N/A
	Examiner Name	N/A
	Title	Factors Affecting Tumor Necrosis factor Receptor Releasing Enzyme Activity

Sir:

Prior to the examination on the merits of the above-referenced application, please amend the application as follows.

AMENDMENTS

In The Claims:

Cancel claims 1-36 without prejudice.

Add the following new claims:

-- 37. (New) An isolated polypeptide having at least one of the following properties:

- a) the polypeptide comprises at least 10 consecutive amino acid residues encoded in any of SEQ ID NOs:8, 9, 1-3, 5-6, or 10; or
- b) the polypeptide comprises at least 15 consecutive amino acids that are at least 80% identical to a sequence encoded in any of SEQ ID NOs:8, 9, 1-3, 5-6, or 10.

38. (New) The polypeptide of claim 37, having at least one of the following properties:

a) the polypeptide comprises at least 10 consecutive amino acid residues contained in SEQ ID NOS: 147-149, 151, or 153-154;

b) the polypeptide comprises at least 15 consecutive amino acids that are at least 80% identical to a sequence contained in SEQ ID NOS: 147-149, 151, or 153-154.

39. (New) The polypeptide of claim 37, which when incubated with COS-1 cells expressing TNF receptor, promotes enzymatic cleavage and release of the receptor.

40. (New) The polypeptide of claim 37, which is immunogenic for an antibody specific for a modulator of TRRE activity.

41. (New) The polypeptide of claim 37, which either:

a) lacks a membrane spanning sequence; or

b) is produced by a process comprising recombinant expression in a host cell followed by purification of the polypeptide from medium in which the cell is cultured.

42. (New) A method of producing the polypeptide according to any of claim 37, comprising:

a) culturing host cells genetically altered to express a polynucleotide comprising an encoding sequence for the polypeptide; and subsequently

b) purifying the polypeptide from the cells.

43. (New) The method of claim 42, comprising harvesting culture medium, and purifying the polypeptide from the culture medium by a process comprising affinity chromatography.

44. (New) An isolated antibody specific for a polypeptide according to claim 37.

45. (New) A method for producing the antibody according to claim 44, comprising immunizing a mammal or contacting an immunocompetent cell or particle with a polypeptide according to claim 37.

46. (New) An assay method for determining altered TRRE activity in a cell or tissue sample, comprising:

a) contacting the sample with a polynucleotide under conditions that permit the polynucleotide to hybridize specifically with nucleic acid that encodes a modulator of TRRE activity, if present in the sample; and

b) determining polynucleotide that has hybridized as a result, as a measure of altered TRRE activity in the sample;

wherein the polynucleotide has at least one of the following properties:

i) the polynucleotide comprises a nucleotide sequence contained in SEQ ID NOs:1-10;

ii) the polynucleotide comprises a nucleotide sequence of at least 30 consecutive nucleotides contained in SEQ ID NOs:1-10;

iii) the polynucleotide comprises a nucleotide sequence of at least 50 consecutive nucleotides at least 90% identical to a sequence contained in SEQ ID NOs:1-10; or

iv) the polynucleotide is capable of hybridizing specifically to a nucleotide sequence contained in SEQ ID NOs:1-10 under stringent conditions.

47. (New) The assay method of claim 46, wherein the polynucleotide comprises a nucleotide sequence of at least 30 consecutive nucleotides contained in SEQ ID NOs:1-10.

48. (New) An assay method for determining altered expression of a modulator of TRRE activity in a cell or tissue sample, comprising:

a) contacting the sample with the antibody of claim 44 under conditions that permit the antibody to bind the modulator if present in the sample, thereby forming an antibody-antigen complex; and

b) determining any complex formed as a measure of altered expression of the modulator.

49. (New) A method for assessing a disease condition associated with altered TRRE activity in a subject, comprising determining altered expression of a TRRE modulator according to claim 46, and then correlating the extent of alteration with the disease condition.

50. (New) A method for decreasing signal transduction from a cytokine into a cell, comprising contacting the cell with a polypeptide having at least one of the following properties:

- a) the polypeptide comprises at least 10 consecutive amino acid residues encoded in any of SEQ ID NOs:1-10; or
- b) the polypeptide comprises at least 15 consecutive amino acids that are at least 80% identical to a sequence encoded in any of SEQ ID NOs:1-10.

51. (New) A method for increasing signal transduction from a cytokine into a cell, comprising contacting the cell with an antibody according to claim 44.

52. (New) The method of claim 50, wherein the cytokine is TNF.

53. (New) A method of screening polynucleotides for an ability to modulate TRRE activity, comprising:

- a) providing cells that express both TRRE and the TNF-receptor;
- b) genetically altering the cells with the polynucleotides to be screened;
- c) cloning the cells genetically altered; and
- d) identifying clones that enzymatically release the receptor at an altered rate.

54. (New) A pharmaceutical composition comprising a polynucleotide in a pharmaceutically compatible excipient, wherein the polynucleotide has at least one of the following properties:

- a) the polynucleotide comprises a nucleotide sequence contained in SEQ ID NOs:1-10;
- b) the polynucleotide comprises a nucleotide sequence of at least 30 consecutive nucleotides contained in SEQ ID NOs:1-10;
- c) the polynucleotide comprises a nucleotide sequence of at least 50 consecutive nucleotides at

least 90% identical to a sequence contained in SEQ ID NOs:1-10;

d) the polynucleotide is capable of hybridizing specifically to a nucleotide sequence contained in SEQ ID NOs:1-10 under stringent conditions; or

e) the polynucleotide comprises a nucleotide sequence that encodes at least 10 consecutive amino acids encoded in SEQ ID NOs:1-10.

55. (New) A pharmaceutical composition comprising a polypeptide in a pharmaceutically compatible excipient, wherein the polypeptide has at least one of the following properties:

a) the polypeptide comprises at least 10 consecutive amino acid residues encoded in any of SEQ ID NOs:1-10; or

b) the polypeptide comprises at least 15 consecutive amino acids that are at least 80% identical to a sequence encoded in any of SEQ ID NOs:1-10.

56. (New) A pharmaceutical composition comprising an antibody according to claim 44 in a pharmaceutically compatible excipient.--

REMARKS

Claims 37-56 are pending after entry of the amendments above.

Claims 1-36 are canceled without prejudice to renewal, without intent to abandon any subject matter therein, and without acquiescing to any rejection which may have been applied. Applicants expressly reserve the right to pursue the subject matter of the canceled claims in a continuing application.

Support for new claims 37-43 is found in, for example, claims 7-14 as originally filed.

Support for new claims 44-45 is found in, for example, claims 16 and 17 as originally filed.

Support for new claims 46-47 is found in, for example, claims 18, 3, 4, and 5 as originally filed.

Support for new claim 48 is found in, for example, claim 19 as originally filed.

Support for new claim 49 is found in, for example, claim 20 as originally filed.

Support for new claim 50 is found in, for example, claims 21, 9, and 10 as originally filed.

Support for new claims 51, 52 and 53 is found in, for example, claims 22-24 as originally filed.

Support for new claim 54 is found in, for example, claims 2-6 and 27 as originally filed, as well as in the specification at page 26, lines 3-28 and page 27, lines 16-30.

Support for new claim 55 is found in, for example, claims 29 and 7-14 as originally filed, as well as in the specification at page 25, line 26 to page 26, line 2 and page 27, lines 16-30.

Support for new claims 56 is found in, for example, claim 28 as originally filed as well as in the specification at page 21, lines 5-9.

No new matter is added.

CONCLUSION

Applicants respectfully submit that the claims are in form for allowance, early notice of which is requested. If, in the opinion of the Examiner a telephone conference would expedite the prosecution of the subject application, the Examiner is invited to call the undersigned at (650) 327-3400.

The Commissioner is hereby authorized to charge any fees under 37 C.F.R. §§ 1.16 and 1.17 which may be required by this paper, or to credit any overpayment, to Deposit Account No. 50-0815.

Respectfully submitted,
BOZICEVIC, FIELD & FRANCIS LLP

Date:

November 13, 2000

By:

Carol L. Francis
Carol L. Francis
Registration No. 36,513

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09/700354

529 Rec'd PCT/PTC 13 NOV 2000

Our Docket: FP-UC 3668

**PATENT COOPERATION TREATY
IN THE INTERNATIONAL BUREAU OF WIPO**

In re International Application)
No. PCT/US99/10793)
Applicant: The Regents of the)
University of California,)
et al.)
Filed: 14 May 1999)
Entitled: FACTORS AFFECTING TUMOR)
NECROSIS FACTOR RECEPTOR)
RELEASING ENZYME ACTIVITY)

International Bureau of WIPO
34, chemin des Colombettes
1221 Geneva 20
Switzerland

LETTER

Sir:

Pursuant to PCT Article 19(1), and responsive to the International Search Report dated December 3, 1999, Applicant respectfully requests that the following amendments and remarks be considered and that the Substitute page submitted herewith be entered for examination.

AMENDMENTS

Applicant respectfully requests replacement of original page 99 with the enclosed substitute page 99. The status of the claims is as follows:

Claims 1 to 32 are unchanged.

New claims 33 to 35 have been added, as follows:

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Filed: 14 May 1999
Docket: FP-UC 3668
Page 2

--33. The polynucleotide according to any of claims 1-5, wherein said nucleotide sequence is not contained in any of the sequences of the following GenBank Accession Nos: AJ003355, AA806165; AI002979; T33896; U52522; AA779203; C06247; AA707194; AA599596; 5453538; U13369; and J03528.

34. The polypeptide according to any of claims 7-10, the sequence of which is not completely encoded by a polynucleotide sequence contained in any of the sequences of the following GenBank Accession Nos: AJ003355, AA806165; AI002979; T33896; U52522; AA779203; C06247; AA707194; AA599596; 5453538; U13369; and J03528.

35. The polynucleotide according to claim 15, the sequence of which is not contained in any of the sequences of the following GenBank Accession Nos: AJ003355, AA806165; AI002979; T33896; U52522; AA779203; C06247; AA707194; AA599596; 5453538; U13369; and J03528.--

REMARKS

The GenBank Accession Numbers recited in new claims 33 to 35 relate to items cited in the International Search Report.

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Docket: FP-UC 3668
Page 3

CONCLUSION

The Examiner is invited to contact the undersigned agent or Cathryn Campbell if there are any questions relating to the subject application.

Respectfully submitted,

1/28/00
Date

Melanie K. Webster
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FACTORS AFFECTING TUMOR NECROSIS FACTOR RECEPTOR
RELEASING ENZYME ACTIVITY

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims the priority benefit of U.S. application 09/081,385,
5 filed May 14, 1998, pending. For purposes of prosecution in the U.S., the priority
application is hereby incorporated herein by reference in its entirety.

FIELD OF THE INVENTION

10 This invention relates generally to the field of signal transduction between
cells, via cytokines and their receptors. More specifically, it relates to enzymatic
activity that cleaves and releases the receptor for TNF found on the cell surface,
and the consequent biological effects. Certain embodiments of this invention are
compositions that affect such enzymatic activity, and may be included in
medicaments for disease treatment.

15 BACKGROUND OF THE INVENTION

Cytokines play a central role in the communication between cells.
Secretion of a cytokine from one cell in response to a stimulus can trigger an
adjacent cell to undergo an appropriate biological response — such as
20 stimulation, differentiation, or apoptosis. It is hypothesized that important
biological events can be influenced not only by affecting cytokine release from
the first cell, but also by binding to receptors on the second cell, which mediates
the subsequent response. The invention described in this patent application
provides new compounds for affecting signal transduction from tumor necrosis
25 factor.

The cytokine known as tumor necrosis factor (TNF or TNF- α) is
structurally related to lymphotoxin (LT or TNF- β). They have about 40 percent
amino acid sequence homology (Old, *Nature* 330:602-603, 1987). These
cytokines are released by macrophages, monocytes and natural killer cells and

play a role in inflammatory and immunological events. The two cytokines cause a broad spectrum of effects both in vitro and in vivo, including: (i) vascular thrombosis and tumor necrosis; (ii) inflammation; (iii) activation of macrophages and neutrophils; (iv) leukocytosis; (v) apoptosis; and (vi) shock. TNF has been associated with a variety of disease states including various forms of cancer, arthritis, psoriasis, endotoxic shock, sepsis, autoimmune diseases, infections, obesity, and cachexia. TNF appears to play a role in the three factors contributing to body weight control: intake, expenditure, and storage of energy (Rothwell, *Int. J. Obesity* 17:S98-S101, 1993). In septicemia, increased endotoxin concentrations appear to raise TNF levels (Beutler et al. *Science* 229:869-871, 1985).

Attempts have been made to alter the course of a disease by treating the patient with TNF inhibitors, with varying degrees of success. For example, the TNF inhibitor dexamabiol provided protection against TNF mediated effects following traumatic brain injury (Shohami et al. *J. Neuroimmun.* 72:169-77, 1997). Some improvement in Crohn's disease was afforded by treatment with anti-TNF antibodies (Neurath et al., *Eur. J. Immun.* 27:1743-50, 1997).

Human TNF and LT mediate their biological activities by binding specifically to two distinct glycoprotein plasma membrane receptors (55 kDa and 75 kDa in size, known as p55 and p75 TNF-R, respectively). The two receptors share 28 percent amino acid sequence homology in their extracellular domains, which are composed of four repeating cysteine-rich regions (Tartaglia and Goeddel, *Immunol. Today* 13:151-153, 1992). However, the receptors lack significant sequence homology in their intracellular domains, and mediate different intracellular responses to receptor activation. In accordance with the different activities of TNF and LT, most human cells express low levels of both TNF receptors: about 2,000 to 10,000 receptors per cell (Brockhaus et al., *Proc. Natl. Acad. Sci. USA* 87:3127-3131, 1990).

Expression of TNF receptors on both lymphoid and non-lymphoid cells can be influenced experimentally by many different agents, such as bacterial lipopolysaccharide (LPS), phorbol myristate acetate (PMA; a protein kinase C

activator), interleukin-1 (IL-1), interferon-gamma (IFN- γ) and IL-2 (Gatanaga et al. *Cell Immunol.* 138:1-10, 1991; Yui et al. *Placenta* 15:819-835, 1994). It has been shown that complexes of human TNF bound to its receptor are internalized from the cell membrane, and then the receptor is either degraded or recycled (Armitage, *Curr. Opin. Immunol.* 6:407-413, 1994). It has been proposed that

5 TNF receptor activity can be modulated using peptides that bind intracellularly to the receptor, or which bind to the ligand binding site, or that affect receptor shedding. See for example patent publications WO 95/31544, WO 95/33051, WO 96/01642, and EP 568 925.

10 TNF binding proteins (TNF-BP) have been identified at elevated levels in the serum and urine of febrile patients, patients with renal failure, and cancer patients, and even certain healthy individuals. Human brain and ovarian tumors produced high serum levels of TNF-BP. These molecules have been purified, characterized, and cloned (Gatanaga et al., *Lymphokine Res.* 9:225-229, 1990a; Gatanaga et al., *Proc. Natl. Acad. Sci USA* 87:8781-8784, 1990b). Human

15 TNF-BP consists of 30 kDa and 40 kDa proteins which are identical to the N-terminal extracellular domains of p55 and p75 TNF receptors, respectively (US Patent No. 5,395,760; EP 418,014). Such proteins have been suggested for use in treating endotoxic shock. Mohler et al. *J. Immunol.* 151:1548-1561, 1993

20 There are several mechanisms possible for the production of secreted proteins resembling membrane bound receptors. One involves translation from alternatively spliced mRNAs lacking transmembrane and cytoplasmic regions. Another involves proteolytic cleavage of the intact membrane receptors, followed by shedding of the cleaved receptor from the cell. The soluble form of p55 and

25 p75 TNF-R do not appear to be generated from mRNA splicing, since only full length receptor mRNA has been detected in human cells *in vitro* (Gatanaga et al., 1991). Carboxyl-terminal sequencing and mutation studies on human p55 TNF-R indicates that a cleavage site may exist between residues Asn 172 and Val 173 (Gullberg et al. *Eur. J. Cell. Biol.* 58:307-312, 1992).

30 There are reports that a specific metalloprotease inhibitor, TNF- α protease inhibitor (TAPI) blocks the shedding of soluble p75 and p55 TNF-R (Crowe et al.

J. Exp. Med. 181:1205-1210, 1995; Mullberg et al. *J. Immunol.* 155:5198-5205, 1995). The processing of pro-TNF on the cell membrane to release the TNF ligand appears to be dependent on a matrix metalloprotease like enzyme (Gearing et al. *Nature* 370:555-557, 1994). This is a family of structurally related matrix-degrading enzymes that play a major role in tissue remodeling and repair associated with development and inflammation (Birkedal-Hansen et al. *Crit. Rev. Oral Biol. Med.* 4:197-250, 1993). The enzymes have Zn^{2+} in their catalytic domains, and Ca^{2+} stabilizes their tertiary structure significantly.

In European patent application EP 657536A1, Wallach et al. suggest that it would be possible to obtain an enzyme that cleaves the 55,000 kDa TNF receptor by finding a mutated form of the receptor that is not cleaved by the enzyme, but still binds to it. The only proposed source for the enzyme is a detergent extract of membranes for cells that appear to have the protease activity. If it were possible to obtain an enzyme according to this scheme, then the enzyme would presumably comprise a membrane spanning region. The patent application does not describe any protease that was actually obtained.

In a previous patent application in the present series (International Patent Publication WO 9820140), methods are described for obtaining an isolated enzyme that cleaves both the p55 and p75 TNF-R from cell surfaces. A convenient source is the culture medium of cells that have been stimulated with phorbol myristate acetate (PMA). The enzyme activity was given the name TRRE (TNF receptor releasing enzyme). In other studies, TRRE was released immediately upon PMA stimulation, indicating that it is presynthesized in an inactive form to be rapidly converted to the active form upon stimulation. Evidence for direct cleavage of TNF-R is that the shedding begins very quickly (~5 min) with maximal shedding within 30 min. TRRE is specific for the TNF-R, and does not cleave IL-1 receptors, CD30, ICAM-1 or CD11b. TRRE activity is enhanced by adding Ca^{++} or Zn^{++} , and inhibited by EDTA and phenantroline.

Given the involvement of TNF in a variety of pathological conditions, it is desirable to obtain a variety of factors that would allow receptor shedding to be

modulated, thereby controlling the signal transduction from TNF at a disease site.

SUMMARY OF THE INVENTION

5 This disclosure provides new compounds that promote enzymatic cleavage and release of TNF receptors from the cell surface. Nine new DNA clones have been selected after repeat screening in an assay that tests the ability to enhance receptor release. The polynucleotide sequences of this invention and the proteins encoded by them have potential as diagnostic aids,
10 and therapeutic compounds that can be used to adjust TNF signal transduction in a beneficial way.

One embodiment of the invention is an isolated polynucleotide comprising a nucleotide sequence with the following properties: a) the sequence is expressed at the mRNA level in Jurkat T cells; b) when COS-1 cells expressing
15 TNF-receptor are genetically transformed to express the sequence, the cells have increased enzymatic activity for cleaving and releasing the receptor. If a polynucleotide sequence is expressed in Jurkat cells, then it can be found in the Jurkat cell expression library deposited with the ATCC (Accession No. TIB-152). It is recognized that the polynucleotide can be obtained from other cell lines, or
20 produced by recombinant techniques.

Included are polynucleotides in which the nucleotide sequence is contained in any of SEQ. ID NOS:1-10. Also embodied are polynucleotides comprising at least 30 and preferably more consecutive nucleotides in said nucleotide sequence, or at least 50 consecutive nucleotides that are homologous
25 to said sequence at a significant level, preferably at the 90% level or more. Also included antisense and ribozyme polynucleotides that inhibit the expression of a TRRE modulator.

Another embodiment of the invention is isolated polypeptides comprising an amino acid sequence encoded by a polynucleotide of this invention. Non-
30 limiting examples are sequences shown in SEQ. ID NOS: 147-158. Fragments

and fusion proteins are included in this invention, and preferably comprise at least 10 consecutive residues encoded by a polynucleotide of this invention, or at least 15 consecutive amino acids that are homologous at a significant level, preferably at least 80%. Preferred polypeptides promote cleavage and release of TNF receptors from the cell surface, especially COS-1 cells genetically transformed to express TNF receptor. The polypeptides may or may not have a membrane spanning domain, and may optionally be produced by a process that involves secretion from a cell. Included are species homologs with the desired activity, and artificial mutants with additional beneficial properties.

Another embodiment of this invention is an antibody specific for a polypeptide of this invention. Preferred are antibodies that bind a TRRE modulator protein, but not other substances found in human tissue samples in comparable amounts.

Another embodiment of the invention is an assay method of determining altered TRRE activity in a cell or tissue sample, using a polynucleotide or antibody of this invention to detect the presence or absence of the corresponding TRRE modulator. The assay method can optionally be used for the diagnosis or evaluation of a clinical condition relating to abnormal TNF levels or TNF signal transduction.

Another embodiment of the invention is a method for increasing or decreasing signal transduction from a cytokine into a cell (including but not limited to TNF), comprising contacting the cell with a polynucleotide, polypeptide, or antibody of this invention.

A further embodiment of the invention is a method for screening polynucleotides for an ability to modulate TRRE activity. The method involves providing cells that express both TRRE and the TNF-receptor; genetically altering the cells with the polynucleotides to be screened; cloning the cells; and identifying clones with the desired activity.

Yet another embodiment of the invention is a method for screening substances for an ability to affect TRRE activity. This typically involves incubating cells expressing TNF receptor with a TRRE modulator of this

invention in the presence or absence of the test substance; and measuring the effect on shedding of the TNF receptor .

The products of this invention can be used in the preparation of a medicament for treatment of the human or animal body. The medicament contains a clinically effective amount for treatment of a disease such as heart failure, cachexia, inflammation, endotoxic shock, arthritis, multiple sclerosis, sepsis, and cancer. These compositions can be used for administration to a subject suspected of having or being at risk for the disease, optionally in combination with other forms of treatment appropriate for their condition.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is a schematic representation of plasmid pCDTR2. This plasmid expresses p75 TNF-R, the ~75 kDa form of the TNF receptor. PCMV stands for cytomegalovirus; BGHpA stands for bovine growth hormone polyadenylation signal.

Figure 2 is a line depicting the levels of p75 TNF-R detected on COS-1 cells genetically altered to express the receptor. Results from the transformed cells, designated C75R (●, upward swooping line) is compared with that from the parental COS-1 cells (■, baseline). The receptor number was calculated by Scatchard analysis (inset).

Figure 3 is a survival graph, showing that TRRE decreases mortality in mice challenged with lipopolysaccharide (LPS) to induce septic peritonitis. (♦) LPS alone; (■) LPS plus control buffer; (●) LPS plus TRRE (2,000 U); (▲) LPS plus TRRE (4,000 U).

Figure 4 is a half-tone reproduction of a bar graph, showing the effect of 9 new clones on TRRE activity on C75R cells (COS-1 cells transfected to express the TNF-receptor. Each of the 9 clones increases TRRE activity by over 2-fold.

Figure 5 is a survival graph, showing the ability of 4 new expressed to save mice challenged with LPS. (♦) saline; (■) BSA; (Δ) Mey-3 (100 μg); (X) Mey-3 (10 μg); (*) Mey-5 (10 μg); (●) Mey-8 (10 μg).

DETAILED DESCRIPTION OF THE INVENTION

It has been discovered that certain cells involved in the TNF transduction pathway express enzymatic activity that causes TNF receptors to be shed from the cell surface. Enzymatic activity for cleaving and releasing TNF receptors has been given the designation TRRE. Phorbol myristate acetate induces release of TRRE from cells into the culture medium. An exemplary TRRE protein had been purified from the supernatant of TNF-1 cells (Example 2). The protease bears certain hallmarks of the metalloprotease family, and is released rapidly from the cell upon activation.

In order to elucidate the nature of this protein, functional cloning was performed. Jurkat cells were selected as being a good source of TRRE. The cDNA from a Jurkat library was expressed, and cell supernatant was tested for an ability to release TNF receptors from cell surfaces. Cloning and testing of the expression product was conducted through several cycles, and nine clones were obtained that more than doubled TRRE activity in the assay (Figure 4). At the DNA level, all 9 clones had different sequences.

Protein expression products from the clones have been tested in a lipopolysaccharide animal model for sepsis. Protein from three different clones successfully rescued animals from a lethal dose of LPS (Figure 5). This points to an important role for these molecules in the management of pathological conditions mediated by TNF.

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The number of new TRRE promoting clones obtained from the expression library was surprising. The substrate specificity of the TRRE isolated in Example 2 distinguishes the 75 kDa and 55 kDa TNF receptors from other cytokine receptors and cell surface proteins. There was little reason beforehand to suspect that cells might have nine different proteases for the TNF receptor. It is possible that one of the clones encodes the TRRE isolated in Example 2, or a related protein. It is possible that some of the other clones have proteolytic activity to cleave TNF receptors at the same site, or at another site that causes release of the soluble form from the cell. It is a hypothesis of this disclosure that some of the clones may not have proteolytic activity themselves, but play a role in promoting TRRE activity in a secondary fashion.

This possibility is consistent with the observations made, because there is an endogenous level of TRRE activity in the cells used in the assay. The cleavage assay involves monitoring TNF receptor release from C75 cells, which are COS-1 cells genetically altered to express p75 TNF-R. The standard assay is conducted by contacting the transformed cells with a fluid believed to contain TRRE. The level of endogenous TRRE activity is evident from the rate of spontaneous release of the receptor even when no exogenous TRRE is added (about 200 units). Accordingly, accessory proteins that promote TRRE activity would increase the activity measured in the assay. Many mechanisms of promotion are possible, including proteins that activate a zymogen form of TRRE, proteins that free TRRE from other cell surface components, or proteins that stimulate secretion of TRRE from inside the cell. It is not necessary to understand the mechanism in order to use the products of this invention in most of the embodiments described.

It is anticipated that several of the clones will have activity not just for promoting TNF receptor cleavage, but also having an effect on other surface proteins. To the extent that cleavage sequences or accessory proteins are shared between different receptors, certain clones would promote phenotypic change (such as receptor release) for the family of related substrates.

[illegible][illegible][illegible][illegible][illegible][illegible]

cells. Those that increase TRRE activity may be referred to as TRRE promoters, and those that decrease TRRE activity may be referred to as TRRE inhibitors. TRRE promoters include compounds that have proteolytic activity for TNF-R, and compounds that augment the activity of TNF-R proteases. The nine
5 polynucleotide clones described in Example 5, and their protein products, are exemplary TRRE promoters. Inhibitors of TRRE activity can be obtained using the screening assays described below.

The term "polynucleotide" refers to a polymeric form of nucleotides of any length, either deoxyribonucleotides or ribonucleotides, or analogs thereof.
10 Polynucleotides may have any three-dimensional structure, and may perform any function, known or unknown. The following are non-limiting examples of polynucleotides: a gene or gene fragment, exons, introns, (mRNA), ribozymes, cDNA, recombinant polynucleotides, branched polynucleotides, plasmids, vectors, nucleic acid probes, and primers. A polynucleotide may comprise
15 modified nucleotides, such as methylated nucleotides and nucleotide analogs. If present, modifications to the nucleotide structure may be imparted before or after assembly of the polymer. The term polynucleotide refers interchangeably to double-and single-stranded molecules. Unless otherwise specified or required, any embodiment of the invention described herein that is a polynucleotide
20 encompasses both the double-stranded form, and each of two complementary single-stranded forms known or predicted to make up the double-stranded form

"Hybridization" refers to a reaction in which one or more polynucleotides react to form a complex that is stabilized via hydrogen bonding between the bases of the nucleotide residues. Hybridization reactions can be performed
25 under conditions of different "stringency". Relevant conditions include temperature, ionic strength, and the presence of additional solutes in the reaction mixture such as formamide. Conditions of increasing stringency are 30°C. in 10X SSC (0.15M NaCl, 15 mM citrate buffer); 40°C. in 6X SSC; 50°C. in 6X SSC 60°C. in 6X SSC, or at about 40°C. in 0.5X SSC, or at about 30°C. in 6X.
30 SSC containing 50% formamide. SDS and a source of fragmented DNA (such as salmon sperm) are typically also present during hybridization. Higher

stringency requires higher minimum complementarity between hybridizing elements for a stable hybridization complex to form. See "Molecular Cloning: A Laboratory Manual", Second Edition (Sambrook, Fritsch & Maniatis, 1989).

It is understood that purine and pyrimidine nitrogenous bases with similar
5 structures can be functionally equivalent in terms of Watson-Crick base-pairing; and the inter-substitution of like nitrogenous bases, particularly uracil and thymine, or the modification of nitrogenous bases, such as by methylation, does not constitute a material substitution.

The percentage of sequence identity for polynucleotides or polypeptides is
10 calculated by aligning the sequences being compared, and then counting the number of shared residues at each aligned position. No penalty is imposed for the presence of insertions or deletions, but are permitted only where required to accommodate an obviously increased number of amino acid residues in one of the sequences being aligned. When one of the sequences being compared is
15 indicated as being "consecutive", then no gaps are permitted in that sequence during the comparison. The percentage identity is given in terms of residues in the test sequence that are identical to residues in the comparison or reference sequence.

As used herein, "expression" of a polynucleotide refers to the production
20 of an RNA transcript. Subsequent translation into protein or other effector compounds may also occur, but is not required unless specified.

"Genetic alteration" refers to a process wherein a genetic element is introduced into a cell other than by mitosis or meiosis. The element may be heterologous to the cell, or it may be an additional copy or improved version of
25 an element already present in the cell. Genetic alternation may be effected, for example, by transducing a cell with a recombinant plasmid or other polynucleotide through any process known in the art, such as electroporation, calcium phosphate precipitation, or contacting with a polynucleotide-liposome complex. Genetic alteration may also be effected, for example, by transduction
30 or infection with a DNA or RNA virus or viral vector. It is preferable that the

genetic alteration is inheritable by progeny of the cell, but this is not generally required unless specified.

The terms "polypeptide", "peptide" and "protein" are used interchangeably herein to refer to polymers of amino acids of any length. The polymer may be linear or branched, it may comprise modified amino acids, and it may be interrupted by non-amino acids. The terms also encompass an amino acid polymer that has been modified; for example, disulfide bond formation, glycosylation, lipidation, acetylation, phosphorylation, or any other manipulation, such as conjugation with a labeling component.

A "fusion polypeptide" is a polypeptide comprising regions in a different position in the sequence than occurs in nature. The regions can normally exist in separate proteins and are brought together in the fusion polypeptide; they can normally exist in the same protein but are placed in a new arrangement in the fusion polypeptide; or they can be synthetically arranged. A "functionally equivalent fragment" of a polypeptide varies from the native sequence by addition, deletion, or substitution of amino acid residues, or any combination thereof, while preserving a functional property of the fragment relevant to the context in which it is being used. Fusion peptides and functionally equivalent fragments are included in the definition of polypeptides used in this disclosure.

It is understood that the folding and the biological function of proteins can accommodate insertions, deletions, and substitutions in the amino acid sequence. Some amino acid substitutions are more easily tolerated. For example, substitution of an amino acid with hydrophobic side chains, aromatic side chains, polar side chains, side chains with a positive or negative charge, or side chains comprising two or fewer carbon atoms, by another amino acid with a side chain of like properties can occur without disturbing the essential identity of the two sequences. Methods for determining homologous regions and scoring the degree of homology are described in Altschul et al. *Bull. Math. Bio.* 48:603-616, 1986; and Henikoff et al. *Proc. Natl. Acad. Sci. USA* 89:10915-10919, 1992.

Substitutions that preserve the functionality of the polypeptide, or confer a new

and beneficial property (such as enhanced activity, stability, or decreased immunogenicity) are especially preferred.

An "antibody" (interchangeably used in plural form) is an immunoglobulin molecule capable of specific binding to a target, such as a polypeptide, through at least one antigen recognition site, located in the variable region of the immunoglobulin molecule. As used herein, the term encompasses not only intact antibodies, but also antibody equivalents that include at least one antigen combining site of the desired specificity. These include but are not limited to enzymatic or recombinantly produced fragments antibody, fusion proteins, humanized antibodies, single chain variable regions, diabodies, and antibody chains that undergo antigen-induced assembly.

An "isolated" polynucleotide, polypeptide, protein, antibody, or other substance refers to a preparation of the substance devoid of at least some of the other components that may also be present where the substance or a similar substance naturally occurs or is initially obtained from. Thus, for example, an isolated substance may be prepared by using a purification technique to enrich it from a source mixture. Enrichment can be measured on an absolute basis, such as weight per volume of solution, or it can be measured in relation to a second, potentially interfering substance present in the source mixture. Increasing enrichments of the embodiments of this invention are increasingly more preferred. Thus, for example, a 2-fold enrichment is preferred, 10-fold enrichment is more preferred, 100-fold enrichment is more preferred, 1000-fold enrichment is even more preferred. A substance can also be provided in an isolated state by a process of artificial assembly, such as by chemical synthesis or recombinant expression.

A "host cell" is a cell which has been genetically altered, or is capable of being transformed, by administration of an exogenous polynucleotide.

The term "clinical sample" encompasses a variety of sample types obtained from a subject and useful in an in vitro procedure, such as a diagnostic test. The definition encompasses solid tissue samples obtained as a surgical removal, a pathology specimen, or a biopsy specimen, cells obtained from a

clinical subject or their progeny obtained from culture, liquid samples such as blood, serum, plasma, spinal fluid, and urine, and any fractions or extracts of such samples that contain a potential indication of the disease.

Unless otherwise indicated, the practice of the invention will employ
5 conventional techniques of molecular biology, microbiology, recombinant DNA, and immunology, within the skill of the art. Such techniques are explained in the standard literature, such as: "Molecular Cloning: A Laboratory Manual", Second Edition (Sambrook, Fritsch & Maniatis, 1989), "Oligonucleotide Synthesis" (M. J. Gait, ed., 1984), "Animal Cell Culture" (R. I. Freshney, ed., 1987); the series
10 "Methods in Enzymology" (Academic Press, Inc.); "Handbook of Experimental Immunology" (D. M. Weir & C. C. Blackwell, Eds.), "Gene Transfer Vectors for Mammalian Cells" (J. M. Miller & M. P. Calos, eds., 1987), "Current Protocols in Molecular Biology" (F. M. Ausubel et al., eds., 1987); and "Current Protocols in Immunology" (J. E. Coligan et al., eds., 1991). The reader may also choose to
15 refer to a previous patent application relating to TRRE, International Patent Application WO 98020140.

For purposes of prosecution in the U.S., and in other jurisdictions where allowed, all patents, patent applications, articles and publications indicated anywhere in this disclosure are hereby incorporated herein by reference in their
20 entirety.

Polynucleotides

Polynucleotides of this invention can be prepared by any suitable technique in the art. Using the data provided in this disclosure, sequences of
25 less than ~50 base pairs are conveniently prepared by chemical synthesis, either through a commercial service or by a known synthetic method, such as the triester method or the phosphite method. A preferred method is solid phase synthesis using mononucleoside phosphoramidite coupling units (Hirose et al., *Tetra. Lett.* 19:2449-2452, 1978; U.S. Patent No. 4,415,732).

30 For use in antisense therapy, polynucleotides can be prepared by chemistry that produce more stable in pharmaceutical preparations. Non-limiting

examples include thiol-derivatized nucleosides (U.S. Patent 5,578,718), and oligonucleotides with modified backbones (U.S. Patent Nos. 5,541,307 and 5,378,825).

Polynucleotides of this invention can also be obtained by PCR
5 amplification of a template with the desired sequence. Oligonucleotide primers spanning the desired sequence are annealed to the template, elongated by a DNA polymerase, and then melted at higher temperature so that the template and elongated oligonucleotides dissociate. The cycle is repeated until the desired amount of amplified polynucleotide is obtained (U.S. Patent Nos.
10 4,683,195 and 4,683,202). Suitable templates include the Jurkat T cell library and other human or animal expression libraries that contain TRRE modulator encoding sequences. The Jurkat T cell library is available from the American Type Culture Collection, 10801 University Blvd., Manassas VA 20110, U.S.A. (ATCC #TIB-152). Mutations and other adaptations can be performed during
15 amplification by designing suitable primers, or can be incorporated afterwards by genetic splicing.

Production scale amounts of large polynucleotides are most conveniently obtained by inserting the desired sequence into a suitable cloning vector and reproducing the clone. Techniques for nucleotide cloning are given in Sambrook,
20 Fritsch & Maniatis (supra) and in U.S. Patent No. 5,552,524. Exemplary cloning and expression methods are illustrated in Example 6.

Preferred polynucleotide sequences are 50%, 70%, 80% , 90%, or 100% identical to one of the sequences exemplified in this disclosure; in order of increasing preference. The length of consecutive residues in the identical or
25 homologous sequence compared with the exemplary sequence can be about 15, 30, 50, 75, 100, 200 or 500 residues in order of increasing preference, up to the length of the entire clone. Nucleotide changes that cause a conservative substitution or retain the function of the encoded polypeptide (in terms of hybridization properties or what is encoded) are especially preferred
30 substitutions.

The polynucleotides of this can be used to measure altered TRRE activity in a cell or tissue sample. This involves contacting the sample with the polynucleotide under conditions that permit the polynucleotide to hybridize specifically with nucleic acid that encodes a modulator of TRRE activity, if present in the sample, and determining polynucleotide that has hybridized as a result of step a). Specificity of the test can be provided in one of several ways. One method involves the use of a specific probe — a polynucleotide of this invention with a sequence long enough and of sufficient identity to the sequence being detected, so that it binds the target and not other nucleic acid that might be present in the sample. The probe is typically labeled (either directly or through a secondary reagent) so that it can be subsequently detected. Suitable labels include ^{32}P and ^{33}P , chemiluminescent and fluorescent reagents. After the hybridization reaction, unreacted probe is washed away so that the amount of hybridized probe can be determined. Signal can be amplified using branched probes (U.S. Patent No. 5,124,246). In another method, the polynucleotide is a primer for a PCR reaction. Specificity is provided by the ability of the paired probes to amplify the sequence of interest. After a suitable number of PCR cycles, the amount of amplification product present correlates with the amount of target sequence originally present in the sample.

Such tests are useful both in research, and in the diagnosis or assessment of a disease condition. For example, TNF activity plays a role in eliminating tumor cells (Example 4), and a cancer may evade the elimination process by activating TRRE activity in the diseased tissue. Hence, under some conditions, high expression of TRRE modulators may correlate with progression of cancer. Diagnostic tests are also of use in monitoring therapy, such as when gene therapy is performed to increase TRRE activity.

Polynucleotides of this invention can also be used for production of polypeptides and the preparation of medicaments, as explained below.

Polypeptides

Short polypeptides of this invention can be prepared by solid-phase chemical synthesis. The principles of solid phase chemical synthesis can be found in Dugas & Penney, Bioorganic Chemistry, Springer-Verlag NY pp 54-92 (1981), and U.S. Patent No. 4,493,795. Automated solid-phase peptide synthesis can be performed using devices such as a PE-Applied Biosystems 430A peptide synthesizer (commercially available from Applied Biosystems, Foster City CA).

Longer polypeptides are conveniently obtained by expression cloning. A polynucleotide encoding the desired polypeptide is operably linked to control elements for transcription and translation, and then transfected into a suitable host cell. Expression may be effected in procaryotes such as *E. coli* (ATCC Accession No. 31446 or 27325), eukaryotic microorganisms such as the yeast *Saccharomyces cerevisiae*, or higher eukaryotes, such as insect or mammalian cells. A number of expression systems are described in U.S. Patent No. 5,552,524. Expression cloning is available from such commercial services as Lark Technologies, Houston TX. The production of protein from 4 exemplary clones of this invention in insect cells is illustrated in Example 6. The protein is purified from the producing host cell by standard methods in protein chemistry, such as affinity chromatography and HPLC. Expression products are optionally produced with a sequence tag to facilitate affinity purification, which can subsequently be removed.

Preferred sequences are 40%, 60%, 80% , 90%, or 100% identical to one of the sequences exemplified in this disclosure; in order of increasing preference. The length of the identical or homologous sequence compared with the native human polynucleotide can be about 7, 10, 15, 20, 30, 50 or 100 residues in order of increasing preference, up to the length of the entire encoding region.

Polypeptides can be tested for an ability to modulate TRRE in a TNF-R cleavage assay. The polypeptide is contacted with the receptor (preferably expressed on the surface of a cell, such as a C75 cell), and the ability of the polypeptide to increase or decrease receptor cleavage and release is

determined. Cleavage of TNF-R by exemplary polypeptides of this invention is illustrated in Example 7.

Polypeptides of this invention can be used as immunogens for raising antibody. Large proteins will raise a cocktail of antibodies, while short peptide fragments will raise antibodies against small region of the intact protein. Antibody clones can be mapped for protein binding site by producing short overlapping peptides of about 10 amino acids in length. Overlapping peptides can be prepared on a nylon membrane support by standard F-Moc chemistry, using a SPOTS™ kit from Genosys according to manufacturer's directions.

Polypeptides of this invention can also be used to affect TNF signal transduction, as explained below.

Antibodies

Polyclonal antibodies can be prepared by injecting a vertebrate with a polypeptide of this invention in an immunogenic form. Immunogenicity of a polypeptide can be enhanced by linking to a carrier such as KLH, or combining with an adjuvant, such as Freund's adjuvant. Typically, a priming injection is followed by a booster injection is after about 4 weeks, and antiserum is harvested a week later. Unwanted activity cross-reacting with other antigens, if present, can be removed, for example, by running the preparation over adsorbants made of those antigens attached to a solid phase, and collecting the unbound fraction. If desired, the specific antibody activity can be further purified by a combination of techniques, which may include protein, A chromatography, ammonium sulfate precipitation, ion exchange chromatography, HPLC, and immunoaffinity chromatography using the immunizing polypeptide coupled to a solid support. Antibody fragments and other derivatives can be prepared by standard immunochemical methods, such as subjecting the antibody to cleavage with enzymes such as papain or pepsin.

Production of monoclonal antibodies is described in such standard references as Harrow & Lane (1988), U.S. Patent Nos. 4,491,632, 4,472,500 and 4,444,887, and *Methods in Enzymology* 73B:3 (1981). Briefly, a mammal is

immunized, and antibody-producing cells (usually splenocytes) are harvested. Cells are immortalized by fusion with a non-producing myeloma, transfecting with Epstein Barr Virus, or transforming with oncogenic DNA. The treated cells are cloned and cultured, and the clones are selected that produce antibody of the
5 desired specificity.

Other methods of obtaining specific antibody molecules (optimally in the form of single-chain variable regions) involve contacting a library of immunocompetent cells or viral particles with the target antigen, and growing out positively selected clones. Immunocompetent phage can be constructed to
10 express immunoglobulin variable region segments on their surface. See Marks et al., *New Eng. J. Med.* 335:730, 1996, International Patent Applications WO 9413804, WO 9201047, WO 90 02809, and McGuinness et al., *Nature Biotechnol.* 14:1449, 1996.

The antibodies of this invention are can be used in immunoassays for
15 TRRE modulators. General techniques of immunoassay can be found in "The Immunoassay Handbook", Stockton Press NY, 1994; and "Methods of Immunological Analysis", Weinheim: VCH Verlags gesellschaft mbH, 1993). The antibody is combined with a test sample under conditions where the antibody will bind specifically to any modulator that might be present, but not any other
20 proteins liable to be in the sample. The complex formed can be measured in situ (U.S. Patent Nos. 4,208,479 and 4,708,929), or by physically separating it from unreacted reagents (U.S. Patent No. 3,646,346). Separation assays typically involve labeled TRRE reagent (competition assay), or labeled antibody (sandwich assay) to facilitate detection and quantitation of the complex. Suitable
25 labels are radioisotopes such as ^{125}I , enzymes such as β -galactosidase, and fluorescent labels such as fluorescein. Antibodies of this invention can also be used to detect TRRE modulators in fixed tissue sections by immunohistology. The antibody is contacted with the tissue, unreacted antibody is washed away, and then bound antibody is detected — typically using a labeled anti-
30 immunoglobulin reagent. Immunohistology will show not only whether the modulator is present, but where it is located in the tissue.

Detection of TRRE modulators is of interest for research purposes, and for clinical use. As indicated earlier, high expression of TRRE modulators may correlate with progression of cancer. Diagnostic tests are also of use in monitoring TRRE modulators that are administered in the course of therapy.

Antibodies of this invention can also be used for preparation of medicaments. Antibodies with therapeutic potential include those that affect TRRE activity — either by promoting clearance of a TRRE modulator, or by blocking its physiological action. Antibodies can be screened for desirable activity according to assays described in the next section.

Screening assays

This invention provides a number of screening methods for selecting and developing products that modulate TRRE, and thus affect TNF signal transduction.

One screening method is for polynucleotides that have an ability to modulate TRRE activity. To do this screening, cells are obtained that express both TRRE and the TNF receptor. Suitable cell lines can be constructed from any cell that expresses a level of functional TRRE activity. These cells are identifiable by testing culture supernatant for an ability to release membrane-bound TNF-R. The level of TRRE expression should be moderate, so that an increase in activity can be detected. The cells can then be genetically altered to express either p55 or p75 TNF-R, illustrated in Example 1. Exemplary is the C75R line: COS-1 cells genetically altered to express the 75 kDa form of the TNF-R. Release of TNF-R from the cell can be measured either by testing residual binding of labeled TNF ligand to the cell, or by immunoassay of the supernatant for released receptor (Example 1).

The screening assay is conducted by contacting the cells expressing TRRE and TNF-R with the polynucleotides to be screened. The effect of the polynucleotide on the enzymatic release of TNF-R from the cell is determined, and polynucleotides with desirable activity (either promoting or inhibiting TRRE activity) are selected. In a variation of this method, cells expressing TRRE

activity but not TNF-R (such as untransfected COS-1 cells) are contacted with the test polynucleotide. Then the culture medium is collected, and used to assay for TRRE activity using a second cell expressing TNF-R (such as C75 cells).

5 This type of screening assay is useful for the selection of polynucleotides from an expression library believed to contain encoding sequences for TRRE modulators. The Jurkat cell expression library (ATCC Accession No. TIB-152) is exemplary. Other cells from which suitable libraries can be constructed are those known to express high levels of TRRE, especially after PMA stimulation, such as THP-1, U-937, HL-60, ME-180, MRC-5, Raji, K-562, and normal human
10 monocytes. The screening involves expressing DNA from the library in the selected cell line being used for screening. Wells with the desired activity are selected, and the DNA is recovered, optionally after replication or cloning of the cells. Repeat cycles of functional screening and selection can lead to identification of new polynucleotide clones that promote or inhibit TRRE activity.
15 This is illustrated below in Example 5. Further experiments can be performed on the selected polynucleotides to determine it modulates TRRE activity inside the cell, or through the action of a protein product. A long open reading frame suggests a role for a protein product, and examination of the amino acid sequence for a signal peptide and a membrane spanning region can help
20 determine whether the protein is secreted from the cell or expressed in the surface membrane.

This type of screening is also useful for further development of the polynucleotides of this invention. For example, expression constructs can be developed that encode functional peptide fragments, fusion proteins, and other
25 variants. The minimum size of polynucleotide sequence that still encodes TRRE modulation activity can be determined by removing part of the sequence and then using the screening assay to determine whether the activity is still present. Mutated and extended sequences can be tested in the same way.

This type of screening assay is also useful for developing compounds that
30 affect TRRE activity by interfering with mRNA that encode a TRRE modulator. Of particular interest are ribozymes and antisense oligonucleotides. Ribozymes

are endoribonucleases that catalyze cleavage of RNA at a specific site. They comprise a polynucleotide sequence that is complementary to the cleavage site on the target, and additional sequence that provide the tertiary structure to effect the cleavage. Construction of ribozymes is described in U.S. Patent Nos. 4,987,071 and 5,591,610. Antisense oligonucleotides that bind mRNA comprise a short sequence complementary to the mRNA (typically 8-25 bases in length). Preferred chemistry for constructing antisense oligonucleotides is outlined in an earlier section. Specificity is provided both by the complementary sequence, and by features of the chemical structure. Antisense molecules that inhibit expression of cell surface receptors are described in U.S. Patent Nos. 5,135,917 and 5,789,573. Screening involves contacting the cell expressing TRRE activity and TNF-R with the compound and determining the effect on receptor release. Ribozymes and antisense molecules effective in altering expression of a TRRE promoter would decrease TNF-R release. Ribozymes and antisense molecules effective in altering expression of a TRRE inhibitor would increase TNF-R release.

Another screening method described in this disclosure is for testing the ability of polypeptides to modulate TRRE activity (Example 7). Cells expressing both TNF-R and a moderate level of TRRE activity are contacted with the test polypeptides, and the rate of receptor release is compared with the rate of spontaneous release. An increased rate of release indicates that the polypeptide is a TRRE promoter, while a decreased rate indicates that the polypeptide is a TRRE inhibitor. This assay can be used to test the activity of new polypeptides, and develop variants of polypeptides already known to modulate TRRE. The minimum size of polypeptide sequence that still encodes TRRE modulation activity can be determined by making a smaller fragment of the polypeptide and then using the screening assay to determine whether the activity is still present. Mutated and extended sequences can be tested in the same way.

Another screening method embodied in this invention is a method for screening substances that interfere with the action of a TRRE modulator at the protein level. The method involves incubating cells expressing TNF receptor

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Medicaments and their use

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surface of cells, which would decrease signal transduction from TNF. Conversely, products that inhibit TRRE activity prevent cleavage of TNF receptors, increasing signal transduction.

The ability to affect TNF signal transduction is of considerable interest in the management of clinical conditions in which TNF signaling contributes to the pathology of the condition. Such conditions include:

- Heart failure. IL-1 β and TNF are believed to be central mediators for perpetuating the inflammatory process, recruiting and activating inflammatory cells. The inflammation depress cardiac function in congestive heart failure, transplant rejection, myocarditis, sepsis, and burn shock.
- Cachexia. The general weight loss and wasting occurring in the course of chronic diseases, such as cancer. TNF is believed to affect appetite, energy expenditure, and metabolic rate.
- Crohn's disease. The inflammatory process mediated by TNF leads to thickening of the intestinal wall, ensuing from lymphedema and lymphocytic infiltration.
- Endotoxic shock. The shock induced by release of endotoxins from gram-negative bacteria, such as E. coli, involves TNF-mediated inflammation
- Arthritis. TNF promotes expression of nitric oxide synthetase, believed to be involved in disease pathogenesis.

Other conditions of interest are multiple sclerosis, sepsis, inflammation brought on by microbe infection, and diseases that have an autoimmune etiology, such as Type I Diabetes.

Polypeptides of this invention that promote TRRE activity can be administered with the objective of decreasing or normalizing TNF signal transduction. For example, in congestive heart failure or Crohn's disease, the polypeptide is given at regular intervals to lessen the inflammatory sequelae.

The treatment is optionally in combination with other agents that affect TNF

signal transduction (such as antibodies to TNF or receptor antagonists) or that lessen the extent of inflammation in other ways.

Polynucleotides of this invention can also be used to promote TRRE activity by gene therapy. The encoding sequence is operably linked to control elements for transcription and translation in human cells. It is then provided in a form that will promote entry and expression of the encoding sequence in cells at the disease site. Forms suitable for local injection include naked DNA, polynucleotides packaged with cationic lipids, and polynucleotides in the form of viral vectors (such as adenovirus and AAV constructs). Methods of gene therapy known to the practitioner skilled in the art will include those outlined in U.S. Patent Nos. 5,399,346, 5,827,703, and 5,866,696.

The ability to affect TNF signal transduction is also of interest where TNF is thought to play a beneficial role in resolving the disease. In particular, TNF plays a beneficial role in the necrotizing of solid tumors. Accordingly, products of this invention can be administered to cancer patients to inhibit TRRE activity, thereby increasing TNF signal transduction and improve the beneficial effect.

Embodiments of the invention that inhibit TRRE activity include antisense polynucleotides. A method of conferring long-standing inhibitory activity is to administer antisense gene therapy. A genetic construct is designed that will express RNA inside the cell which in turn will decrease the transcription of the target gene (U.S. Patent No. 5,759,829). In humans, a more frequent form of antisense therapy is to administer the effector antisense molecule directly, in the form of a short stable polynucleotide fragment that is complementary to a segment of the target mRNA (U.S. Patent Nos. 5,135,917 and 5,789,573) — in this case, the transcript that encodes the TRRE modulator. Another embodiment of the invention that inhibits TRRE are ribozymes, constructed as described in an earlier section. The function of ribozymes in inhibiting mRNA translation is described in U.S. Patent Nos. 4,987,071 and 5,591,610.

Once a product of this invention is found to have suitable TRRE modulation activity in the in vitro assays described in this disclosure, it is preferable to also test its effectiveness in an animal model of a TNF mediated

disease process. Example 3 describes an LPS model for sepsis that can be used to test promoters of TRRE activity. Example 4 describes a tumor necrosis model, in which TRRE inhibitors could be tested for an ability to enhance necrotizing activity. Those skilled in the art will know of other animal models suitable for testing effects on TNF signal transduction or inflammation. Other illustrations are the cardiac ischemia reperfusion models of Weyrich et al. (*J. Clin. Invest.* 91:2620, 1993) and Garcia-Criado et al. (*J. Am. Coll. Surg.* 181:327, 1995); the pulmonary ischemia reperfusion model of Steinberg et al. (*J. Heart Lung Transplant.* 13:306, 1994), the lung inflammation model of International Patent Application WO 9635418; the bacterial peritonitis model of Sharar et al. (*J. Immunol.* 151:4982, 1993), the colitis model of Meenan et al. (*Scand. J. Gastroenterol.* 31:786, 1996), and the diabetes model of von Herrath et al. (*J. Clin. Invest.* 98:1324, 1996). Models for septic shock are described in Mack et al. *J. Surg. Res.* 69:399, 1997; and Seljelid et al. *Scand. J. Immunol.* 45:683-7.

For use as an active ingredient in a pharmaceutical preparation, a polypeptide, polynucleotide, or antibody of this invention is generally purified away from other reactive or potentially immunogenic components present in the mixture in which they are prepared. Typically, each active ingredient is provided in at least about 90% homogeneity, and more preferably 95% or 99% homogeneity, as determined by functional assay, chromatography, or SDS polyacrylamide gel electrophoresis. The active ingredient is then compounded into a medicament in accordance with generally accepted procedures for the preparation of pharmaceutical preparations, such as described in *Remington's Pharmaceutical Sciences 18th Edition* (1990), E.W. Martin ed., Mack Publishing Co., PA. Steps in the compounding of the medicament depend in part on the intended use and mode of administration, and may include sterilizing, mixing with appropriate non-toxic and non-interfering excipients and carriers, dividing into dose units, and enclosing in a delivery device. The medicament will typically be packaged with information about its intended use.

Mode of administration will depend on the nature of the condition being treated. For conditions that are expected to require moderate dosing and that are at well perfused sites (such as cardiac failure), systemic administration is acceptable. For example, the medicament may be formulated for intravenous administration, intramuscular injection, or absorption sublingually or intranasally. Where it is possible to administer the active ingredient locally, this is usually preferred. Local administration will both enhance the concentration of the active ingredient at the disease site, and minimize effects on TNF receptors on other tissues not involved in the disease process. Conditions that lend themselves to administration directly at the disease site include cancer and rheumatoid arthritis. Solid tumors can be injected directly when close to the skin, or when they can be reached by an endoscopic procedure. Active ingredients can also be administered to a tumor site during surgical resection, being implanted in a gelatinous matrix or in a suitable membrane such as Gliadel® (Guilford Sciences). Where direct administration is not possible, the administration may be given through an arteriole leading to the disease site. Alternatively, the pharmaceutical composition may be formulated to enhance accumulation of the active ingredient at the disease site. For example, the active ingredient can be encapsulated in a liposome or other matrix structure that displays an antibody or ligand capable of binding a cell surface protein on the target cell. Suitable targeting agents include antibodies against cancer antigens, ligands for tissue-specific receptors (e.g., serotonin for pulmonary targeting). For compositions that decrease TNF signal transduction, an appropriate targeting molecule may be the TNF ligand, since the target tissue may likely display an unusually high density of the TNF receptor.

Effective amounts of the compositions of the present invention are those that alter TRRE activity by at least about 10%, typically by at least about 25%, more preferably by about 50% or 75%. Where near complete ablation of TRRE activity is desirable, preferred compositions decrease TRRE activity by at least 90%. Where increase of TRRE activity is desirable, preferred compositions increase TRRE activity by at least 2-fold. A minimum effective amount of the

active compound will depend on the disease being treated, which of the TRRE modulators is selected for use, and whether the administration will be systemic or local. For systemic administration, an effective amount of activity will generally be an amount of the TRRE modulator that can cause a change in the enzyme
5 activity by 100 to 50,000 Units — typically about 10,000 Units. The mass amount of protein, nucleic acid, or antibody is chosen accordingly, based on the specific activity of the active compound in Units per gram.

The following examples provided as a further guide to the practitioner, and are not intended to limit the invention in any way.

EXAMPLES

Example 1: Assay system for TRRE activity.

This Example illustrates an assay system that measures TRRE activity on the human TNF-R in its native conformation in the cell surface membrane

15 Membrane-associated TNF-R was chosen as the substrate, as having microenvironment similar to that of the substrate for TRRE in vivo. Membrane-associated TNF-R also requires more specific activity, which would differentiate less-specific proteases. Cells expressing an elevated level of the p75 form of TNF-R were constructed by cDNA transfection into monkey COS-1 cells which
20 express little TNF-R of either the 75 kDa or 55 kDa size.

The procedure for constructing these cells was as follows: cDNA of human p75 TNF-R was cloned from a λ gt10 cDNA library derived from human monocytic U-937 cells (Clontech Laboratories, Palo Alto, CA). The first 300 bp on both 5' and 3' ends of the cloned fragment was sequenced and compared to the
25 reported cDNA sequence of human p75 TNF-R. The cloned sequence was a 2.3 kb fragment covering positions 58-2380 of the reported p75 TNF-R sequence, which encompasses the full length of the p75 TNF-R-coding sequence from positions 90-1475. The 2.3 kb p75 TNF-R cDNA was then subcloned into the multiple cloning site of the pCDNA3 eukaryotic expression vector. The

orientation of the p75 TNF-R cDNA was verified by restriction endonuclease mapping.

Figure 1 illustrates the final 7.7 kb construct, pCDTR2. It carries the neomycin-resistance gene for the selection of transfected cells in G418, and the expression of the p75 TNF-R is driven by the cytomegalovirus promoter. The pCDTR2 was then transfected into monkey kidney COS-1 cells (ATCC CRL-1650) using the calcium phosphate-DNA precipitation method. The selected clone in G418 medium was identified and subcultured. This clone was given the designation **C75R**.

To determine the level of p75 TNF-R expression on C75R cells, 2×10^5 cells/well were plated into a 24-well culture plate and incubated for 12 to 16 hours in 5% CO₂ at 37°C. They were then incubated with 2-30 ng ¹²⁵I human recombinant TNF (radiolabeled using the chloramine T method) in the presence or absence of 100-fold excess of unlabeled human TNF at 4°C for 2 h. After three washes with ice-cold PBS, cells were lysed with 0.1N NaOH and bound radioactivity was determined in a Pharmacia Clinigamma counter (Uppsala, Sweden).

Figure 2 shows the results obtained. C75R had a very high level of specific binding of radiolabeled ¹²⁵I-TNF, while parental COS-1 cells did not. The number of TNF-R expressed on C75R was determined to be 60,000-70,000 receptors per cell by Scatchard analysis (Figure 2, inset). The K_d value calculated was 5.6×10^{-10} M. This K_d value was in close agreement to the values previously reported for native p75 TNF-R.

TRRE was obtained by PHA stimulation of THP-1 cells (WO 9802140). THP-1 cells (ATCC 45503) growing in logarithmic phase were collected and resuspended to 1×10^6 cells/ml of RPMI-1640 supplemented with 1% FCS and incubated with 10^{-6} M PMA for 30 min in 5% CO₂ at 37 °C. The cells were collected and washed once with serum-free medium to remove PMA and resuspended in the same volume of RPMI-1640 with 1% FCS. After 2 hours incubation in 5% CO₂ at 37°C, the cell suspension was collected, centrifuged, and the cell-free supernatant was collected as the source of TRRE.

In order to measure the effect of TRRE on membrane-bound TNF-R in the COS-1 cell constructs, the following experiment was performed. C75R cells were seeded at a density of 2×10^5 cells/well in a 24-well cell culture plate and incubated for 12 to 16 hours at 37°C in 5% CO₂. The medium in the wells was aspirated, replaced with fresh medium alone or with TRRE medium, and incubated for 30 min at 37°C. The medium was then replaced with fresh medium containing 30 ng/ml ¹²⁵I-labeled TNF. After 2 hours at 4°C, the cells were lysed with 0.1 N NaOH and the level of bound radioactivity was measured. The level of specific binding of C75R by ¹²⁵I-TNF was significantly decreased after incubation with TRRE. The radioactive count was 1,393 cpm on the cells incubated with TRRE compared to 10,567 cpm on the cells not treated with TRRE, a loss of 87% of binding capacity.

In order to determine the size of the p75 TNF-R cleared from C75R by TRRE, the following experiment was performed. 15×10^6 C75R cells were seeded in a 150 mm cell culture plate and incubated at 37°C in 5% CO₂ for 12 to 16 hours. TRRE medium was incubated with C75R cells in the 150 mm plate for 30 min and the resulting supernatant was collected and centrifuged. The concentrated sample was applied to 10% acrylamide SDS-PAGE and electrophoretically transferred to a polyvinylidene difluoride membrane (Immobilon). Immunostaining resulted in a single band of 40 kDa, similar to the size found in biological fluids. Thus, transfected COS-1 cells expressed high levels of human p75 TNF-R in a form similar to native TNF-R.

The following assay method was adopted for routine measurement of TRRE activity. C75R cells and COS-1 cells were seeded into 24-well culture plates at a density of 2.5×10^5 cells/ml/well and incubated overnight (for 12 to 16 hours) in 5% CO₂ at 37°C. After aspirating the medium in the well, 300 µl of TRRE medium was incubated in each well of both the C75R and COS-1 plates for 30 min in 5% CO₂ at 37°C (corresponding to A and C mentioned below, respectively). Simultaneously, C75R cells in 24-well plates were also incubated with 300 µl of fresh medium or buffer. The supernatants were collected,

centrifuged, and then assayed for the concentration of soluble p75 TNF-R by ELISA.

ELISA assay for released TNF-R (WO 9802140) was performed as follows: Polyclonal antibodies to human p75 TNF-R were generated by immunization of New Zealand white female rabbits (Yamamoto et al. *Cell. Immunol.* 38:403-416, 1978). The IgG fraction of the immunized rabbit serum was purified using a protein G (Pharmacia Fine Chemicals, Uppsala, Sweden) affinity column (Ey et al. (1978) *Immunochemistry* 15:429-436, 1978). The IgG fraction was then labeled with horseradish peroxidase (Sigma Chemical Co., St. Louis, MO) (Tijssen and Kurstok, *Anal. Biochem.* 136:451-457, 1984). In the first step of the assay, 5 µg of unlabeled IgG in 100 µl of 0.05 M carbonate buffer (pH 9.6) was bound to a 96-well ELISA microplate (Corning, Corning, NY) by overnight incubation at 4°C. Individual wells were washed three times with 300 µl of 0.2% Tween-20 in phosphate buffered saline (PBS). The 100 µl of samples and recombinant receptor standards were added to each well and incubated at 37°C for 1 to 2 hours. The wells were then washed in the same manner, 100 µl of horseradish peroxidase-labeled IgG added and incubated for 1 hour at 37 °C. The wells were washed once more and the color was developed for 20 minutes (min) at room temperature with the substrates ABTS (Pierce, Rockford, IL) and 30% H₂O₂ (Fisher Scientific, Fair Lawn, NJ). Color development was measured at 405 nm.

When C75R cells were incubated with TRRE medium, soluble p75 TNF-R was released into the supernatant which was measurable by ELISA. The amount of receptors released corresponded to the amount of TRRE added. There was also a level of spontaneous TNF-R release in C75R cells incubated with just medium alone. It is hypothesized that this is due to an endogenous source of proteolytic enzyme, a homolog of the human TRRE of monkey origin.

The following calculations were performed. A = (amount of soluble p75 TNF-R in a C75R plate treated with the TRRE containing sample); i.e. the total amount of sTNF-R in a C75R plate. B = (amount of soluble p75 TNF-R spontaneously released in a C75R plate treated with only medium or buffer

containing the same reagent as the corresponding samples but without exogenous TRRE); i.e. the spontaneous release of sTNF-R from C75R cells. C = (amount of soluble p75 TNF-R in a COS-1 plate treated with the TRRE sample or the background level of soluble p75 TNF-R released by THP-1.); i.e. the degraded value of transferred (pre-existing) sTNF-R in the TRRE sample during 30 min incubation in a COS-1 plate. This corresponds to the background level of sTNF-R degraded in a C75R plate. The net release of soluble p75 TNF-R produced only by TRRE activity existing in the initial sample is calculated as follows: (Net release of soluble p75 TNF-R only by TRRE) = A - B - C.

Unit activity of TRRE was defined as follows: 1 pg of soluble p75 TNF-R net release (A-B-C) in the course of the assay is one unit (U) of TRRE activity.

Using this assay, the time course of receptor shedding by TRRE was measured in the following experiment. TRRE-medium was incubated with C75R and COS-1 cells for varying lengths of time. The supernatants were then collected and assayed for the level of soluble p75 TNF-R by ELISA and the net TRRE activity was calculated. Detectable levels of soluble receptor were released by TRRE within 5 min and increased up to 30 min. Longer incubation times showed that the level of TRRE remained relatively constant after 30 min, presumably from the depletion of substrates. Therefore, 30 min was determined to be the optimal incubation time.

The induction patterns of TRRE and known MMPs by PMA stimulation are quite different. In order to induce MMPs, monocytic U-937 cells, fibrosarcoma HT-1080 cells, or peritoneal exudate macrophages (PEM) usually have to be stimulated for one to three days with LPS or PMA. On the other hand, as compared with this prolonged induction, TRRE is released very quickly in culture supernatant following 30 min of PMA-stimulation. The hypothesis that TRRE and sTNF-R form a complex *in vitro* was confirmed by the experiment that 25% TRRE activity was recovered from soluble p75 TNF-R affinity column. This means that free TRRE has the ability to bind to its catalytic product, sTNF-R. The remaining 75% which did not combine to the affinity column may already be

bound to sTNF-R or may not have enough affinity to bind to sTNF-R even though it is in a free form.

Example 2: Characterization of TRRE obtained from THP-1 cells.

5 TRRE obtained by PHA stimulation of THP-1 cells was partially purified from the culture medium (WO 9802140). First, protein from the medium was concentrated by 100% saturated ammonium sulfate precipitation at 4°C. The precipitate was pelleted by centrifugation at 10,000 x g for 30 min and resuspended in PBS in approximately twice the volume of the pellet. This
10 solution was then dialyzed at 4°C against 10 mM Tris-HCl, 60 mM NaCl, pH 7.0. This sample was loaded on an anion-exchange chromatography, Diethylaminoethyl (DEAE)-Sephadex A-25 column (Pharmacia Biotech) (2.5 x 10 cm) previously equilibrated with 50 mM Tris-HCl, 60 mM NaCl, pH 8.0. TRRE was then eluted with an ionic strength linear gradient of 60 to 250 mM
15 NaCl, 50 mM Tris-HCl, pH 8.0. Each fraction was measured for absorbance at 280 nm and assayed for TRRE activity. The DEAE fraction with the highest specific activity (the highest value of TRRE units/A280) was pooled and used in the characterizations of TRRE described in this example.

In the next experiment, the substrate specificity of the enzyme was
20 elucidated using immunohistochemical techniques. Fluorescein isothiocyanate (FITC)-conjugated anti-CD54, FITC-conjugated goat anti-rabbit and mouse antibodies, mouse monoclonal anti-CD30, anti-CD11b and anti-IL-1R (Serotec, Washington D.C.) were used. Rabbit polyclonal anti-p55 and p75 TNF-R were obtained according to Yamamoto et al. (1978) *Cell Immunol.* 38:403-416. THP-
25 1 cells were treated for 30 min with 1,000 and/or 5,000 U/ml of TRRE eluted from the DEAE-Sephadex column, and then transferred to 12 x 75 mm polystyrene tubes (Fischer Scientific, Pittsburgh, PA) at 1×10^5 cells/100 μ l/tube. The cells were then pelleted by centrifugation at 350 x g for 5 min at 4°C and stained directly with 10 μ l FITC-conjugated anti-CD54 (diluted in cold PBS/0.5% sodium
30 aside), indirectly with FITC-conjugated anti-mouse antibody after treatment of

mouse monoclonal anti-CD11b, IL-1R and CD30 and also indirectly with FITC-conjugated anti-rabbit antibody after treatment of rabbit polyclonal anti-p55 and p75 TNF-R.

THP-1 cells stained with each of the antibodies without treatment of TRRE were used as negative controls. The tubes were incubated for 45 min at 4°C, agitated every 15 min, washed twice with PBS/2% FCS, repelleted and then resuspended in 200µl of 1% paraformaldehyde. These labeled THP-1 cells were analyzed using a fluorescence activated cell sorter (FACS) (Becton-Dickinson, San Jose, CA) with a 15 mW argon laser with an excitation of 488 nm. Fluorescent signals were gated on the basis of forward and right angle light scattering to eliminate dead cells and aggregates from analysis. Gated signals (10⁴) were detected at 585 BP filter and analyzed using Lysis II software. Values were expressed as percentage of positive cells, which was calculated by dividing mean channel fluorescence intensity (MFI) of stained THP-1 cells treated with TRRE by the MFI of the cells without TRRE treatment (negative control cells).

To test the *in vitro* TNF cytolytic assay by TRRE treatment the L929 cytolytic assay was performed according to the method described by Gatanaga et al. (1990b). Briefly, L929 cells, an adherent murine fibroblast cell line, were plated (70,000 cells/0.1ml/well in a 96-well plate) overnight. Monolayered L929 cells were pretreated for 30 min with 100, 500 or 2,500 U/ml of partially-purified TRRE and then exposed to serial dilutions of recombinant human TNF for 1 hour. After washing the plate with RPMI-1640 with 10% FCS to remove the TRRE and TNF, the cells were incubated for 18 hours in RPMI-1640 with 10% FCS containing 1 µg/ml actinomycin D at 37°C in 5% CO₂. Culture supernatants were then aspirated and 50 µl of 1% crystal violet solution was added to each well. The plates were incubated for 15 min at room temperature. After the plates were washed with tap water and air-dried, the cells stained with crystal violet were lysed by 100 µl per well of 100 mM HCl in methanol. The absorbance at

550 nm was measured using an EAR 400 AT plate reader (SLT-Labinstruments, Salzburg, Austria).

To investigate whether TRRE also truncates the ~55 kDa size of TNF-R, partially-purified TRRE was applied to THP-1 cells which express low levels of both p55 and p75 TNF-R (approximately 1,500 receptors/cell by Scatchard analysis). TRRE eluate from the DEAE-Sephadex column was added to THP-1 cells (5×10^6 cells/ml) at a final TRRE concentration of 1,000 U/ml for 30 min. The concentration of soluble p55 and p75 TNF-R in that supernatant was measured by soluble p55 and p75 TNF-R ELISA. TRRE was found to truncate both human p55 and p75 TNF-R on THP-1 cells and released 2,382 and 1,662 pg/ml soluble p55 and p75 TNF-R, respectively.

Therefore, TRRE obtained by PHA stimulation of THP-1 cells is capable of enzymatically cleaving and releasing human p75 TNF-R on C75R cells, and both human p55 and p75 TNF-R on THP-1 cells.

Partial inhibition of TRRE activity was obtained by chelating agents such as 1,10-phenanthroline, EDTA and EGTA (% TRRE activity remaining were 41%, 67% and 73%, respectively, at 2 mM concentration). On the other hand, serine protease inhibitors such as PMSF, AEBSF and 3,4-DCI, and serine and cysteine protease inhibitors such as TLCK and TPCK had no effect on the inhibition of TRRE. TRRE was slightly activated in the presence of Mn^{2+} , Ca^{2+} , Mg^{2+} , and Co^{2+} (% TRRE activities remaining were 157%, 151%, 127%, and 123%, respectively), whereas partial inhibition occurred in the presence of Zn^{2+} and Cu^{2+} (% TRRE activities remaining were 23% and 47%, respectively) (WO 9802140).

TRRE fractions from the most active DEAE fraction (60 mM to 250 mM NaCl) can be purified further. In one method (WO 9802140), the fractions were concentrated to 500 μ L with a Centriprep-10 filter (10,000 MW cut-off membrane) (Amicon). This concentrated sample was applied to 6% PAGE under non-denaturing native conditions. The gel was sliced horizontally into 5 mm strips and each was eluted into 1 ml PBS. The eluates were then tested according to the assay (Example 1) for TRRE activity.

Example 3: TRRE activity alleviates septic shock

The following protocol was used to test the effects of TRRE in preventing mortality in a model for septic shock. Mice were injected with lethal or sublethal levels of LPS, and then with a control buffer or TRRE. Samples of peripheral blood were then collected at intervals to establish if TRRE blocked TNF-induced production of other cytokines in the bloodstream. Animals were assessed for the ability of TRRE to block the clinical effects of shock, and then euthanized and tissues examined by histopathological methods.

Details were as follows: adult Balb/c mice, were placed in a restraining device and injected intravenously via the tail vein with a 0.1 ml solution containing 10 ng to 10 mg of LPS in phosphate buffer saline (PBS). These levels of LPS induce mild to lethal levels of shock in this strain of mice. Shock results from changes in vascular permeability, fluid loss, and dehydration, and is often accompanied by symptoms including lethargy, a hunched, stationary position, rumped fur, cessation of eating, cyanosis, and, in serious cases, death within 12 to 24 hours. Control mice received an injection of PBS. Different amounts (2,000 or 4,000 U) of purified human TRRE were injected IV in a 0.1 ml volume within an hour prior to or after LPS injection. Serum (0.1 ml) was collected with a 27 gauge needle and 1 ml syringe IV from the tail vein at 30, 60 and 90 minutes after LPS injection. This serum was heparinized and stored frozen at -20°C. Samples from multiple experiments were tested by ELISA for the presence of sTNF-R, TNF, IL-8 and IL-6. Animals were monitored over the next 12 hours for the clinical effects of shock. Selected animals were euthanized at periods from 3 to 12 hours after treatment, autopsied and various organs and tissues fixed in formalin, imbedded in paraffin, sectioned and stained by hematoxylin-eosin (H and E). Tissue sections were subjected to histopathologic and immunopathologic examination.

Figure 3 shows the results obtained. (♦) LPS alone; (■) LPS plus control buffer; (●) LPS plus TRRE (2,000 U); (▲) LPS plus TRRE (4,000 U).

Mice injected with LPS alone or LPS and a control buffer died shortly after injection. 50% of the test animals were dead after 8 hours (LPS) or 9 hours (LPS plus control buffer), and 100% of the animals were dead at 15 hours. In contrast, animals treated with TRRE obtained as described in Example 1 did much better. When injections of LPS were accompanied by injections of a 2,000 U of TRRE, death was delayed and death rates were lower. Only 40% of the animals were dead at 24 hours. When 4,000 U of TRRE was injected along with LPS, all of the animals had survived at 24 hours. Thus, TRRE is able to counteract the mortality induced by LPS in test animals.

Example 4: TRRE activity decreases tumor necrotizing activity

The following protocol was followed to test the effects of TRRE on tumor necrosis in test animals in which tumors were produced, and in which TNF was subsequently injected.

On Day 0, cutaneous Meth A tumors were produced on the abdominal wall of fifteen BALB/c mice by intradermal injection of 2×10^5 Meth A tumor cells. On Day 7, the mice were divided into three groups of five mice each and treated as follows:

- Group 1: Injected intravenously with TNF ($1 \mu\text{g}/\text{mouse}$).
- Group 2: Injected intravenously with TNF ($1 \mu\text{g}/\text{mouse}$) and injected intratumorally with TRRE obtained as in Example 1 (400 units/mouse, 6, 12 hours after TNF injection).
- Group 3: Injected intravenously with TNF ($1 \mu\text{g}/\text{mouse}$) and injected intratumorally with control medium (6, 12 hours after TNF injection).

On Day 8, tumor necrosis was measured with the following results: Group 1: 100% of necrosis (5/5); Group 2: 20% (1/5); Group 3: 80% (4/5). Injections of TRRE greatly reduced the ability of TNF to induce necrosis in Meth A tumors in BALB/c mice.

Since adding TRRE activity ablates the beneficial necrotizing activity of TNF, blocking endogenous TRRE activity would promote the beneficial effects of TNF.

5 **Example 5: Nine new polynucleotide clones that affect TRRE activity**

A number of cells have been found to express high levels of TRRE activity, especially after PMA stimulation. These include the cell lines designated THP-1, U-937, HL-60, ME-180, MRC-5, Raji, K-562. Jurkat cells have a high TRRE activity (850 TRRE U/mL at 10^{-2} PMA). In this experiment, the expression
10 library of the Jurkat T cell (ATCC #TIB-152) was obtained and used to obtain 9 polynucleotide clones that augment TRRE activity.

Selection of expression sequences in the library was done by repeated cycles of transfection into COS-1 cells, followed by assaying of the supernatant as in Example 1 for the presence of activity cleaving and releasing the TNF
15 receptor. Standard techniques were used in the genetic manipulation. Briefly, the DNA of 10^6 Jurkat cells was extracted using an InVitrogen plasmid extraction kit according to manufacturer's directions. cDNA was inserted in the ZAP Express™/EcoRI vector (cat. no. 938201, Stratagene, La Jolla CA. The library was divided into 48 groups of DNA and transformed into COS-1 cells using the
20 CaCl transfection method. Once the cells were grown out, the TRRE assay was performed, and five positive groups were selected. DNA from each of these five groups was obtained, and transfected into *E. coli*, with 15 plates per group. DNA was prepared from these cells and then transfected into COS-1 cells once more. The cells were grown out, and TRRE activity was tested again. Two positive
25 groups were selected and transfected into *E. coli*, yielding 98 colonies. DNA was prepared from 96 of these colonies and transfected into COS-1 cells. The TRRE activity was performed again, and nine clones were found to substantially increase TRRE activity in the assay. These clones were designated 2-8, 2-9, 2-14, 2-15, P2-2, P2-10, P2-13, P2-14, and P2-15.

30 **Figure 4** is a bar graph showing the TRRE activity observed when the 9 clones were tested with C75 cells in the standard assay (Example 1).

These nine clones were then sequenced according to the following procedure:

1. Plasmid DNA was prepared using a modified alkaline lysis procedure.
2. DNA sequencing was performed using DyeDeoxy termination reactions (ABI). Base-specific fluorescent dyes were used as labels.
3. Sequencing reactions were analyzed on 5.75% Long Ranger™ gels by an ABI 373A-S or on 5.0% Long Ranger™ gels by an ABI 377 automated sequencer.
4. Subsequent data analysis was performed using Sequencher™ 3.0 software.

Standard primers T7X, T3X, -40, -48 Reverse, and BK Reverse (BKR) were used in sequencing reactions. For each clone, several additional internal sequencing primers (listed below) were synthesized.

NCBI BLAST (Basic Local Alignment Search Tool) sequence analysis (Altschul et al. (1990) *J. Mol. Biol.* 215:403-410) was performed to determine if other sequences were significantly similar to these sequences. Both the DNA sequences of the clones and the corresponding ORFs (if any) were compared to sequences available in databases.

The following clones were obtained and sequenced:

TABLE 1: DNA sequences affecting TRRE activity

Clone	Sequence Designation	SEQ ID NO:	Approx Length (bp)	Expression Designation	Related sequences (potential homology)
2-9	AIM2	1	4,047		—
2-8	AIM3T3 (partial sequence)	2	739		<i>M. musculus</i> 45S pre-rRNA gene
	AIM3T7 (partial sequence)	3	233		
2-14	AIM4	4	2,998	Mey3	human arfaptin 2 and others (see below)
2-15	AIM5	5	4,152		—
P2-2	AIM6	6	3,117	Mey5	—
P2-10	AIM7	7	3,306	Mey6	Human Insulin-like Growth factor II Receptor
P1-13	AIM8	8	4,218		—
P2-14	AIM9	9	1,187	Mey8	—
P2-15	AIM10	10	3,306		E1b-55kDa-associated protein

Clone 2-9 (AIM2): The internal primers used for sequencing are shown in SEQ. ID NOS:11-38. The sequence of AIM2 is presented in SEQ ID NO:1. The complementary strand of the AIM2 sequence is SEQ ID NO:147. The longest open reading frame (ORF) in the AIM2 sequence is 474 AA long and represented in SEQ ID NO:148.

Clone 2-8 (AIM3): Two partial sequences of length 739 and 233 were obtained and designated AIM3T3 and AIM3T7. The internal primers used for sequencing are shown in SEQ. ID NOS:39-46. The sequences of AIM3T3 and

AIM3T7 are presented in SEQ ID NOs:2 and 3, respectively. The BLAST search revealed that the AIM3T3 sequence may be homologous to the mouse (*M. musculus*) 28S ribosomal RNA (Hassouna et al. *Nucleic Acids Res.* 12:3563-3583, 1984) and the *M. musculus* 45S pre-rRNA genes (Accession No. X82564.

- 5 The complementary sequence of the AIM3T3 sequence showed 99% similarity over 408 bp beginning with nt 221 of SEQ ID NO:2 to the former and 97% similarity over the same span to the latter.

Clone 2-14 (AIM4). The internal primers used for sequencing are shown in SEQ. ID NOS:14-65. The sequence of AIM4 is presented in SEQ ID NO:4. The
10 complementary strand of the AIM4 sequence is SEQ ID NO:149. The longest ORF in the AIM4 sequence is 236 AA long and represented in SEQ ID NO:150. AIM4 has significant alignments to human sequences arfaptin 2, ADE2H1 mRNA showing homologies to SAICAR synthetase, polypyrimidine tract binding protein (heterogeneous nuclear ribonucleoprotein I) mRNA, several PTB genes for
15 polypyrimidine tract binding proteins, mRNA for por1 protein. Human arfaptin 2 is a putative target protein of ADP-ribosylation factor that interacts with RAC1 by binding directly to it. RAC1 is involved in membrane ruffling. Arfaptin 2 has possible transmembrane segments, potential CK2 phosphorylation sites, PKC phosphorylation site and RGD cell attachment sequence.

- 20 *Clone 2-15 (AIM5):* The internal primers used for sequencing are shown in SEQ. ID NOS:66-80. The sequence of AIM5 is presented in SEQ ID NO:5. The BLAST search revealed that the AIM5 sequence displays some similarity to Human Initiation Factor 5A (eIF-5A) Koettwitz et al. (1995) *Gene* 159:283-284, 1995 and Human Initiation Factor 4D (eIF 4D) Smit-McBride et al. (1989) *J. Biol.*
25 *Chem.* 264:1578-1583, 1989.

Clone P2-2 (AIM6): The internal primers used for sequencing are shown in SEQ. ID NOS:81-93. The sequence of AIM6 is presented in SEQ ID NO:6. The longest ORF in the AIM6 sequence is 1038 AA long and represented in SEQ ID NO:151.

- 30 *Clone P2-10 (AIM7):* The internal primers used for sequencing are shown in SEQ. ID NOS:94-106. The sequence of AIM7 is presented as SEQ ID NO:7.

The longest ORF in the AIM7 sequence is 849 AA long and represented in SEQ ID NO:152. The BLAST search revealed that this clone may be related to the Human Insulin-like Growth Factor II Receptor (Morgan et al. *Nature* 329:301-307, 1987 or the Human Cation-Independent Mannose 6-Phosphate Receptor mRNA (Oshima et al. *J. Biol. Chem.* 263:2553-2562, 1988). The AIM7 sequence showed roughly 99% identity to both sequences over 2520 nucleotides beginning with nt 12 of SEQ ID NO:7 and 99% similarity to the latter over the same span.

Clone P2-13 (AIM8): The internal primers used for sequencing are shown in SEQ. ID NOS:107-118. The sequence of AIM8 is presented as SEQ ID NO:8. The longest ORF in the AIM8 sequence is 852 AA long and represented in SEQ ID NO:153.

Clone P2-14 (AIM9): The internal primers used for sequencing are shown in SEQ. ID NOS:119-124. The sequence of AIM9 is presented as SEQ ID NO:9. The longest ORF was about 149 amino acids in length.

Clone P2-15 (AIM10): The internal primers used for sequencing are shown in SEQ. ID NOS:125-146. The sequence of AIM10 is presented as SEQ ID NO:10. The longest ORF in the AIM10 sequence is 693 AA long and represented in SEQ ID NO:154. Sequence 10 on BLASTN search of non-redundant databases at NCBI aligns with Human mRNA for E1b-55kDa-associated protein, locus HSA7509 (Accession AJ007509, NID g3319955).

Clonal DNA may be directly injected into test animals in order to test the ability of these nucleic acids to induce TRRE activity, counteract septic shock and/or affect tumor necrosis, as is described in detail in Examples 3 and 4. Alternatively, proteins or RNA can be generated from the clonal DNA for similar testing.

Example 6: Expression of newly obtained clones

Example 5 describes 9 new clones which enhance TRRE activity in a cell surface assay system. The clones were obtained in the pBK-CMB Phagmid vector .

The following work was done on contract through the commercial laboratory Lark Technologies, Houston, TX. The clones were removed from shuttle vectors and inserted into expression vectors in the following manner. Recombinant plasmid (pBK-CMV containing insert) was digested with appropriate restriction enzyme(s) such as Spe I, Xba I, EcoR I or others, as appropriate. The Baculovirus Transfer Vector (pAcGHLT-A Baculovirus Transfer Vector, PharMingen, San Diego, CA, Cat. No. 21460P) was also cut with appropriate restriction enzyme(s) within or near the multiple cloning site to receive the insert removed from the shuttle vector.

The fragment of interest being subcloned was isolated from the digest using Low-Melting agarose electrophoresis and purified from the gel using a Qiaquick Gel Extraction Kit following Lark SOP MB 020602. If necessary, the receiving vector was treated with alkaline phosphatase according to Lark SOP MB 090201. The fragment was ligated into the chosen site of the vector pAcGHLT-A. The recombinant plasmid was transformed into *E. coli* XL1 Blue MRF' cells and the transformed bacterial cells were selected on LB agar plates containing ampicillin (100µg/ml). Ampicillin resistant colonies were picked and grown on LB broth containing ampicillin for plasmid preparation.

Plasmid DNA was prepared using Alkaline Minilysate Procedure (Lark SOP MB 010802 and digested with appropriate restriction enzyme(s). Selected subclones were confirmed to be of the correct size. Subclones were digested with other appropriate restriction enzyme(s) to ascertain correct orientation of the insert by confirming presence of fragments of proper size(s). A subclone was grown in 100 ml of LB broth containing ampicillin (100µg/ml) and the plasmid DNA prepared using Qiagen Midi Plasmid Preparation Kit (Lark SOP MB 011001). The DNA concentration was determined by measuring the absorbance at 260 nm and the DNA sample was verified to be originated from correct subclone by restriction digestion.

Thus were produced the expression constructs for Mey3, Mey5, Mey6, Mey8 now with the coding sequence of interest fused to GST gene with polyhistitidine tag, protein kinase A site and thrombin cleavage site. The GST

gene and now the fusion protein are under the polyhedrin promotor. PharMingen (San Diego, CA) incorporated the vector with insert into functional baculovirus particles by co-inserting the transfer vector (pAcGHLT) into susceptible insect cell line S along with linearized virus DNA (PharMingen, San Diego, CA, BaculoGold viral DNA, Cat. No. 21100D). The functional virus particles were grown again on the insect cells to generate a high titer stock. Protein production was then done by infecting a large culture of cells in Tini cell. The cells were harvested when the protein yield reached a maximum and before the virus killed the cells. Fusion proteins were collected on a glutathione-agarose column, washed and released with glutathionine.

Proteins collected from the affinity column were quantified by measuring OD₂₈₀ and were assayed on gels using SDS-PAGE and Western blotting with labeled anti-GST (PharMingen, San Diego, CA, mAbGST Cat. No. 21441A) to confirm that all the bands present included the GST portion.

Four of the ten sequences have been cloned, expressed in baculovirus infected insect cells, and then purified.

TABLE 2: Expressed protein from Jurkat library clones		
Name	Sequence in insert	Amount of protein (mg/mL)
Mey3	AIM4	4.7, 5.0
Mey5	AIM6	1.36, 1.50
Mey6	AIM7	0.33
Mey8	AIM9	1.53

Gels indicated the presence of the GST protein in addition to larger proteins that were also positive with the anti-GST antibody in Western analyses. Mey3 repeatedly exhibited the presence of proteins around 32kDa, 56kDa, bands around 60-70kDa and another larger than 70kDa. Mey5 consistently had proteins migrating as approximately 34kDa, 38kDa, 58kDa, around 60-70kDa, and others larger than 70kDa. Mey6 had protein bands around 34kDa, 56kDa,

58kDa, and bands around 60-70kDa. Mey8 had protein bands around 36kDa, 58kDa and bands around 60-70kDa. All of the indicated bands were positive for GST. The bands may represent the desired fusion protein or degradation/cleavage product generated during growth and purification.

5

Example 7: Assay of expression products for effect on TNF-R cleaving activity

The following method was used to measure TRRE activity of Mey 3, 5, 6 and 8. C75R cells and COS-1 cells were seeded into 24-well culture plates at a density of 2.5×10^5 cells/ml/well and incubated overnight (for 12 to 16 hours) in 5% CO₂ at 37°C. After aspirating the medium in the well, 300µl of 1 µg of Mey 3, 5 and 8 were incubated in each well of both the C75R and COS-1 plates for 30 min in 5% CO₂ at 37°C (corresponding to A and C mentioned below, respectively). Simultaneously, C75R cells in 24-well plates were also incubated with 300µl of fresh medium or buffer (corresponding to B mentioned below). The supernatants were collected, centrifuged, and then assayed for the concentration of soluble p75 TNF-R by ELISA as described in Example 1.

The following results were obtained:

TABLE 3: Enzymatic activity of expressed clones	
Clone No.	TNF-receptor releasing activity U/mg
Mey-3	341
Mey-5	671
Mey-6	452
Mey-8	191

20

Example 8: Effectiveness of expression products in treating septic shock

The protocol outlined in Example 3 was used to test the effects of the expression products from the new clones in preventing mortality in the septic shock model.

- 5 Different amounts of recombinant Mey 3, 5, and 8 (10 – 100 ug/mouse) were injected i.v. in a 0.05 ml volume within an hour prior to or after injection of a lethal dose of LPS. Serum (0.1ml) was collected using a 27 gauge needle and 1 ml syringe from the tail vein at 30, 60 and 90 minutes after LPS injection. This serum was heparinized and stored frozen at -20°C. Samples from multiple
10 experiments were tested by ELISA for the presence of solubilized TNR-R, the TNR ligand, IL-8, and IL-6. Animals were monitored over the next 12 hours for the clinical effects of shock. Selected animals were euthanized from 3 to 12 hours after treatment, autopsied and various organs and tissues fixed in formalin, imbedded in paraffin, sectioned and stained by hematoxylin-eosin (H and E).
15 Tissue sections were subjected to histopathologic and immunopathologic examination.

Figure 5 shows the results obtained. (♦) saline; (■) BSA; (Δ) Mey-3 (100 µg); (X) Mey-3 (10 µg); (*) Mey-5 (10 µg); (●) Mey-8 (10 µg).

- Mice injected with LPS alone or LPS, a control buffer or control protein
20 (BSA) died rapidly. All of the animals in this group were dead at 24 hours. In contrast, when injections of LPS were accompanied by injections of a 10 – 100 ug of Mey 3, 5 and 8, death was delayed and death rates were lower. None of the animal were dead at 24 hours that had been treated with Mey 3 and Mey 5. Only 66 % of the animals were dead at 24 hours that had been treated with Mey
25 8. Thus, Mey 3, 5 and 8 were able to counteract the mortality induced by LPS in test animals.

ART 34 AMDT

CLAIMS

1. An isolated polynucleotide comprising a nucleotide sequence expressed at the mRNA level in human mononuclear leukocytes having cell-surface TNF receptor, thereby increasing cleavage and release of the receptor from the surface of the cell.
2. The polynucleotide of claim 1, wherein the nucleotide sequence is contained in a sequence selected from the group consisting of
 - a) SEQ. ID NO:1;
 - b) SEQ. ID NO:2 and SEQ. ID NO:3;
 - c) SEQ. ID NO:4;
 - d) SEQ. ID NO:5;
 - e) SEQ. ID NO:6;
 - f) SEQ. ID NO:7;
 - g) SEQ. ID NO:8;
 - h) SEQ. ID NO:9; and
 - i) SEQ. ID NO:10.
3. An isolated polynucleotide comprising at least 30 consecutive nucleotides in said nucleotide sequence of a polynucleotide according to any of claims 1-2.
4. An isolated polynucleotide comprising a linear sequence of at least 50 consecutive nucleotides at least 90% identical to a sequence contained in said nucleotide sequence of the polynucleotide of claim 1.
5. An isolated polynucleotide of at least 50 nucleotides capable of hybridizing specifically to said nucleotide sequence of a polynucleotide according to any of claims 1-3 at 68°C in 0.5 M phosphate buffer pH 7, 7% SDS, and 100 µg/mL salmon sperm DNA, followed by washing in a buffer containing 3X SSC.
6. An antisense polynucleotide or ribozyme comprising at least 10 consecutive nucleotides in said nucleotide sequence of a polynucleotide according to claim 1 or 2, which inhibits the expression of a TRRE modulator.
7. An isolated polypeptide comprising an amino acid sequence encoded by a polynucleotide according to any of claims 1-5.

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ART 34 AMDI

8. The polypeptide of claim 7, selected from the group consisting of SEQ. ID NOS: 147-154.
9. An isolated polypeptide, comprising at least 10 consecutive residues in said amino acid sequence of a polypeptide according to claim 7 or 8.
10. An isolated polypeptide, comprising at least 15 consecutive amino acids which are at least 80% identical to a sequence contained in said amino acid sequence of the polypeptide according to claim 7 or 8.
11. The polypeptide of claim 7-10, which when incubated with COS-1 cells expressing TNF receptor, promotes enzymatic cleavage and release of the receptor.
12. The polypeptide of claims 7-11, which either:
- a) lacks a membrane spanning sequence; or
 - b) is produced by a process comprising recombinant expression in a host cell followed by purification of the polypeptide from medium in which the cell is cultured.
13. A method of producing the polypeptide according to any of claims 7 to 11, comprising the steps of:
- a) culturing host cells genetically altered to express the polynucleotide according to claim 3; and subsequently
 - b) purifying the polypeptide from the cells.
14. The method according to claim 13, comprising harvesting culture medium following step a); and purifying the polypeptide from the culture medium by a process comprising affinity chromatography.
15. An isolated polynucleotide encoding the polypeptide of claim 8 or 9.
16. An isolated antibody specific for a polypeptide according any of claims 7-11.
17. A method for producing the antibody according to claim 16, comprising immunizing a mammal or contacting an immunocompetent cell or particle with a polypeptide according to claim 9 or 10.

ART 34 AMDT

18. An assay method of determining altered TRRE activity in a cell or tissue sample, comprising the steps of:
 - a) contacting the sample with the polynucleotide of claim 4 or 5 under conditions that permit the polynucleotide to hybridize specifically with nucleic acid that encodes a modulator of TRRE activity, if present in the sample, and
 - b) determining polynucleotide that has hybridized as a result of step a), as a measure of altered TRRE activity in the sample.
19. An assay method for determining altered expression of a modulator of TRRE activity in a cell or tissue sample, comprising the steps of:
 - a) contacting the sample with the antibody of claim 16 under conditions that permit the antibody to bind the modulator if present in the sample, thereby forming an antibody-antigen complex; and
 - b) determining complex formed in step a), as a measure of the modulator.
20. A method for assessing a disease condition associated with altered TRRE activity in a subject, comprising determining altered TRRE activity in the sample from the subject according to claim 18, or determining altered expression of a TRRE modulator according to claim 19, and then correlating the extent of alteration with the disease condition.
21. A method for decreasing signal transduction from a cytokine into a cell, comprising contacting the cell with a polypeptide according to any of claims 7-8 and 11-12, or with a polynucleotide according to any of claims 1-3 and 15.
22. A method for increasing signal transduction from a cytokine into a cell, comprising contacting the cell with a polynucleotide according to claim 6, or with an antibody according to claim 16.
23. The method according to claim 21 or claim 22, wherein the cytokine is TNF.
24. A method for screening polynucleotides for an ability to modulate TRRE activity, comprising the steps of:
 - a) providing cells that express both TRRE and the TNF-receptor;
 - b) genetically altering the cells with the polynucleotides to be screened;
 - c) cloning the cells genetically altered in step b); and
 - d) identifying clones that enzymatically release the receptor at an altered rate.

ART 34 AMDT

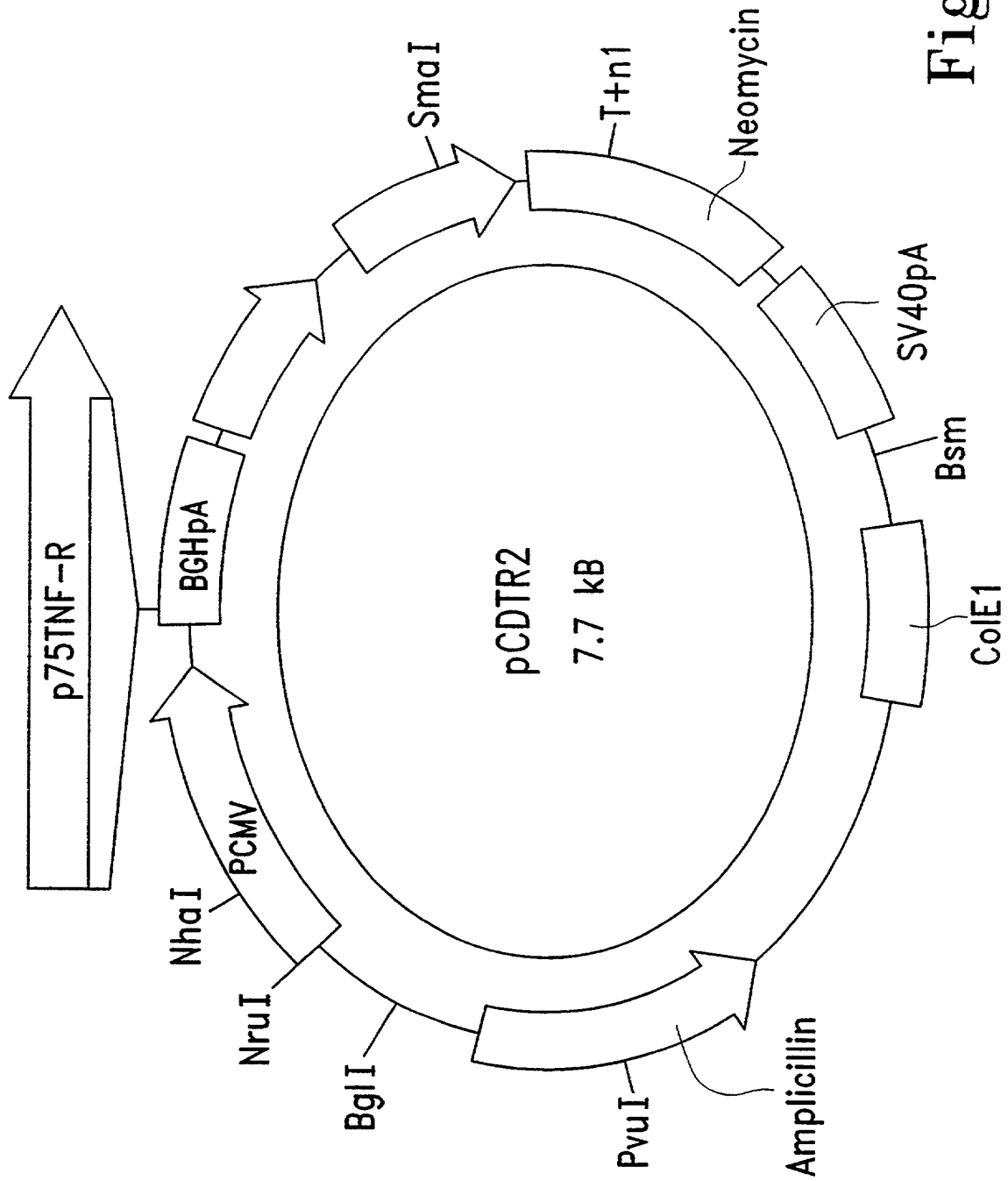
25. A method for screening substances for an ability to affect TRRE activity, comprising the steps of:
- incubating cells expressing TNF receptor with a polypeptide according to claim 9 in the presence of the substance;
 - incubating cells expressing TNF receptor with a polypeptide according to claim 9 in the absence of the substance;
 - measuring any TNF receptor released from the cells in steps a) and b); and
 - correlating an increase or decrease of the receptor released in step a) relative to that in step b) with an ability of the substance to enhance or diminish TRRE activity.
26. Use of a polypeptide according to any of claims 7-8 or 11-12, in the preparation of a medicament for treatment of the human or animal body by surgery or therapy.
27. Use of a polynucleotide according to any of claims 1-3, 6, or 15 in the preparation of a medicament for treatment of the human or animal body by surgery or therapy.
28. Use of an antibody according to claim 16, in the preparation of a medicament for treatment of the human or animal body by surgery or therapy.
29. Use of a polypeptide according to any of claims 7-8 and 11-12, a polynucleotide according to any of claims 1-3 and 15 or an antibody according to claim 16, in the preparation of a medicament for treatment of a disease selected from the group consisting of heart failure, cachexia, inflammation, endotoxic shock, arthritis, multiple sclerosis, and sepsis.
30. A method of treating cancer in a subject, comprising increasing signal transduction from TNF into cells at the site of the cancer in the subject according to claim 22 or 23.
31. A method of treating a disease selected from the group consisting of heart failure, cachexia, inflammation, endotoxic shock, arthritis, multiple sclerosis, and sepsis, comprising decreasing signal transduction from TNF into cells at the site of the disease in the subject according to claim 21 or 23.
32. The method of claim 31, comprising administering to the subject an effective amount of the polypeptide of any of claims 7-8 or 11-12.

ART 34 AMEND

33. The polynucleotide according to any of claims 1-5, wherein said nucleotide sequence is not contained in any of the sequences of the following GenBank Accession Nos: AJ003355, AA806165; A1002979; T33896; U52522; AA779203; C06247; AA707194; AA599596; 5453538; U13369; and J03528.
34. The polypeptide according to any of claims 7-10, the sequence of which is not completely encoded by a polynucleotide sequence contained in any of the sequences of the following GenBank Accession Nos: AJ003355, AA806165; A1002979; T33896; U52522; AA779203; C06247; AA707194; AA599596; 5453538; U13369; and J03528.
35. The polynucleotide according to claim 15, the sequence of which is not contained in any of the sequences of the following GenBank Accession Nos: AJ003355, AA806165; A1002979; T33896; U52522; AA779203; C06247; AA707194; AA599596; 5453538; U13369; and J03528.
36. The isolated polynucleotide of claim 1, wherein the nucleotide sequence is expressed at the mRNA level in Jurkat T cells; and when COS-1 cells expressing TNF receptor are genetically altered to express the sequence, the cells have increased enzymatic activity for cleaving and releasing the receptor.

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Fig. 1



2/5

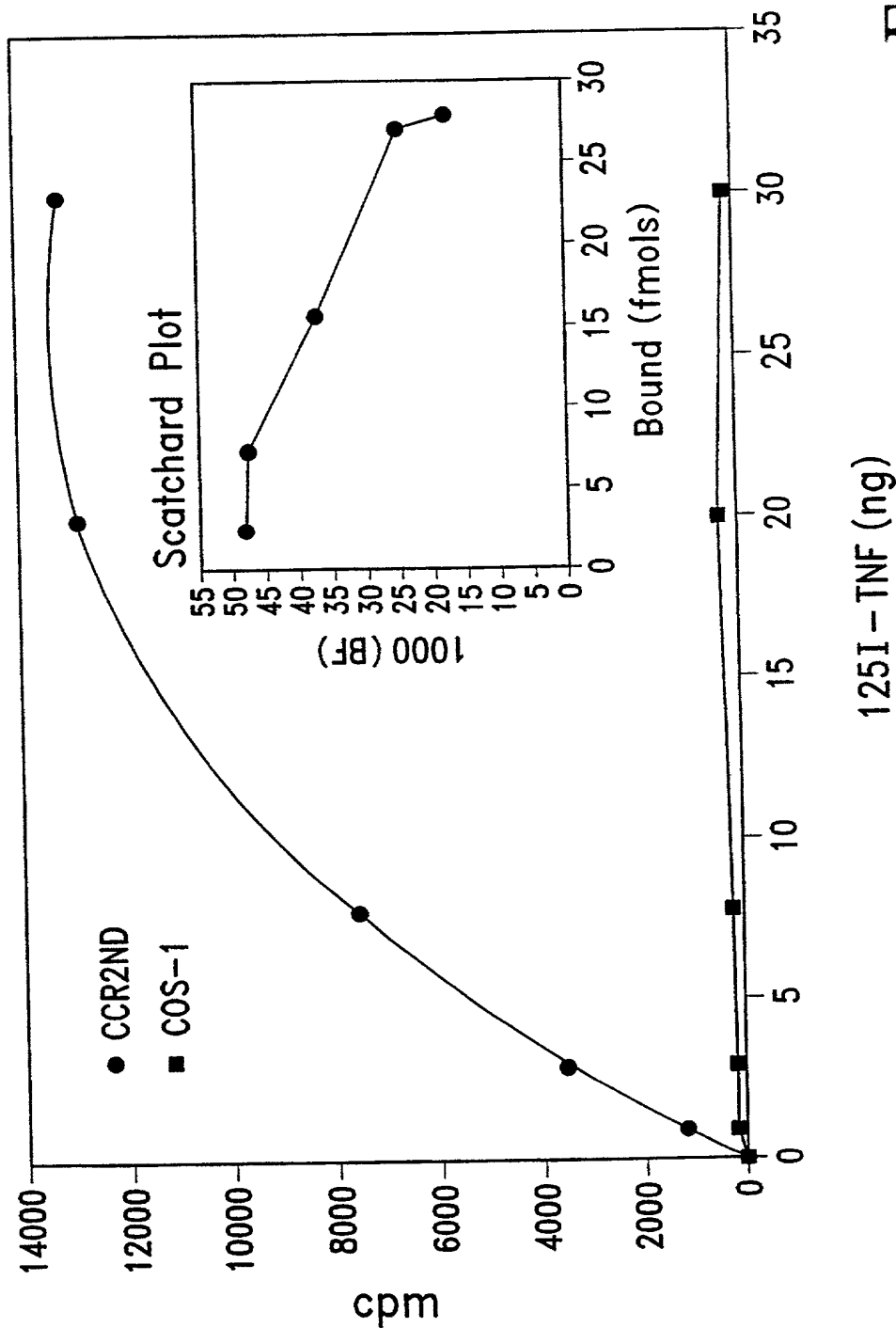


Fig. 2

3/5

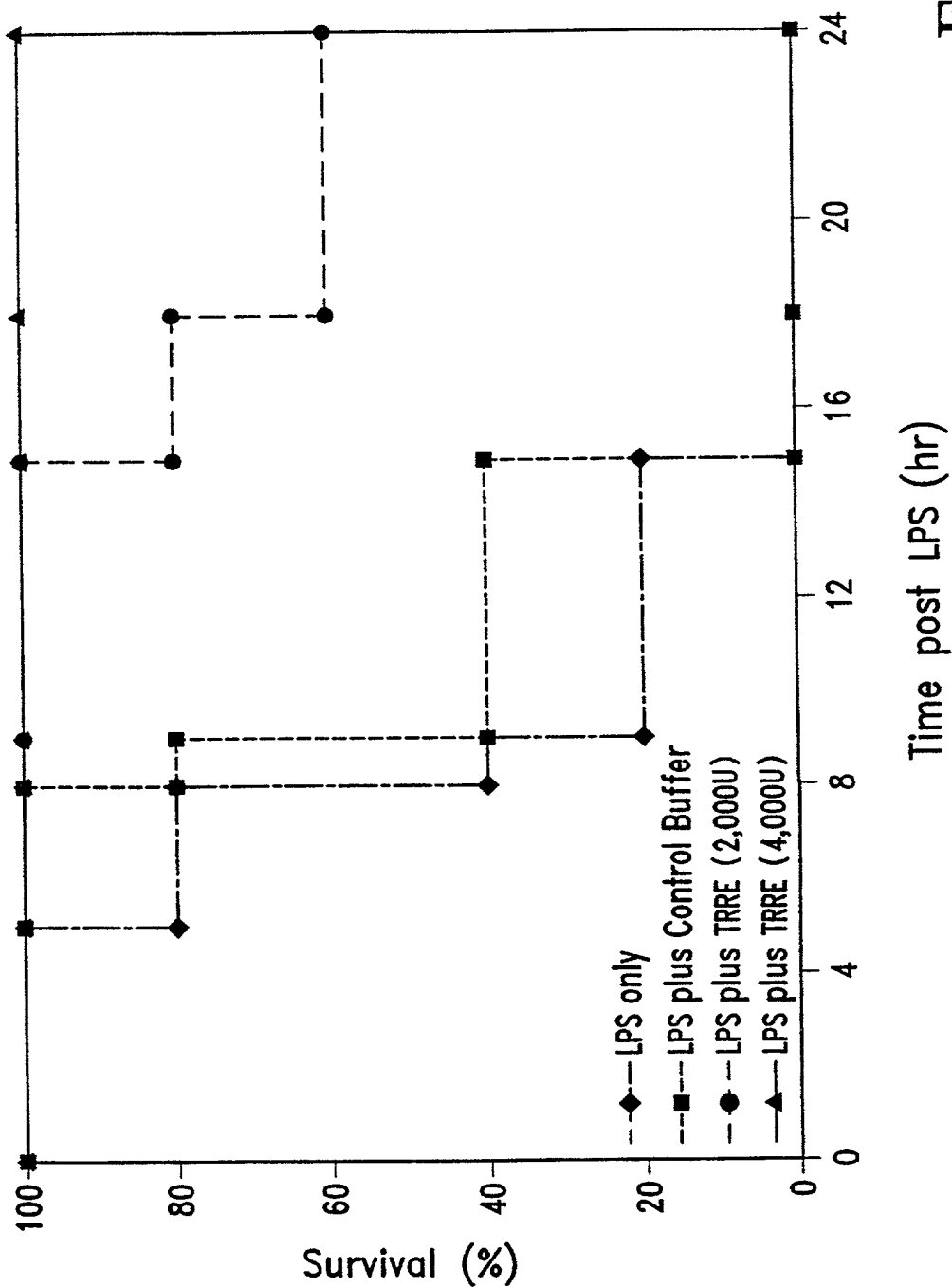
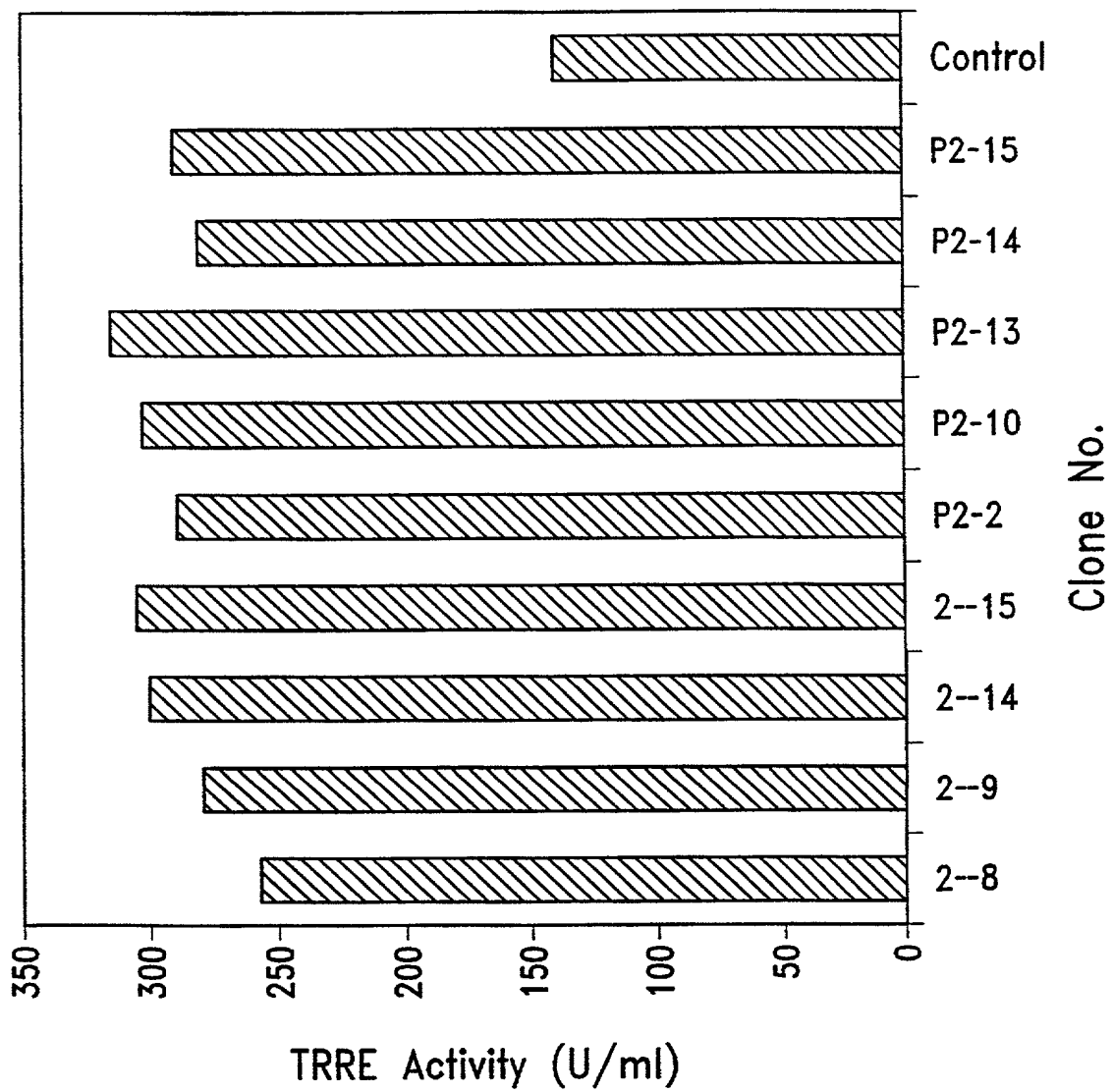


Fig. 3

4/5

Fig. 4



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5/5

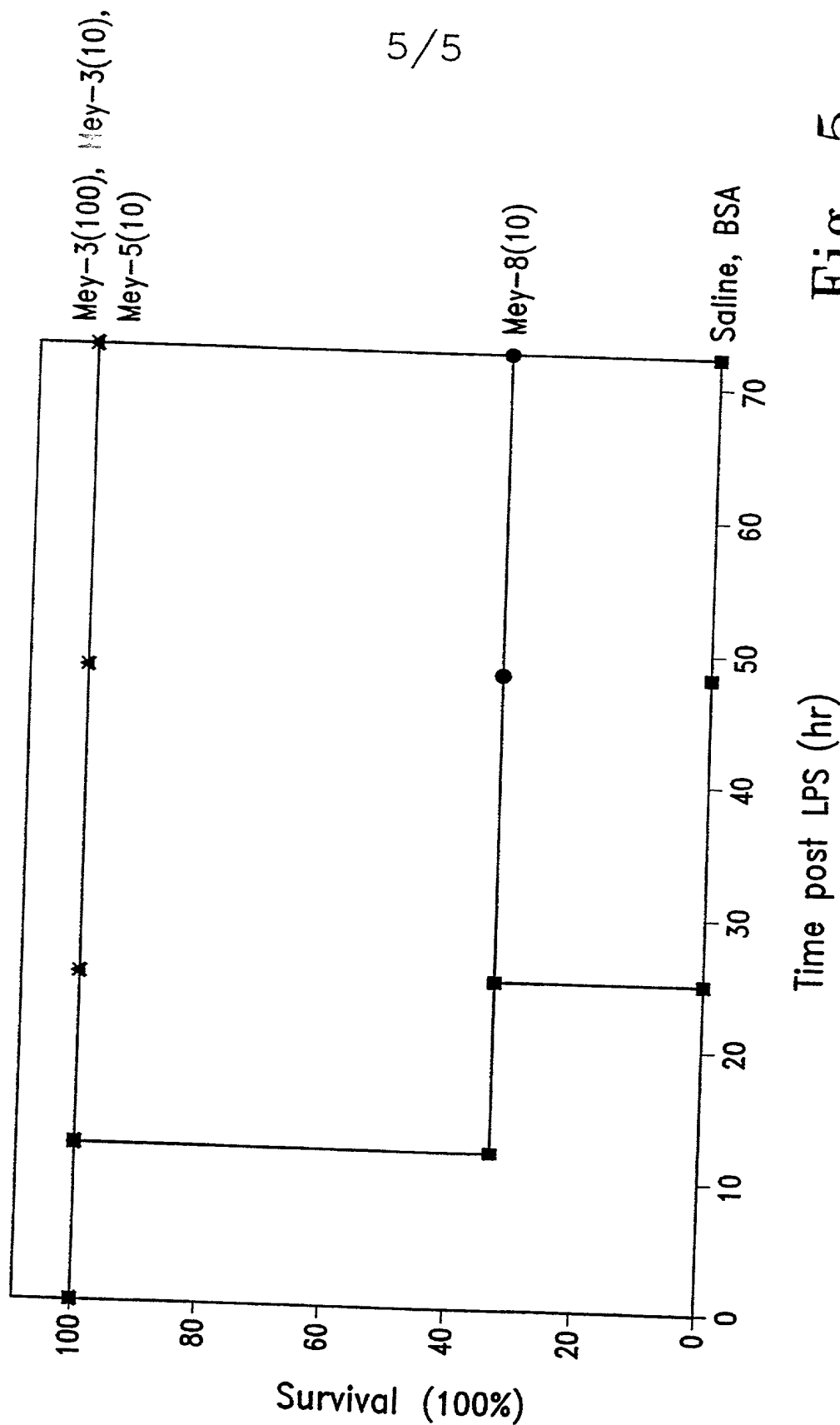


Fig. 5

COMBINED DECLARATION FOR PATENT APPLICATION AND POWER OF ATTORNEY
(Includes Reference to PCT International Applications)

ATTORNEY'S DOCKET NUMBER

IRVN-007CIP2

As a below-named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name,

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

Factors Affecting Tumor Necrosis Factor Receptor Releasing Enzyme Activity ✓

the specification of which (check only one item below):

☐ is attached hereto.

☐ was filed as United States application

Serial No. _____

on _____,

and was amended

on _____ (if applicable).

☒ was filed as PCT international application

Number US99/10793 ✓

on May 14, 1999 ✓

and was amended under PCT Article 19

on January 28, 2000 (if applicable).

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, §1.56(a).

I hereby claim foreign priority benefits under Title 35, United States Code, §119 of any foreign application(s) for patent or inventor's certificate or of any PCT international application(s) designating at least one country other than the United States of America listed below and have also identified below any foreign application(s) for patent or inventor's certificate or any PCT international application(s) designating at least one country other than the United States of America filed by me on the same subject matter having a filing date before that of the application(s) of which priority is claimed:

PRIOR FOREIGN/PCT APPLICATION(S) AND ANY PRIORITY CLAIMS UNDER 35 U.S.C. 119:

COUNTRY (if PCT, indicate "PCT")	APPLICATION NUMBER	DATE OF FILING (day, month, year)	PRIORITY CLAIMED UNDER 35 USC 119
PCT	PCT/US99/10793	14 May 1999	<input checked="" type="checkbox"/> YES <input type="checkbox"/> NO
			<input type="checkbox"/> YES <input type="checkbox"/> NO
			<input type="checkbox"/> YES <input type="checkbox"/> NO

COMBINED DECLARATION FOR PATENT APPLICATION AND POWER OF ATTORNEY
(Includes Reference to PCT International Applications)

ATTORNEY'S DOCKET NUMBER

IRVN-007CIP2

As a below-named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name,

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

Factors Affecting Tumor Necrosis Factor Receptor Releasing Enzyme Activity
the specification of which (check only one item below):

☐ is attached hereto.

☐ was filed as United States application

Serial No. _____

on _____,

and was amended

on _____ (if applicable).

☒ was filed as PCT international application

Number US99/10793

on May 14, 1999,

and was amended under PCT Article 19

on January 28, 2000 (if applicable).

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

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PRIOR FOREIGN/PCT APPLICATION(S) AND ANY PRIORITY CLAIMS UNDER 35 U.S.C. 119:

COUNTRY (if PCT, indicate "PCT")	APPLICATION NUMBER	DATE OF FILING (day, month, year)	PRIORITY CLAIMED UNDER 35 USC 119
PCT	PCT/US99/10793	14 May 1999	<input checked="" type="checkbox"/> YES <input type="checkbox"/> NO
			<input type="checkbox"/> YES <input type="checkbox"/> NO
			<input type="checkbox"/> YES <input type="checkbox"/> NO

NOV-10-00 FRI 04:36 PM BOZICEVIC FIELD&FRANCIS FAX NO. 650 327+3231

P. 04

Combined Declaration For Patent Application and Power of Attorney (Continued) (Includes Reference to PCT International Applications)				ATTORNEY'S DOCKET NUMBER IRVN-007CIP2	
I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) or PCT international application(s) designating the United States of America that is/are listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in that/those prior application(s) in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, §1.56(a) which occurred between the filing date of the prior application(s) and the national or PCT international filing date of this application:					
PRIOR U.S. APPLICATIONS OR PCT INTERNATIONAL APPLICATIONS DESIGNATING THE U.S. FOR BENEFIT UNDER 35 U.S.C. 120:					
U.S. APPLICATIONS			STATUS (Check one)		
U.S. APPLICATION NUMBER	U.S. FILING DATE	PATENTED	PENDING	ABANDONED	
09/081,385 -	May 14, 1998 ✓		XX		
POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith. (List name and registration number)					
<div style="display: flex; justify-content: space-between;"> <div> Karl Bozicevic, Reg. No. 28,807 Bret E. Field, Reg. No. 37,620 Dianne L. DeVore, Reg. No. 42,484 Alan W. Cannon, Reg. No. 34,977 </div> <div> Carol L. Francis, Reg. No. 36,513 Pamela J. Sherwood, Reg. No. 36,677 Paula A. Borden, Reg. No. 42,344 Nicole Verona, Reg. No. P-47,153 </div> </div>					
Send Correspondence to: BOZICEVIC, FIELD & FRANCIS-LLP 200 Middlefield Road, Suite 200 Menlo Park, CA 94025 Telephone: (650) 327-3400 Facsimile: (650) 327-3231				Direct Telephone Calls to: (name and telephone number) Name: <u>Carol L. Francis</u> Registration No. 36,513 Telephone: (650) 327-3400	
1-00	FULL NAME OF INVENTOR	FAMILY NAME	FIRST GIVEN NAME	SECOND GIVEN NAME	
	RESIDENCE & CITIZENSHIP	CITY	STATE OR FOREIGN COUNTRY	COUNTRY OF CITIZENSHIP	
	POST OFFICE ADDRESS	CITY	STATE & ZIP CODE/COUNTRY		
		Gatanaga	Tetsuya		
		Irvine	California CA	Japan ✓	
		77 Wellesley	Irvine	California 92612	
202	FULL NAME OF INVENTOR	FAMILY NAME	FIRST GIVEN NAME	SECOND GIVEN NAME	
	RESIDENCE & CITIZENSHIP	CITY	STATE OR FOREIGN COUNTRY	COUNTRY OF CITIZENSHIP	
	POST OFFICE ADDRESS	CITY	STATE & ZIP CODE/COUNTRY		
		Granger	Gale	A.	
		Laguna Beach	California	United States	
		31562 Santa Rosa	Laguna Beach	California	
I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.					
SIGNATURE OF INVENTOR 201		SIGNATURE OF INVENTOR 202		SIGNATURE OF INVENTOR 203	
DATE 11/13/00		DATE		DATE	

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NOV-13-00 MON 09:19 AM

BOZICEVIC FIELD&FRANCIS

FAX NO. 650 327+3231

P. 08

Combined Declaration For Patent Application and Power of Attorney (Continued) (Includes Reference to PCT International Applications)				ATTORNEY'S DOCKET NUMBER IRVN-007CIP2	
<p>I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) or PCT international application(s) designating the United States of America that is/are listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in that/those prior application(s) in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, §1.56(a) which occurred between the filing date of the prior application(s) and the national or PCT international filing date of this application:</p>					
<p>PRIOR U.S. APPLICATIONS OR PCT INTERNATIONAL APPLICATIONS DESIGNATING THE U.S. FOR BENEFIT UNDER 35 U.S.C. 120:</p>					
U.S. APPLICATIONS			STATUS (Check one)		
U.S. APPLICATION NUMBER	U.S. FILING DATE	PATENTED	PENDING	ABANDONED	
09/081,385	May 14, 1998		XX		
<p>POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith. (List name and registration number)</p>					
<p>Karl Bozicovic, Reg. No. 28,807 Bret E. Field, Reg. No. 37,620 Dianne L. DeVoro, Reg. No. 42,484 Alan W. Cannon, Reg. No. 34,977</p>			<p>Carol L. Francis, Reg. No. 36,513 Pamela J. Sherwood, Reg. No. 36,677 Paula A. Borden, Reg. No. 42,344 Nicole Varano, Reg. No. P-47,153</p>		
<p>Send Correspondence to: BOZICEVIC, FIELD & FRANCIS LLP 200 Middlefield Road, Suite 200 Menlo Park, CA 94025 Telephone: (650) 327-3400 Facsimile: (650) 327-3231</p>				<p>Direct Telephone Calls to: (Name and telephone number) Name: Carol L. Francis Registration No. 36,513 Telephone: (650) 327-3400</p>	
201	FULL NAME OF INVENTOR	FAMILY NAME	FIRST GIVEN NAME	SECOND GIVEN NAME	
	RESIDENCE & CITIZENSHIP	CITY	STATE OR FOREIGN COUNTRY	COUNTRY OF CITIZENSHIP	
	POST OFFICE ADDRESS	CITY	STATE & ZIP CODE/COUNTRY		
		Gatanaga	Tetsuya		
		Irvine	California	Japan	
		77 Wellesley	Irvine	California 92612	
202	FULL NAME OF INVENTOR	FAMILY NAME	FIRST GIVEN NAME	SECOND GIVEN NAME	
	RESIDENCE & CITIZENSHIP	CITY	STATE OR FOREIGN COUNTRY	COUNTRY OF CITIZENSHIP	
	POST OFFICE ADDRESS	CITY	STATE & ZIP CODE/COUNTRY		
		Granger	Gale	A.	
		Laguna Beach	California CA	United States ✓	
		31562 Santa Rosa	Laguna Beach	California	
<p>I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.</p>					
SIGNATURE OF INVENTOR 201		SIGNATURE OF INVENTOR 202		SIGNATURE OF INVENTOR 203	
		Gale A. Granger			
DATE		DATE		DATE	
		13 NOV 00			

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SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: Gatanaga, Tetsuya
Granger, Gale A.
- (ii) TITLE OF INVENTION: Factors Altering Tumor Necrosis
Factor Receptor Releasing Enzyme Activity.
- (iii) NUMBER OF SEQUENCES: 154
- (iv) CORRESPONDENCE ADDRESS:
(A) ADDRESSEE: BOZICEVIC, FIELD, & FRANCIS, LLP
(B) STREET: 200 MIDDLEFIELD ROAD, #200
(C) CITY: Menlo Park
(D) STATE: CA
(E) COUNTRY: USA
(F) ZIP: 94025
- (v) COMPUTER READABLE FORM:
(A) MEDIUM TYPE: Diskette
(B) COMPUTER: IBM Compatible
(C) OPERATING SYSTEM: Windows
(D) SOFTWARE: FastSEQ for Windows Version 2.0b
- (vi) CURRENT APPLICATION DATA:
(A) APPLICATION NUMBER: 09/700,354
(B) FILING DATE: 13-NOV-2000
(C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: 09/081,385
(B) FILING DATE: 14-MAY-1998

(A) APPLICATION NUMBER: PCT/US99/10793
(B) FILING DATE: 14-MAY-1999
- (viii) ATTORNEY/AGENT INFORMATION:
(A) NAME: Francis, Carol L.
(B) REGISTRATION NUMBER: 36,513
(C) REFERENCE/DOCKET NUMBER: IRVN-007CIP2
- (ix) TELECOMMUNICATION INFORMATION:
(A) TELEPHONE: 650-327-3400
(B) TELEFAX: 650-327-3231
(C) TELEX:

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 4047 base pairs
(B) TYPE: nucleic acid

09/700354

(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: Genomic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

AAGCTTTTTG	CTTTCCTTCC	CCGGGAAAGG	CCGGGGCCAG	AGACCCGCAC	TCGGACCAGG	60
CGGGGGCTGC	GGGGCCAGAG	TGGGCTGGGG	AGGGCTGGGA	GGGCGTCTGG	GGCCGGCTCC	120
TCCAGGCTGG	GGGCCGCCAG	CTCCGGGAAG	GCAGTCCTGG	CCTGCGGATG	GGGCCGCGCG	180
TGGGGCCCCG	CGGGGCGGCC	TCGGGAGGCG	TCCAGGCTGC	GGGAGCGGGA	GGAGCGGCCG	240
TGCGGGCGCC	AGCGCCGTGG	GTGGAGGTCG	CCGTCCCTCC	TGAGGGGCAG	CCAGTGCGTT	300
TGGGACCCGG	GAGCAGAGCC	CGCGCCTCCC	CAGCGGCCCTC	CCCGGGGGTC	TCACCGGGTC	360
ACCCGAGAGC	GGAGGCCCGG	GCTCCGCAGA	AACCCGGGGC	GGCCGCGGGG	AAGCAGCGCC	420
CTCAGGCGTC	GGAGGAGCCC	CCAGAAGGAC	CTCGCGCCTT	CCCGCCGGGC	TCCGACCGCC	480
TGGGTTTCGGT	GCGGGACGGC	CCAGGCCGCC	AGGACCCCCA	AGCGCAGCTC	AGTCTGCGGG	540
GCACGACCCA	GAGGCCAGCA	GCAGAGGACG	GGGCCGGGGC	CGGGAGAGGG	CGGGGAGGGC	600
GCTCCTGGGA	GGTCAAGGCC	AGGGCTAGAC	TTTCAGGGTC	ATGGCCTGGC	CCCTCATCCC	660
CAGGGAGGTG	AGGGGGCTCT	GTGAGCAGAG	GGGGCCCCGG	TGGAGAAGGC	GCTGCTAGCC	720
AGGGGCGGGG	CAGGAGCCCA	GGTGGGGACT	TAAGGTGGC	TGAAGGGACC	CTCAGGCTGC	780
AGGGATAGGG	AGGGAAGCTA	GGGGTGTGGC	TTGGGGAGGT	GCTGGGGGAC	CGCGGGCGCC	840
CTTTATTCTG	AAGCCGAATG	TGCTGCCGGA	GTCCCCAGTG	ACCTAGAAAT	CCATTTCAAG	900
ATTTTCAGGA	GTTTCAGGTG	GAGACAAAGG	CCAGGCCCAG	GTGAAAATGT	GGCAGTGACA	960
GAGTATGGGG	TGAGAACCAC	GGAGAGAGGA	AGTCCCCGAG	GCGGATGATG	GGACAGAGAG	1020
CGGGGACCAG	AATTTTTTAA	AACGCATCTG	AGATGCGTTT	GGCAGACTCA	TAGTTGTTTT	1080
CCTTTACCGG	AGAAAGTGTG	GGCAGAAGCC	AGCTCTAAAG	CCCAGGCTGC	CCAGCCTGCA	1140
CTGGCAGAGC	TGACGGAAGG	CCAGGGCAGA	GCCTTCCCTC	CCTGTACACG	ACATGAGCCC	1200
TGGAGATCTG	GAATGAGGCA	GATGTGCCCA	GGGAAAGCTG	ATCCGCCCCG	ACCCAGGGCC	1260
CCCCGGGTGC	CCCTTTGAGC	GTGGAATCGT	TGCCAGGTCA	TGGCTCCCTG	CTATCGAACA	1320
CCGGACACGG	GTCTGTGTCT	GCACCTGGCA	GTTGCAGGAC	CGACACCCAC	AATGCCTTAA	1380
GAGGTGATGA	CTGCCTTCCA	GGGGCCTGGC	TGGCTGACAC	TTTGCATGGC	TCCTGGAGAA	1440
GAGGGATTGA	GTGGAGTCCA	CGGGTCATGG	CCACGTCTCG	GGTGCTGCCT	CTGAGGCAGG	1500
GCCCGGCTGG	GGTGAGAAGG	GGCTGGAGAC	AGGTTCCCTG	CAGTTCAGCC	TCTAACCGGT	1560
GGTCTTCATG	CCTAGGAACC	CACTGGGGGC	TTATGAAACT	GCAGGTGGCT	GAGTCCTTGC	1620
CATGGGGTCT	CTCCTTCAGG	AGGTCTGGGT	GGGGCCGGAG	ACTGTACCCC	ACAAAGGCTC	1680
CCAGGTGAGG	CGGATGTGGC	CTGGCGCTGT	GTGGCTCTGG	ACCTAGTCCT	TGGGCTTGGG	1740
CTGGCGCCCA	GGGCCTGGGC	TTGAGACAGC	TGTGACGCAG	GCAAGCCATT	TACCCCGTTT	1800
GTGGGGACAT	TACATCTTCC	TAGCTTGAA	CACACAGGCA	GCCAGGGTTG	TTATCCACAT	1860
TCCTCCTCCA	TGTTCTTCTC	TTGAGAACTT	TTACCAGGTA	TGTCAGGAGC	TGGGCTCCAC	1920
CAGGGAGACT	CAAGTGGA	GCCCTCATCC	TTGTCTCCA	GGAGACAGGA	AAACCTATGG	1980
TTACAATTCC	AGGGACAAGA	GCGATGCATG	TGAGGTGTGG	CAAATCTCAC	TGTTCAACTG	2040
GAGAAATCAG	AGACAGCTTC	CTGGAGGCAG	TGACACCTGG	ACAGGCTTCT	CCACAGGAGG	2100
AAGCGAGTGA	GAGAAGCCAA	CTGGGATGGA	CCCATCATGT	AGGGGGAACA	GTGCGCGCAG	2160
AACCAACAAC	CACCCCCACC	CTAGGCCCCAG	AGCTCACGGA	GAGAGCTGGG	CCTCTCGGGG	2220
TGACTACATA	GTTCCCTGCT	GGATCTTAGG	TCTTGTCTCT	GGGCAGCTCT	GCTGAGACCT	2280
CTATGCCTGT	TCCAGGCTGC	ACCAAGGTTT	TGTGACTATT	GGTCTGGGGT	TGTTTTGCAG	2340
CAACTGAAGT	GTTCTGTTGT	AAAACAGGCA	CTTGATTGTC	TGGAAGGAAT	GCTGTTTGT	2400
CTTGCTGCGA	CAACACTTGA	GCAGCATTTA	GTGGGCGGTT	TATATCTTGT	GGAGTAATGG	2460
TGTTTTTTGA	AGTCTGTCTC	GGGTACTGCA	CATTAAGAGG	AATATCATT	TCTGAAACAT	2520
TGCTATTTTC	CACACCAGAA	ATCATATCCT	CTTGCTGGTC	CATGTCTGAA	GACCTTACAC	2580
GAGAAAGTCT	TAATGTAAGT	TTAGTAGAGT	CCTTGATGG	AGAACTAATT	ATATCATACA	2640
TTGCCGCTTT	CTCACTCTGC	TCTTTTTTCAT	CCTTGCCTAA	TTTCATTTTC	TTCTGCTTCT	2700
TTTGTTTTCT	TTCTGGAGAA	TCTAGCAAGA	TATCTGGTGG	AACATCTCGA	GGTGATGAAC	2760
AAGGTAGAGA	CTGAGATTGT	AGGATTAAAG	GTGGTCTTGA	GCCTTTAGGA	GTTCTTTCAC	2820
TTCCAGCAGG	GGAGCATACT	GGCTGTGGAG	ATCTCAAGGG	AAAAGATGCA	GCATTCCTCA	2880
TTGTTGAAGA	ATCTCCATCG	TCACTACTTA	GCCTGTGCAC	CATGTGTAGG	TAGTCCTCAC	2940
TTGAACCATG	TCTAGGATTA	TCAGCATGAT	GATTAGCTGA	ATTGCCAGAC	AACGGACCAG	3000

AAACTTTATT	ATCATGTATG	TTTCTCAAAC	CACCTGCAAC	AATGGGACTT	GATACCGATG	3060
CTTGTTGCAT	CTGTGGATGT	GTTGTGTAAC	TTGAAGGATG	GGAATATGGC	ATGTATCCTG	3120
CAGGGCTTTG	TGGGGCGTAT	GGACTAGGCA	CTGGGCTATT	TTGCTGTGGC	ATAAATCTGT	3180
TCCCAGAGCT	TGCTCTGTGGT	GGCACAAACC	GGCTGGAGGG	GCTATGTGAG	ATAGTGGTTT	3240
GTTGATAAAT	GGAAGATGCA	GGACTACTGT	GCATGGAATT	CTGAGAAAGT	TTATACTGAG	3300
ACATCATCAT	TCCACTTTGT	ACATATCTGT	TCTGCATGCT	TTTCTCCCTG	AAAACATTAG	3360
GACTCCTTGC	CAGGACGGCC	TGCAACAAGA	CTGGTATGTC	ACCTTCTGGG	TCATCACTGC	3420
CAAGGTTATC	TTTCAACTCT	ATGTGATCTG	TTGATACCTG	GTTGAGGCTA	TGGACAAGCT	3480
GTGAAACCAA	ATTGTCATCC	CTACAAGCCA	AAAGGCAGTT	CACCTCTTCT	GCTATTTCGTG	3540
CATTAAAGAG	AAGGCTCTTT	GTAGTTGTAG	CAGGTAAAGG	AGATGGAAGA	GGCAGCTGGT	3600
TCAGGAGGTC	TGTGAGACTA	GCAATCCCCG	CAAGAGTAGT	AATGGGGACA	TGGGGCATAT	3660
CCCCATTTCAT	CCTGAATTTT	TGGAATGGTG	TTGCCTATAA	AAGTACTTAG	TTCAGGTGCC	3720
AGCTGTCAAT	ACTTCCCATT	TCCCAAACAC	TGGGCGAATC	GGCGTCTGAA	TCCAAGGGGA	3780
GGCCGAGGCC	GCTGTGGCGA	GAGACTATAA	TCCGGGCCGG	GAGGGGGGGC	GGCTACGGCT	3840
CCTCTTCCGT	CTCCTCAGTG	CGGGGAACAT	GTAGAGCCGG	GGGGAGACCA	GCCGAGAAGA	3900
CAAATCGTTG	CTTCTTCTTC	CTCCTCCTCC	TCCTTCTCCC	ACATAGAAAC	ACTCACAAAC	3960
ACCCGACCAC	GGGCCCCGAGC	TACCGGGGGG	GCATCGCCGC	GGGCCCCGGA	ACCAATTCTC	4020
CTGTGCGCGG	GGGCGTCCTT	TGGATCC				4047

CCAGGCTCTC	GAAATCCCTA	TAGTCCAGCC	CCTCCCAGACA	TGCATAGAGG	CACCTCGATGA	2100
CCTCGCGGCT	CTCCAGCGCA	CGTGAAGCGCA	CGCTGAAACC	AGCCAGGTAG	CCATGGAAGT	2160
AGTGGTGGAT	CGACAAAGGG	TCTCCTTGGG	TGGTGTCTGT	ACTGTTGTCT	CCCTTTTCCT	2220
TCTCTTTGTT	CTTCTCCTCA	GTCCAGCAGG	CCCCAATCAT	GAGAGCAGGC	TCCCTTCGGG	2280
GTGGGTGGAT	GAGGCCATTG	TCATGGATGA	GGGCAGGGTC	GAAGGAGATG	CCGTCGGTAT	2340
AGAGTGTGAC	TGTGGGGAAC	TCGAGGTTCA	GAGCGTAGTG	GTGCCACTCA	TCATCACAGA	2400
CCTGCTCCAG	CTTCCAGAGG	AACTTGACTG	GGCGGGCACT	CTCAAGCAGG	GGCCAGTAGA	2460
GGAAGGCAAT	CCTACAGCCG	TGGACAGTCA	GCGAGTAGTG	AGAGAAGCCG	TCCTCATTCT	2520
GGACAGTGTT	ACATACGATG	GTTTCCTCTT	CCTTCTTGCC	CTTGTTGGGA	GTTACGCCAT	2580
GCTTCATCCA	GAAGGACAGG	GTGAAGTGGT	CACTGAGGCT	GTCCTGGGGC	CCAGAGCCCCA	2640
GCCCACTGGG	GCCACCCAGG	GGCACCTGCA	CAGCCTGGGT	GCCATTGAAC	CAGTAGATCA	2700
GGCTGCTGTC	CTGGCTGTAG	TGCACCGAGA	GTCTCTGCTG	CCAGTTGGCA	TTGGGGCCAG	2760
GCATGGGCAA	CAGATCCACT	TCCCCAGTGG	CAGCACCACA	GAGTTTCCGC	AGCGCCCCT	2820
CTGAGTAGTT	GTCACGGTCA	CAGCCCTTGG	CCACATGGCT	GGTCTGCAGC	TCTATGGTGG	2880
CCTGAATGTT	CCAGAGTGGT	TCATCACAGG	TCTCCAGGCG	GATACCAGGG	AACAAGGCCA	2940
AGCTCCCAGC	ACCTGGTGCA	TATTCGATCC	TTTTGTTCCA	GCCTTGCCAG	CTGGGTTTAC	3000
AGGTGGGCTT	CACCTGAATC	TCCACCTCAG	CATCATCTGC	TGCCCCGCTT	TTCCCACAGT	3060
CATAAGCTGT	CACGTGAAAC	TTATAGAGCC	TCTCACCCT	GTACTGCAGC	TTCTCTGTGT	3120
TCTCAATGTT	CCCTGCATTG	TCAATGAGGA	AAGGGGTGTT	GGGTGTGAGA	ATCTCATAGT	3180
AGCAGATCTG	GCTGTACTGG	GGGGAGCAGT	CACCGTCAAT	GGCTTCCACC	CGCAGGATGC	3240
GATCGTACAG	CTTCCCCTCT	GTCACAGCCG	CACGATACAG	CCGTTCCACA	AACACTGGGG	3300
CAAACTCGTT	CACATCGTTG	ACCCGCACAT	GCACAGTGGC	CTTGTGGGAC	TTCTTGGTGT	3360
TGGCCCCGTC	GGGGCCCTCG	CCACAGTCAT	AGGCCTGGAT	GGTGAAGGTG	TGTTCTTTCT	3420
GGGCCTCGCA	GTCCACAGGC	TCCTTGCCCC	GGATCAGCCC	CTCTCCTGTC	GCCTTGTCAA	3480
GGATCACAGC	CTCAAAGGGC	ACCCAGACC	CATGGAGCCG	GAAGCCGCAG	ATCTCACCTG	3540
CATAGCGCAG	CGGGGCATCC	TTGTCCAAGG	CAAAGAGTGG	TGGATTTCAGT	AGGACCGTGT	3600
TGTCATTCTC	CATGACGATG	CCCTGGTACT	CTGCCTCAAT	CCATGGCTTG	TGCTTGTTGG	3660
CTTTGTTACA	GGAGCAGGAC	GCGAGCAGAG	AGGCCAGCAG	AAGGGGCAGC	AGCAGGAGGG	3720
TCATGGTGCG	GCGTGGGGCA	GGGCAGGGCC	AGGCGTTTGC	CTCCCCTGGG	AGCCTCCAGC	3780
CTGCGGATTC	CACCTTGCGG	GAGGGATACA	GGGGGGGAAA	ACCAAATAAA	AACGCTCAAAT	3840
AAATTGTGTA	GGAGGAGTCC	AGCTTAGGAC	CGGGCCAGAG	CCAGGCCAGG	CTCGGGGAGG	3900
GGGCCTCTGC	AGGTTCCAGG	GATCACTGCT	GCCACCACCG	CCACCCTGGG	AGCCAGTTAT	3960
TTTGCCATTG	CCTTGATTGC	AACAGTGCC	TCTCTGTCA	TGGCAGACAG	CACCGTGATC	4020
AGGATCTCTT	CTCCACAGTC	GTACTTCTGC	TCAATCTCCT	TGCCAAGGTC	TCCCTCAGGG	4080
AGACGAAGGT	CCTCTCGTAC	CTCCCCGCTG	TCCTGGAGCA	GTGATAGGTA	CCCATCCTGG	4140
ATCTTTGGAT	CC					4152

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 3117 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: Genomic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

GGATCCAAAG	ATTCGGCACG	AGTGGCCACA	TCATGAACCT	CCAGGCCCAG	CCCAAGGCTC	60
AGAACAAGCG	GAAGCGTTGC	CTCTTTGGGG	GCCAGGAACC	AGCTCCCAAG	GAGCAGCCCC	120
CTCCCCTGCA	GCCCCCCCAG	CAGTGCATCA	GAGTGAAGGA	GGAGCAGTAC	CTCGGGCACG	180
AGGGTCCAGG	AGGGCCAGTC	TCCACCTCTC	AGCCTGTGGA	ACTGCCCCCT	CCTAGCAGCC	240
TGGCCCTGCT	GAACTCTGTG	GTGTATGGGC	CTGAGCGGAC	CTCAGCAGCC	ATGCTGTCCC	300
AGCAGGTGGC	CTCAGTAAAG	TGGCCCCAAT	CTGTGATGGC	TCCAGGGCGG	GGCCCGGAGC	360
GTGGAGGAGG	TGGGGGTGTC	AGTGACAGCA	GCTGGCAGCA	GCAGCCAGGC	CAGCCTCCAC	420
CCCATTCAAC	ATGGAAGTGC	CACAGTCTGT	CCCTCTACAG	TGCAACCAAG	GGGAGCCCGC	480

ATCCTGGAGT	GGGAGTCCCC	ACTTACTATA	ACCACCCTGA	GGCACTGAAG	CGGGAGAAAG	540
CGGGGGGCCC	ACAGCTGGAC	CGCTATGTGC	GACCAATGAT	GCCACAGAAG	GTGCAGCTGG	600
AGGTAGGGCG	GCCCCAGGCA	CCCTTGAATT	CTTTCCACGC	AGCCAAGAAA	CCCCCAAACC	660
AGTCACTGCC	CCTGCAACCC	TTCCAGCTGG	CATTTCGGCCA	CCAGGTGAAC	CGGCAGGTCT	720
TCCGGCAGGG	CCCACCGCCC	CCAAACCCGG	TGGCTGCCTT	CCCTCCACAG	AAGCAGCAGC	780
AGCAGCAGCA	ACCACAGCAG	CAGCAGCAGC	AGCAGCAGGC	AGCCCTACCC	CAGATGCCGC	840
TCTTTGAGAA	CTTCTATTCC	ATGCCACAGC	AACCCTCGCA	GCAACCCCAG	GACTTTGGCC	900
TGCAGCCAGC	TGGGCCACTG	GGACAGTCCC	ACCTGGCTCA	CCACAGCATG	GCACCCTACC	960
CCTTCCCCCC	CAACCCAGAT	ATGAACCCAG	AACTGCGCAA	GGCCCTTCTG	CAGGACTCAG	1020
CCCCGCAGCC	AGCGCTACCT	CAGGTCCAGA	TCCCCTTCCC	CCGCCGCTCC	CGCCGCCTCT	1080
CTAAGGAGGG	TATCCTGCCT	CCCAGCGCCC	TGGATGGGGC	TGGCACCCAG	CCTGGGCAGG	1140
AGGCCACTGG	CAACCTGTTC	CTACATCACT	GGCCCCCTGCA	GCAGCCGCCA	CCTGGCTCCC	1200
TGGGGCAGCC	CCATCCTGAA	GCTCTGGGAT	TCCCGCTGGA	GCTGAGGGAG	TCGCAGCTAC	1260
TGCCTGATGG	GGAGAGACTA	GCACCCAATG	GCCGGGAGCG	AGAGGCTCCT	GCCATGGGCA	1320
GCGAGGAGGG	CATGAGGGCA	GTGAGCACAG	GGGACTGTGG	GCAGGTGCTA	CGGGGCGGAG	1380
TGATCCAGAG	CACGCGACGG	AGGCGCCGGG	CATCCCAGGA	GGCCAATTTG	CTGACCCTGG	1440
CCCAGAAGGC	TGTGGAGCTG	GCCTCACTGC	AGAATGCAAA	GGATGGCAGT	GGTTCTGAAG	1500
AGAAGCGGAA	AAGTGTATTG	GCCTCAACTA	CCAAGTGTGG	GGTGGAGTTT	TCTGAGCCTT	1560
CCTTAGCCAC	CAAGCGAGCA	CGAGAAGACA	GTGGGATGGT	ACCCCTCATC	ATCCCACTGT	1620
CTGTGCCTGT	GCGAACTGTG	GACCCAACCTG	AGGCAGCCCC	GGCTGGAGGT	CTTGATGAGG	1680
ACGGGAAGGG	TCTTGAACAG	AACCCTGCTG	AGCAACAAGC	ATCAGTCATC	GTCAACCGCA	1740
GGCGGTCCAC	CCGAATCCCC	GGGACAGATG	CTCAAGCTCA	GGCGGAGGAC	ATGAATGTCA	1800
AGTTGGAGGG	GGAGCCTTCC	GTGCGGAAAC	CAAAGCAGCG	GCCCAGGCCC	GAGCCCCCTCA	1860
TCATCCCCAC	CAAGCGTGGG	ACTTTCATCG	CCCCTCCCGT	CTACTCCAAC	ATCACCCCAT	1920
ACCAGAGCCA	CCTGCGCTCT	CCCGTGCGCC	TAGCTGACCA	CCCCCTCTGAG	CGGAGCTTTG	1980
AGCTACCTCC	CTACACGCCG	CCCCCCATCC	TCAGCCCTGT	GCGGGAAGGC	TCTGGCCTCT	2040
ACTTCAATGC	CATCATATCA	ACCAGCACCA	TCCCTGCCCC	TCCTCCCATC	ACGCCTAAGA	2100
GTGCCCATCG	CACGCTGCTC	CGGACTAACA	GTGCTGAAGT	AACCCCGCCT	GTCTCTCTCTG	2160
TGATGGGGGA	GGCCACCCCA	GTGAGCATCG	AGCCACGGAT	CAACGTGGGC	TCCCGGTTCC	2220
AGGCAGAAAT	CCCCTTGATG	AGGGACCGTG	CCCTGGCAGC	TGCAGATCCC	CACAAGGCTG	2280
ACTTGGTGTG	GCAGCCATGG	GAGGACCTAG	AGAGCAGCCG	GGAGAAGCAG	AGGCAAGTGG	2340
AAGACCTGCT	GACAGCCGCC	TGCTCCAGCA	TTTTCCCTGG	TGCTGGCACC	AACCAGGAGC	2400
TGGCCCTGCA	CTGTCTGCAC	GAATCCAGAG	GAGACATCCT	GGAAACGCTG	AATAAGCTGC	2460
TGCTGAAGAA	GCCCCTGCGG	CCCCACAACC	ATCCGCTGGC	AACTTATCAC	TACACAGGCT	2520
CTGACCAGTG	GAAGATGGCC	GAGAGGAAGC	TGTTCAACAA	AGGCATTGCC	ATCTACAAGA	2580
AGGATTTCTT	CCTGGTGCAG	AAGCTGATCC	AGACCAAGAC	CGTGGCCCCG	TGCGTGGAGT	2640
TCTACTACAC	CTACAAGAAG	CAGGTGAAAA	TCGGCCGCAA	TGGGACTCTA	ACCTTTGGGG	2700
ATGTGGATAC	GAGCGATGAG	AAGTCGGCCC	AGGAAGAGGT	TGAAGTGGAT	ATTAAGACTT	2760
CCCAAAAGTT	CCCAAGGGTG	CCTCTTCCCA	GAAGAGAGTC	CCCAAGTGAA	GAGAGGCTGG	2820
AGCCCAAGAG	GGAGGTGAAG	GAGCCCCAGGA	AGGAGGGGGA	GGAGGAGGTG	CCAGAGATCC	2880
AAGAGCAAGGA	AGGAGCAGAA	GAGGGGCGAG	AGCGCAGCAG	GCGGGCAGCG	GCAGTCAAAG	2940
CCACGCAGAC	ACTACAGGCC	AATGAGTCGG	CCAGTGACAT	CCTCATCCTC	CGGAGCCACG	3000
AGTCCAACGC	CCCTGGGTCT	GCCGGTGGCC	AGGCCTCGGA	GAAGCCAAGG	GAAGGGACAG	3060
GGAAGTCACG	AAGGGCACTA	CCTTTTTTCAG	AAAAAAAAAA	AAAAAAACAA	AAAGCTT	3117

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 3306 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: Genomic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

GAATTCGGCA	CGAGGTCAGT	TTCTGTGGA	ACACAGAGGC	TGCTGTCCC	ATTCAGACAA	60
CGACGGATAC	AGACCAAGCT	TGCTCTATAA	GGGATCCCAA	CAGTGGATTT	GTGTTTAATC	120
TTAATCCGCT	AAACAGTTCG	CAAGGATATA	ACGTCTCTGG	CATTGGGAAG	ATTTTTATGT	180
TTAATGTCG	CGGCACAATG	CCTGTCTGTG	GGACCATCCT	GGGAAAACCT	GCTTCTGGCT	240
GTGAGGCAGA	AACCCAAACT	GAAGAGCTCA	AGAATTGGAA	GCCAGCAAGG	CCAGTCGGAA	300
TTGAGAAAAG	CCTCCAGCTG	TCCACAGAGG	GCTTCATCAC	TCTGACCTAC	AAAGGGCCTC	360
TCTCTGCCAA	AGGTACCGCT	GATGCTTTTA	TCGTCCGCTT	TGTTTGCAAT	GATGATGTTT	420
ACTCAGGGCC	CCTCAAATTC	CTGCATCAAG	ATATCGACTC	TGGGCAAGGG	ATCCGAAACA	480
CTTACTTTGA	GTTTGAAACC	GCGTTGGCCT	GTGTTCCCTC	TCCAGTGGAC	TGCCAAGTCA	540
CCGACCTGGC	TGGAAATGAG	TACGACCTGA	CTGGCCTAAG	CACAGTCAGG	AAACCTTGA	600
CGGCTGTTGA	CACCTCTGTC	GATGGGAGAA	AGAGGACTTT	CTATTTGAGC	GTTTGCAATC	660
CTCTCCCTTA	CATTCTTGGA	TGCCAGGGCA	GCGCAGTGGG	GTCTTGCTTA	GTGTCAGAAG	720
GCAATAGCTG	GAATCTGGGT	GTGGTGCAGA	TGAGTCCCCA	AGCCGCGGCG	AATGGATCTT	780
TGAGCATCAT	GTATGTCAAC	GGTGACAAGT	GTGGGAACCA	GCGCTTCTCC	ACCAGGATCA	840
CGTTTGAGTG	TGCTCAGATA	TCGGGCTCAC	CAGCATTTCA	GCTTCAGGAT	GGTTGTGAGT	900
ACGTGTTTTAT	CTGGAGAACT	GTGGAAGCCT	GTCCCCTTGT	CAGAGTGGAA	GGGGACAACT	960
GTGAGGTGAA	AGACCCAAGG	CATGGCAACT	TGTATGACCT	GAAGCCCCTG	GGCCTCAACG	1020
ACACCATCGT	GAGCGCTGGC	GAATACACTT	ATTACTTCCG	GGTCTGTGGG	AAGCTTTCCT	1080
CAGACGTCG	CCCCACAAGT	GACAAGTCCA	AGGTGGTCTC	CTCATGTGAG	GAAGAGCGGG	1140
AACCGCAGGG	ATTTACAAAA	GTGGCAGGTC	TCTGACTCA	GAAGCTAACT	TATGAAAATG	1200
GCTTGTTAAA	AATGAACCTC	ACCGGGGGGG	ACACTTGCCA	TAAGGTTTTAT	CAGCGCTCCA	1260
CAGCCATCTT	CTTCTACTGT	AGCCGCGGCA	CCCAGCGGCC	AGTATTTCTA	AAGGAGACTT	1320
CAGATTGTTT	CTACTTGTTT	GAGTGGCGAA	CGCAGTATGC	CTGCCCACCT	TTCGATCTGA	1380
CTGAATGTTT	ATTCAAAGAT	GGGGCTGGCA	ACTCCTTCGA	CCTCTCGTCC	CTGTCAAGGT	1440
ACAGTGACAA	CTGGGAAGCC	ATCACTGGGA	CGGGGGACCC	GGAGCACTAC	CTCATCAATG	1500
TCTGCAAGTC	TCTGGCCCCG	CAGGCTGGCA	CTGAGCCGTG	CCCTCCAGAA	GCAGCCGCGT	1560
GTCTGCTGGG	TGGCTCCAAG	CCCGTGAACC	TCGGCAGGGT	AAGGGACGGA	CCTCAGTGGA	1620
GAGATGGCAT	AATTGTCTCT	AAATACGTTG	ATGGCGACTT	ATGTCCAGAT	GGGATTCGGA	1680
AAAAGTCAAC	CACCATCCGA	TTCACCTGCA	GCGAGAGCCA	AGTGAACTCC	AGGCCCATGT	1740
TCATCAGCGC	CGTGGAGGAC	TGTGAGTACA	CCTTTGCCTG	GCCCACAGCC	ACAGCCTGTC	1800
CCATGAAGAG	CAACGAGCAT	GATGACTGCC	AGGTACACAA	CCCAAGCACA	GGACACCTGT	1860
TTGATCTGAG	CTCCTTAAGT	GGCAGGGCGG	GATTCACAGC	TGCTTACAGC	GAGAAGGGGT	1920
TGGTTTACAT	GAGCATCTGT	GGGGAGAAAT	AAAAGTCCCC	TCCTGGCGTG	GGGGCTGCT	1980
TTGGACAGAC	CAGGATTAGC	GTGGGCAAGG	CCAACAAGAG	GCTGAGATAC	GTGGACCAGG	2040
TCCTGCAGCT	GGTGTACAAG	GATGGGTCCC	CTGTCCCTC	CAAAATCCGC	CTGAGCTATA	2100
AGAGTGTGAT	CAGTTTCGTG	TGCAGGCTGT	AGGCCGGGCC	AACCAATAGG	CCCATGCTCA	2160
TCTCCCTGA	AAGTAGACA	TGCATCTCT	TCTTCTCCTG	GCACACGCCG	CTGGCCTGCG	2220
AGCAAGCGAC	CGAAGGTTCC	GTGAGGAATG	GAAGCTCTAT	TGTTGACTTG	TCTCCCCTTA	2280
TTCATCGCAC	TGGTGGTTAT	GAGGCTTATG	ATGAGAGTGA	GGATGATGCC	TCCGATACCA	2340
ACCCTGATTT	CTACATCAAT	ATTTGTCAGC	CACTAAATCC	CATGCACGGA	GTGCCCTGTC	2400
CTGCCGGAGC	CGCTGTGTGC	AAAGTTCCTA	TTGATGGTCC	CCCCATAGAT	ATCGGCCGGG	2460
TAGCAGGACC	ACCAATACTC	AATCCAATAG	CAAATGAGAT	TACTTTGAAT	TTTGAAAGCA	2520
GTA CTCTTG	CCAGGAATTC	AGTTGTAAAT	AAAATTGAAC	CTGCTCAACA	GCTGAGGGAG	2580
ACTAGAAATG	ATGGGTCCAT	ATCCTGGTGC	ATTGTCATAC	AATTCAAACA	ATGGTGCAGC	2640
TACCAGCTTG	TAATTTTTTAG	GGACTGCAAA	CAAGGCTTTT	TCTTGAAGCT	GAACCAGAAA	2700
CAACTTCTTA	TGTTCTTTAG	GCTTTGTAAT	ATGTGCAGGA	ATATATGGAT	ACTGAGGAGG	2760
TTCAAAATTT	GGTCTCCACC	AGTTACCAAT	GCAATCGTCA	ATGACCCAGT	CTTGCAAAAC	2820
TCCATCCTGA	CGACCCAGTA	TCTCTGTCAT	TAAGCGTTTT	AGTCCTTCAA	CTTCATCTTC	2880
TCCTGGGTTA	AGTTACCAC	CAGGTAGTTT	GAAGAAAGTT	GTTCACAGTC	GCAGCAGTAA	2940
CACATGGGGT	AGCCGGTGCT	CATGTACAAT	CAGAACCCCT	TCTACAGTCC	TCCTCATTTCC	3000
AATTTTATCA	AATTCTTTCC	TCATAGCGTG	AAATCTGGCT	GCAACAGAGC	TGTCCTTTCTC	3060
GTAGAGGGC	TCTTTTGTAC	CAAAAGGATA	ATTGGTAAGA	GGGTACAGGT	TGATGGTGCG	3120
CTCAGGGGTG	AGGGGCTTCG	TCTGCTGGAT	GTACTTGTTG	CCGAACTGAG	TGACCCCCCG	3180
GGCCAGCCG	GTCTGCGAGC	GATTGGGCGG	TACCACAGAC	ATGCTGGCGA	GCTCCGGCGC	3240
TGACGGCGAG	CAGAAAGTGG	CAGGCAGGGT	AGACTTTCCC	CGTGCGGGAA	GCCTCGTGCC	3300
GAATTC						3306

- 8 -

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 4218 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: Genomic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

GAATTCGGCA	CGAGAATGGA	TCAACCTCAA	CAACACGTTA	AAGCTAGACG	AAAGAAGTAA	60
TACACAGTGT	ATGAGTCTCA	CATGAAATAC	CCGGATGTAA	ATCCAAAGAA	ACAGGAAGCA	120
GATTGGTGGT	TGCCAGGGAC	AAGGGCGGTG	GGAGGAGAAA	ATGGAGAGTA	ACGGGACTTT	180
ACTTTTGGAG	TGATGAGAAT	GTTTTGGAGC	TAGATAGAAG	TGGTGGTTGT	ACACCATTGT	240
GGATGTACTA	CCACTTAATT	GTTCACTTAA	AAAGTTAATT	TATGTGAATT	GCATCTTAAT	300
TAAAAACAAG	GATAACATTC	CAACTCCTGG	ACATTATCCT	TCCTTTCCAT	TTGATGTCAG	360
GCCCGTGTTA	GAATTCTCAT	CCGTTTGGT	CACCTGCACTT	AAGATGTGGA	GAAATTAGGA	420
CGCACAGTTA	AGAGGAAGGA	TAACACTGAT	TAAGGTAGTG	CTTTTCTAGG	TTTCCCTTAA	480
ACAATTTAAC	AGATGGATAG	TGGCACCCT	TACGAGATGG	AAAAACCAGC	GGAAGGAAGA	540
TTTGGGGGAG	AAGTTAAGTT	TGTCTTGGGC	CTGTGTTTTG	CAACCTGACT	GTAAAAGACA	600
TATGTTAAGT	CTTCAGTGGC	GAAACACTAA	AACTAGAAAT	GGATCAGAAT	TTTATCTTTG	660
GATGTGACTT	CTCAAGGATG	GTCTTGTAC	TTCAGTGCCT	GGTCAAATGA	CAAGATGGGC	720
AATCTTTTCC	TGAAGGTCCA	AGCACCTGAA	CGTGGCAGGG	TGACCCGATT	CCGATTTGCT	780
TAGAACAATC	CTAGTTCATG	CCTATTGTCC	CTCATGTAAT	TAATATCACT	CTCAAAATGT	840
CTCATTTTGT	GCAATAAATT	CTGCAACGTG	ATGGCGCGAC	TCTCGCGGCC	CGAGCGGCCG	900
GACCTTGTCT	TCGAGGAAGA	GGACCTCCCC	TATGAGGAGG	AAATCATGCG	GAACCAATTC	960
TCTGTCAAAT	GCTGGCTTCA	CTACATCGAG	TTCAAACAGG	GCGCCCCGAA	GCCCAGGCTC	1020
AATCAGCTAT	ACGAGCGGGC	ACTCAAGCTG	CTGCCCTGCA	GCTACAAACT	CTGGTACCGA	1080
TACCTGAAGG	CGCGTCGGGC	ACAGGTGAAG	CATCGCTGTG	TGACCGACCC	TGCCTATGAA	1140
GATGTCAACA	ACTGTCATGA	GAGGGCCTTT	GTGTTTCATG	ACAAGATGCC	TCGTCTGTGG	1200
CTAGATTACT	GCCAGTTTCT	CATGGACGAC	GGGCGCGTCA	CACACACCCG	CCGCACCTTC	1260
GACCGTGCCC	TCCGGGCAC	GCCCATCACG	CAGCACTCTC	GAATTTGGCC	CCTGTATCTG	1320
CGCTTCCTGC	GCTCACACCC	ACTGCCTGAG	ACAGCTGTGC	GAGGCTATCG	GCGCTTCCTC	1380
AGCTGTAGTC	CTGAGAGTGC	AGAGGAGTAC	ATTGAGTACC	TCAAGTCAAG	TGACCGGCTG	1440
GATGAGGCCG	CCCAGCGCCT	GGCCACCGTG	GTGAACGACG	AGCGTTTTCG	GTCTAAGGCC	1500
GGCAAGTCCA	ACTACCAGCT	GTGGCACGAG	CTGTGCGACC	TCATCTCCCA	GAATCCGGAC	1560
AAGGTACAGT	CCCTCAATGT	GGACGCCATC	ATCCGCGGGG	GCCTCACCCG	CTTCACCGAC	1620
CAGCTGGGCA	AGCTCTGGTG	TTCTCTCGCC	GACTACTACA	TCCGCAGCGG	CCATTTTCGAG	1680
AAGGCTCGGG	ACGTGTACGA	GGAGGCCATC	CGGACAGTGA	TGACCGTGCG	GGACTTCACA	1740
CAGGTGTTTG	ACAGCTACGC	CCAGTTCGAG	GAGAGCATGA	TCGCTGCAAA	GATGGAGACC	1800
GCCTCGGAGC	TGGGGCGCGA	GGAGGAGGAT	GATGTGGACC	TGGAGCTGCG	CCTGGCCCCG	1860
TTGAGCAGC	TCATCAGCCG	GCGGCCCTTG	CTCCTCAACA	GCGTCTTGCT	GCGCCAAAAC	1920
CCACACCACG	TGCACGAGTG	GCACAAGCGT	GTCGCCCTGC	ACCAGGGCCG	CCCCCGGGAG	1980
ATCATCAACA	CCTACACAGA	GGCTGTGCAG	ACGGTGGACC	CCTTCAAGGC	CACAGGCAAG	2040
CCCCACACTC	TGTGGGTGGC	GTTTGCCAAG	TTTTATGAGG	ACAACGGACA	GCTGGACGAT	2100
GCCCGTGTC	TCCTGGAGAA	GGCCACCAAG	GTGAACCTCA	AGCAGGTGGA	TGACCTGGCA	2160
AGCGTGTGGT	GTCAGTGCGG	AGAGCTGGAG	CTCCGACACG	AGAACTACGA	TGAGGCCTTG	2220
CGGCTGCTGC	GAAAGGCCAC	GGCGCTGCCT	GCCCGCCGGG	CCGAGTACTT	TGATGGTTCA	2280
GAGCCCGTGC	AGAACCGCGT	GTACAAGTCA	CTGAAGGTCT	GGTCCATGCT	CGCCGACCTG	2340
GAGGAGAGCC	TCGGCACCTT	CCAGTCCACC	AAGGCCGTGT	ACGACCGCAT	CCTGGACCTG	2400
CGTATCGCAA	CACCCAGAT	CGTCATCAAC	TATGCCATGT	TCCTGGAGGA	GCACAAGTAC	2460
TTGAGGAGA	GCTTCAAGGC	GTACGAGCGC	GGCATCTCGC	TGTTCAAGTG	GCCCAACGTG	2520
TCCGACATCT	GGAGCACCTA	CCTGACCAAA	TTCATTGCCC	GCTATGGGGG	CCGCAAGCTG	2580
GAGCGGGCAC	GGGACCTGTT	TGAACAGGCT	CTGGACGGCT	GCCCCCAAA	ATATGCCAAG	2640
ACCTTGTAAC	TGCTGTACGC	ACAGCTGGAG	GAGGAGTGGG	GCCTGGCCCC	GCATGCCATG	2700
GCCGTGTACG	AGCGTGCCAC	CAGGGCCGTG	GAGCCCCGCC	AGCAGTATGA	CATGTTCAAC	2760

1187

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 3306 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: Genomic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

CCCTCACTAA	AGGGAACAAA	AGCTGGAGCT	CGCGCGCCTG	CAGGTCGACA	CTAGTGGATC	60
GAAAGTTTCGT	TACGCCAAGC	TCGAAATTAA	CTCTGGGCTG	ACCCATAAAC	ATTTGTCTGA	120
TCTAGGATAT	AGTTGCGTTT	CTTGGCGGCA	GCAATCTGGA	TGAGGCGGTT	GAGGCACTGG	180
GTGGCCTGCT	GGATCAGGAC	ATCCCAGCGG	CCAGCATAGT	TCCGCTGCCG	GCGTAGGCCCC	240
ATCACCCGCA	TCTTATCCAT	GATGGCATTG	GTACCCAGGA	TGTTGTACTT	CTTGGAAGGG	300
TTGGAGGCTG	CATGTTTGAT	GGCCCATGTG	GTCTTGCCAG	CAGCAGGCAG	GCCCACCATC	360
ATCAGAATCT	CACATTCTGC	CTTGCTCTTT	GGTCCAACGG	TGCCCCGGAT	ACGCTCACTA	420
AGGGGAAGGT	GCTGGATGAA	GGTAAACCCC	GGGAGGACAG	AACAGTAGGG	CTCTGCTCTC	480
TGTCCGAAGT	TGAACTCCAC	TGCGCAATTC	TTCACCAGGA	CATGAGGATA	GAGGGCCTGA	540
CCCCCAAGG	CTTCCTTCTG	GATTTCGAAA	GCAATGCCCA	TCCACTTTCC	ATTCTTGGTA	600
AAAGACAGTT	CCACGTCATT	TCCACATTCA	AAATCCGCAA	AGCAGCCAAT	CACCGGAGAG	660
CTCTGCGGTG	CTAGGAGAGC	GGCTGGGCCC	GCAGACTGGG	GGGAAAGCTC	CGCAGCCGCA	720
GTGGGCCCCA	GGATCAGGCC	CCGCGTGGCC	TGGAGAAGCC	CAGTCTGGGC	TGGAGCGGGA	780
GCTGGACAGT	GTGGCCTTGC	GTTTCGCCCC	GGGAGCGCTG	CGAGTGTGCG	GGCCTCGGGT	840
GGATTTGCTG	AGCACCAATA	CCTCACGGTT	GCCAACCTGG	GGTTTTAGCT	CCCTTGGTTT	900
TAATCCCCTA	GGGGCGGGTG	GGGGCACGGG	AGGAAGGATG	GGCCAGCTGG	GTGCAATCCT	960
GCTGTAAGCC	AGCCATTCCCT	TGATTTCTTA	GAATTAACTA	AACGGTCGCG	CGGGAGGCCG	1020
CGGGGGGCCG	AGCGGAGCAG	CCGCGGCTGA	GGTTCCCAG	TCGGCCGCTC	GCGGTGCGC	1080
TCCGCCGCCG	GGACCCCGGC	CTCTGGCCGC	GCCGGCTCCG	GCCTCCGGGG	GGGCCGGGGC	1140
CGCCGGGACA	TGGTGCCAGT	CGCACCCCTT	CCCCCGCCGC	GCTGAGCTCG	CCGGCCGCGC	1200
CCGGGCTGGG	ACGTCGCGAG	GGGAAGATGT	TTTCCGCCCT	GAAGAAGCTG	GTGGGGTCGG	1260
ACCAGGCCCC	GGGCCGGGAC	AAGAATCATCC	CCGCCGGGCT	GCAGTCCATG	AACCAGGCGT	1320
TGCAGAGGCG	CTTCGCCAAG	GGGGTGCAGT	ACAACATGAA	GATAGTGATC	CGGGGAGACA	1380
GGAACACGGG	CAAGACAGCG	CTGTGGCACC	GCCTGCAGGG	CCGGCCGTTT	GTGGAGGAGT	1440
ACATCCCCAC	ACAGGAGATC	CAGGTCACCA	GCATCCACTG	GAGCTACAAG	ACCACGGATG	1500
ACATCGTGAA	GGTTGAAGTC	TGGGATGTAG	TAGACAAAGG	AAAATGCAAA	AAGCGAGGCG	1560
ACGGCTTAAA	GATGGAGAAC	GACCCCCAGG	AGNCGGAGTC	TGAAATGGCC	CTGGATGCTG	1620
AGTTCTCTGGA	CGTGTAACAAG	AACTGCAACG	GGTGGTTCAT	GATGTTTCGAC	ATTACCAAGC	1680
AGTGGACCTT	CAATTACATT	CTCCGGGAGC	TTCCAAAAGT	GCCCACCCAC	GTGCCAGTGT	1740
GCGTGCTGGG	GAACTACCGG	GACATGGGCG	AGCACCGAGT	CATCCTGCCG	GACGACGTGC	1800
GTGACTTCAT	CGACAACCTG	GACAGACCTC	CAGGTTCCCTC	CTACTTCCGC	TATGCTGAGT	1860
CTTCCATGAA	GAACAGCTTC	GGCCTAAAGT	ACCTTCATAA	GTTCTTCAAT	ATCCCATTTT	1920
TGCAGCTTCA	GAGGGAGACG	CTGTTGCGGC	AGCTGGAGAC	GAACCAGCTG	GACATGGACG	1980
CCACGCTGGA	GGAGCTGTCTG	GTGCAGCAGG	AGACGGAGGA	CCAGAACTAC	GGCATCTTCC	2040
TGGAGATGAT	GGAGGCTCGC	AGCCGTGGCC	ATGCGTCCCC	ACTGGCGGCC	AACGGGCAGA	2100
GCCCATCCCC	GGGCTCCCAG	TCACCAGTCC	TGCTTGCCAC	CGTGTGTCTC	ACGGGGAGCT	2160
CCAGCCCCGG	CACACCCCAG	CCCGCCCCAC	AGTGCCCCCT	CAATGCTGCC	CCACCATCCT	2220
CTGTGCCCCC	TGTACCACCC	TCAGAGGCCC	TGCCCCACC	TGCGTGCCCC	TCAGCCCCCG	2280
CCCCACGGCG	CAGCATCATC	TCTAGGCTGT	TTGGGACGTC	ACCTGCCACC	GAGGCAGCCC	2340
CTCCACCTCC	AGAGCCAGTC	CCGGCCGCAC	AGGGCCAGC	AACGGTCCAG	AGTGTGGAGG	2400
ACTTTGTTCC	TGACGACCGC	CTGGACCGCA	GCTTCCTGGA	AGACACAACC	CCC GCCAGGG	2460
ACGAGAAGAA	GGTGGGGGCC	AAGGCTGCCC	AGCAGGACAG	TGACAGTGAT	GGGGAGGCCC	2520
TGGGCGGCAA	CCCGATGGTG	GCAGGGTTCC	AGGACGATGT	GGACCTCGAA	GACCAGCCAC	2580

[illegible]

- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

GCCTGGGTTC GGTGCGGGAC

20

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

TGGTCGGGTG TTTGTGAGTG

20

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

CCTCTCCGT CTCCTCAGTG

20

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

GGATTGCTAG TCTCACAGAC

20

(2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

[illegible]

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

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- (i) SEQUENCE CHARACTERISTICS:

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

20

- (i) SEQUENCE CHARACTERISTICS:

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

20

- (i) SEQUENCE CHARACTERISTICS:

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

20

- (i) SEQUENCE CHARACTERISTICS:

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SECRET

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(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

20

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

20

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

20

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

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[illegible]

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[illegible][illegible]

- [illegible]

[illegible]

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|---|---|---|---|---|---|---|---|---|---|---|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|
| 0 | 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 | 17 | 18 | 19 | 20 | 21 | 22 | 23 | 24 | 25 | 26 | 27 | 28 | 29 | 30 | 31 | 32 | 33 | 34 | 35 | 36 | 37 | 38 | 39 | 40 | 41 | 42 | 43 | 44 | 45 | 46 | 47 | 48 | 49 | 50 | 51 | 52 | 53 | 54 | 55 | 56 | 57 | 58 | 59 | 60 | 61 | 62 | 63 | 64 | 65 | 66 | 67 | 68 | 69 | 70 | 71 | 72 | 73 | 74 | 75 | 76 | 77 | 78 | 79 | 80 | 81 | 82 | 83 | 84 | 85 | 86 | 87 | 88 | 89 | 90 | 91 | 92 | 93 | 94 | 95 | 96 | 97 | 98 | 99 |

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[illegible]

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[illegible][illegible]

- [illegible]

- (C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

GTTCCCAGAG CTTGTCTGTG

20

(2) INFORMATION FOR SEQ ID NO:37:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

GTTTGGCAGA CTCATAGTTG

20

(2) INFORMATION FOR SEQ ID NO:38:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

TAGCAGGGAG CCATGACCTG

20

(2) INFORMATION FOR SEQ ID NO:39:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

CTTGCGCCA GAAGCGAGAG

20

(2) INFORMATION FOR SEQ ID NO:40:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

20

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

TCCCCGCTGA TTCCGCCAAG

20

(2) INFORMATION FOR SEQ ID NO:42:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

CTTTTGAAT TCGGCACGAG

20

(2) INFORMATION FOR SEQ ID NO:43:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

CCCCTGGTCC GCACCAGTTC

20

(2) INFORMATION FOR SEQ ID NO:44:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

GAGAAGGGTC GGGGCGGCAG

20

(2) INFORMATION FOR SEQ ID NO:45:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs

ANNALS

[illegible]

AAATCACATC GCGTCAACAC

20

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

TAAGAGAGTC ATAGTTACTC

20

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

GCTCTAGAAG TACTCTCGAG

20

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

ACTCTGGCCA TCAGGAGATC

20

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

[illegible]

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(D) TOPOLOGY: linear

20

(D) TOPOLOGY: linear

20

(D) TOPOLOGY: linear

20

(D) TOPOLOGY: linear

20

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(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:54:

GCTACCATGC AGAGACACAG

20

(2) INFORMATION FOR SEQ ID NO:55:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:55:

CAGGCTGACA AGAAAATCAG

20

(2) INFORMATION FOR SEQ ID NO:56:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:56:

GGCACGCATA GAGGAGAGAC

20

(2) INFORMATION FOR SEQ ID NO:57:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:57:

TGGGTGATGC CTTTGCTGAC

20

(2) INFORMATION FOR SEQ ID NO:58:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

09700354 11300

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:58:

AAAACAAGAT CAAGGTGATG

20

(2) INFORMATION FOR SEQ ID NO:59:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:59:

TTGCCCACAT TGCTATGGTG

20

(2) INFORMATION FOR SEQ ID NO:60:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:60:

GACCAAGATC AGAAGTAGAG

20

(2) INFORMATION FOR SEQ ID NO:61:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:61:

- CCCCTGGGCC AATGATGTTG

20

(2) INFORMATION FOR SEQ ID NO:62:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 19 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:62:

TCTTCCCACC ATAGCAATG

19

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(2) INFORMATION FOR SEQ ID NO:68:

(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

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(2) INFORMATION FOR SEQ ID NO:69:

(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

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(2) INFORMATION FOR SEQ ID NO:70:

(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

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(2) INFORMATION FOR SEQ ID NO:71:

(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

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(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:72:

20

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:73:

20

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:74:

20

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:75:

20

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs

CONFERENCES

Year	1977	1978	1979	1980	1981	1982	1983	1984	1985	1986	1987	1988	1989	1990	1991	1992	1993	1994	1995	1996	1997	1998	1999	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017	2018	2019	2020	2021	2022	2023	2024	2025	2026	2027	2028	2029	2030																																																																																																																															
Population	1000000	1050000	1100000	1150000	1200000	1250000	1300000	1350000	1400000	1450000	1500000	1550000	1600000	1650000	1700000	1750000	1800000	1850000	1900000	1950000	2000000	2050000	2100000	2150000	2200000	2250000	2300000	2350000	2400000	2450000	2500000	2550000	2600000	2650000	2700000	2750000	2800000	2850000	2900000	2950000	3000000	3050000	3100000	3150000	3200000	3250000	3300000	3350000	3400000	3450000	3500000	3550000	3600000	3650000	3700000	3750000	3800000	3850000	3900000	3950000	4000000	4050000	4100000	4150000	4200000	4250000	4300000	4350000	4400000	4450000	4500000	4550000	4600000	4650000	4700000	4750000	4800000	4850000	4900000	4950000	5000000	5050000	5100000	5150000	5200000	5250000	5300000	5350000	5400000	5450000	5500000	5550000	5600000	5650000	5700000	5750000	5800000	5850000	5900000	5950000	6000000	6050000	6100000	6150000	6200000	6250000	6300000	6350000	6400000	6450000	6500000	6550000	6600000	6650000	6700000	6750000	6800000	6850000	6900000	6950000	7000000	7050000	7100000	7150000	7200000	7250000	7300000	7350000	7400000	7450000	7500000	7550000	7600000	7650000	7700000	7750000	7800000	7850000	7900000	7950000	8000000	8050000	8100000	8150000	8200000	8250000	8300000	8350000	8400000	8450000	8500000	8550000	8600000	8650000	8700000	8750000	8800000	8850000	8900000	8950000	9000000	9050000	9100000	9150000	9200000	9250000	9300000	9350000	9400000	9450000	9500000	9550000	9600000	9650000	9700000	9750000	9800000	9850000	9900000	9950000	10000000

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(2) INFORMATION FOR SEQ ID NO:77:

(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

20

(2) INFORMATION FOR SEQ ID NO:78:

(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

20

(2) INFORMATION FOR SEQ ID NO:79:

(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

20

(2) INFORMATION FOR SEQ ID NO:80:

(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

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20

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

TCTTGGATCT CTGGCACCTC

20

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

CCATCAGAGT GAAGGAGGAG

20

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

CCATCTTCCA CTGGTCAGAG

20

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

CTCCTTCTCT TGGATCTCTG

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- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:85:
TTACTTCAGC ACTGTTAGTC

(2) INFORMATION FOR SEQ ID NO:86:

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:86:

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(2) INFORMATION FOR SEQ ID NO:87:

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:87:

20

(2) INFORMATION FOR SEQ ID NO:88:

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:88:

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(2) INFORMATION FOR SEQ ID NO:89:

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(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

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(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

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(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

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(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

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(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:94:

CCTGCAGGTC GACACTAGTG

20

(2) INFORMATION FOR SEQ ID NO:95:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:95:

AATTGGAATG AGGAGGACTG

20

(2) INFORMATION FOR SEQ ID NO:96:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:96:

GCTCTAGAAG TACTCTCGAG

20

(2) INFORMATION FOR SEQ ID NO:97:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:97:

ATTGTATGAC AATGCACCAG

20

(2) INFORMATION FOR SEQ ID NO:98:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single

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TCCACAGAGG GCTTCATCAC

20

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

CCTGACTGGC CTAAGCACAG

20

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

AAGCCTCATA ACCACCAGTG

20

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

TGTCAACGGT GACAAGTGTG

20

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

TTGTACACCA GCTGCAGGTC

20

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:116:

CGGCATGGCA ATGAGGACAC

20

(2) INFORMATION FOR SEQ ID NO:117:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:117:

AGGACGAGAT GGACCTCCAG

20

(2) INFORMATION FOR SEQ ID NO:118:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:118:

CCCTCTGTCC TCTAGCCCAC

20

(2) INFORMATION FOR SEQ ID NO:119:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:119:

TCTTGAGGGG ACTGACTCTG

20

(2) INFORMATION FOR SEQ ID NO:120:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

- (C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:138:

CGCAACAGCG TCTCCCTCTG

20

(2) INFORMATION FOR SEQ ID NO:139:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:139:

AGTACCTTCA TAAGTTCTTC

20

(2) INFORMATION FOR SEQ ID NO:140:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:140:

TCCCAGACTT CAACCTTCAC

20

(2) INFORMATION FOR SEQ ID NO:141:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:141:

AAACATCTTC CCGGTCGGAC

20

(2) INFORMATION FOR SEQ ID NO:142:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:142:

Ser	Asn	Tyr	Gln	Gln	Thr	Thr	Ile	Ser	His	Ser	Pro	Ser	Ser	Arg	Phe	
140					145					150					155	
GTG	CCA	CCA	CAG	ACA	AGC	TCT	GGG	AAC	AGA	TTT	ATG	CCA	CAG	CAA	AAT	890
Val	Pro	Pro	Gln	Thr	Ser	Ser	Gly	Asn	Arg	Phe	Met	Pro	Gln	Gln	Asn	
				160					165						170	
AGC	CCA	GTG	CCT	AGT	CCA	TAC	GCC	CCA	CAA	AGC	CCT	GCA	GGA	TAC	ATG	938
Ser	Pro	Val	Pro	Ser	Pro	Tyr	Ala	Pro	Gln	Ser	Pro	Ala	Gly	Tyr	Met	
			175					180					185			
CCA	TAT	TCC	CAT	CCT	TCA	AGT	TAC	ACA	ACA	CAT	CCA	CAG	ATG	CAA	CAA	986
Pro	Tyr	Ser	His	Pro	Ser	Ser	Tyr	Thr	Thr	His	Pro	Gln	Met	Gln	Gln	
		190					195					200				
GCA	TCG	GTA	TCA	AGT	CCC	ATT	GTT	GCA	GGT	GGT	TTG	AGA	AAC	ATA	CAT	1034
Ala	Ser	Val	Ser	Ser	Pro	Ile	Val	Ala	Gly	Gly	Leu	Arg	Asn	Ile	His	
	205					210					215					
GAT	AAT	AAA	GTT	TCT	GGT	CCG	TTG	TCT	GGC	AAT	TCA	GCT	AAT	CAT	CAT	1082
Asp	Asn	Lys	Val	Ser	Gly	Pro	Leu	Ser	Gly	Asn	Ser	Ala	Asn	His	His	
220					225					230					235	
GCT	GAT	AAT	CCT	AGA	CAT	GGT	TCA	AGT	GAG	GAC	TAC	CTA	CAC	ATG	GTG	1130
Ala	Asp	Asn	Pro	Arg	His	Gly	Ser	Ser	Glu	Asp	Tyr	Leu	His	Met	Val	
				240					245					250		
CAC	AGG	CTA	AGT	AGT	GAC	GAT	GGA	GAT	TCT	TCA	ACA	ATG	AGG	AAT	GCT	1178
His	Arg	Leu	Ser	Ser	Asp	Asp	Gly	Asp	Ser	Ser	Thr	Met	Arg	Asn	Ala	
			255					260					265			
GCA	TCT	TTT	CCC	TTG	AGA	TCT	CCA	CAG	CCA	GTA	TGC	TCC	CCT	GCT	GGA	1226
Ala	Ser	Phe	Pro	Leu	Arg	Ser	Pro	Gln	Pro	Val	Cys	Ser	Pro	Ala	Gly	
		270					275					280				
AGT	GAA	GGA	ACT	CCT	AAA	GGC	TCA	AGA	CCA	CCT	TTA	ATC	CTA	CAA	TCT	1274
Ser	Glu	Gly	Thr	Pro	Lys	Gly	Ser	Arg	Pro	Pro	Leu	Ile	Leu	Gln	Ser	
	285					290					295					
CAG	TCT	CTA	CCT	TGT	TCA	TCA	CCT	CGA	GAT	GTT	CCA	CCA	GAT	ATC	TTG	1322
Gln	Ser	Leu	Pro	Cys	Ser	Ser	Pro	Arg	Asp	Val	Pro	Pro	Asp	Ile	Leu	
300					305					310					315	
CTA	GAT	TCT	CCA	GAA	AGA	AAA	CAA	AAG	AAG	CAG	AAG	AAA	ATG	AAA	TTA	1370
Leu	Asp	Ser	Pro	Glu	Arg	Lys	Gln	Lys	Lys	Gln	Lys	Lys	Met	Lys	Leu	
				320				325					330			
GGC	AAG	GAT	GAA	AAA	GAG	CAG	AGT	GAG	AAA	GCG	GCA	ATG	TAT	GAT	ATA	1418
Gly	Lys	Asp	Glu	Lys	Glu	Gln	Ser	Glu	Lys	Ala	Ala	Met	Tyr	Asp	Ile	
			335					340				345				
ATT	AGT	TCT	CCA	TCC	AAG	GAC	TCT	ACT	AAA	CTT	ACA	TTA	AGA	CTT	TCT	1466
Ile	Ser	Ser	Pro	Ser	Lys	Asp	Ser	Thr	Lys	Leu	Thr	Leu	Arg	Leu	Ser	
			350				355					360				
CGT	GTA	AGG	TCT	TCA	GAC	ATG	GAC	CAG	CAA	GAG	GAT	ATG	ATT	TCT	GGT	1514
Arg	Val	Arg	Ser	Ser	Asp	Met	Asp	Gln	Gln	Glu	Asp	Met	Ile	Ser	Gly	

													165						170						175																															
Arg	Glu	Lys	Ala	Gly	Gly	Pro	Gln	Leu	Asp	Arg	Tyr	Val	Arg	Pro	Met																																									
													180											185											190																					
Met	Pro	Gln	Lys	Val	Gln	Leu	Glu	Val	Gly	Arg	Pro	Gln	Ala	Pro	Leu																																									
													195											200											205																					
Asn	Ser	Phe	His	Ala	Ala	Lys	Lys	Pro	Pro	Asn	Gln	Ser	Leu	Pro	Leu																																									
													210											215											220																					
Gln	Pro	Phe	Gln	Leu	Ala	Phe	Gly	His	Gln	Val	Asn	Arg	Gln	Val	Phe																																									
													225											230											235											240										
Arg	Gln	Gly	Pro	Pro	Pro	Pro	Asn	Pro	Val	Ala	Ala	Phe	Pro	Pro	Gln																																									
													245											250											255																					
Lys	Gln	Gln	Gln	Gln	Gln	Gln	Pro	Gln	Gln	Gln	Gln	Gln	Gln	Gln	Gln																																									
													260											265											270																					
Ala	Ala	Leu	Pro	Gln	Met	Pro	Leu	Phe	Glu	Asn	Phe	Tyr	Ser	Met	Pro																																									
													275											280											285																					
Gln	Gln	Pro	Ser	Gln	Gln	Pro	Gln	Asp	Phe	Gly	Leu	Gln	Pro	Ala	Gly																																									
													290											295											300																					
Pro	Leu	Gly	Gln	Ser	His	Leu	Ala	His	His	Ser	Met	Ala	Pro	Tyr	Pro																																									
													305											310											315											320										
Phe	Pro	Pro	Asn	Pro	Asp	Met	Asn	Pro	Glu	Leu	Arg	Lys	Ala	Leu	Leu																																									
													325											330											335																					
Gln	Asp	Ser	Ala	Pro	Gln	Pro	Ala	Leu	Pro	Gln	Val	Gln	Ile	Pro	Phe																																									
													340											345											350																					
Pro	Arg	Arg	Ser	Arg	Arg	Leu	Ser	Lys	Glu	Gly	Ile	Leu	Pro	Pro	Ser																																									
													355											360											365																					
Ala	Leu	Asp	Gly	Ala	Gly	Thr	Gln	Pro	Gly	Gln	Glu	Ala	Thr	Gly	Asn																																									
													370											375											380																					
Leu	Phe	Leu	His	His	Trp	Pro	Leu	Gln	Gln	Pro	Pro	Pro	Gly	Ser	Leu																																									
													385											390											395											400										
Gly	Gln	Pro	His	Pro	Glu	Ala	Leu	Gly	Phe	Pro	Leu	Glu	Leu	Arg	Glu																																									
													405											410											415																					
Ser	Gln	Leu	Leu	Pro	Asp	Gly	Glu	Arg	Leu	Ala	Pro	Asn	Gly	Arg	Glu																																									
													420											425											430																					
Arg	Glu	Ala	Pro	Ala	Met	Gly	Ser	Glu	Glu	Gly	Met	Arg	Ala	Val	Ser																																									
													435											440											445																					
Thr	Gly	Asp	Cys	Gly	Gln	Val	Leu	Arg	Gly	Gly	Val	Ile	Gln	Ser	Thr																																									
													450											455											460																					
Arg	Arg	Arg	Arg	Arg	Ala	Ser	Gln	Glu	Ala	Asn	Leu	Leu	Thr	Leu	Ala																																									
													465											470											475											480										
Gln	Lys	Ala	Val	Glu	Leu	Ala	Ser	Leu	Gln	Asn	Ala	Lys	Asp	Gly	Ser																																									
													485											490											495																					
Gly	Ser	Glu	Glu	Lys	Arg	Lys	Ser	Val	Leu	Ala	Ser	Thr	Thr	Lys	Cys																																									
													500											505											510																					
Gly	Val	Glu	Phe	Ser	Glu	Pro	Ser	Leu	Ala	Thr	Lys	Arg	Ala	Arg	Glu																																									
													515											520											525																					
Asp	Ser	Gly	Met	Val	Pro	Leu	Ile	Ile	Pro	Val	Ser	Val	Pro	Val	Arg																																									
													530											535											540																					
Thr	Val	As																																																						

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Ala	Gly	Thr	Phe	Ile	Ala	Pro	Pro	Val	Tyr	Ser	Asn	Ile	Thr	Pro	Tyr	625	630	635	640
Gln	Ser	His	Leu	Arg	Ser	Pro	Val	Arg	Leu	Ala	Asp	His	Pro	Ser	Glu	645	650	655	
Arg	Ser	Phe	Glu	Leu	Pro	Pro	Tyr	Thr	Pro	Pro	Pro	Ile	Leu	Ser	Pro	660	665	670	
Val	Arg	Glu	Gly	Ser	Gly	Leu	Tyr	Phe	Asn	Ala	Ile	Ile	Ser	Thr	Ser	675	680	685	
Thr	Ile	Pro	Ala	Pro	Pro	Pro	Ile	Thr	Pro	Lys	Ser	Ala	His	Arg	Thr	690	695	700	
Leu	Leu	Arg	Thr	Asn	Ser	Ala	Glu	Val	Thr	Pro	Pro	Val	Leu	Ser	Val	705	710	715	720
Met	Gly	Glu	Ala	Thr	Pro	Val	Ser	Ile	Glu	Pro	Arg	Ile	Asn	Val	Gly	725	730	735	
Ser	Arg	Phe	Gln	Ala	Glu	Ile	Pro	Leu	Met	Arg	Asp	Arg	Ala	Leu	Ala	740	745	750	
Ala	Ala	Asp	Pro	His	Lys	Ala	Asp	Leu	Val	Trp	Gln	Pro	Trp	Glu	Asp	755	760	765	
Leu	Glu	Ser	Ser	Arg	Glu	Lys	Gln	Arg	Gln	Val	Glu	Asp	Leu	Leu	Thr	770	775	780	
Ala	Ala	Cys	Ser	Ser	Ile	Phe	Pro	Gly	Ala	Gly	Thr	Asn	Gln	Glu	Leu	785	790	795	800
Ala	Leu	His	Cys	Leu	His	Glu	Ser	Arg	Gly	Asp	Ile	Leu	Glu	Thr	Leu	805	810	815	
Asn	Lys	Leu	Leu	Leu	Lys	Lys	Pro	Leu	Arg	Pro	His	Asn	His	Pro	Leu	820	825	830	
Ala	Thr	Tyr	His	Tyr	Thr	Gly	Ser	Asp	Gln	Trp	Lys	Met	Ala	Glu	Arg	835	840	845	
Lys	Leu	Phe	Asn	Lys	Gly	Ile	Ala	Ile	Tyr	Lys	Lys	Asp	Phe	Phe	Leu	850	855	860	
Val	Gln	Lys	Leu	Ile	Gln	Thr	Lys	Thr	Val	Ala	Gln	Cys	Val	Glu	Phe	865	870	875	880
Tyr	Tyr	Thr	Tyr	Lys	Lys	Gln	Val	Lys	Ile	Gly	Arg	Asn	Gly	Thr	Leu	885	890	895	
Thr	Phe	Gly	Asp	Val	Asp	Thr	Ser	Asp	Glu	Lys	Ser	Ala	Gln	Glu	Glu	900	905	910	
Val	Glu	Val	Asp	Ile	Lys	Thr	Ser	Gln	Lys	Phe	Pro	Arg	Val	Pro	Leu	915	920	925	
Pro	Arg	Arg	Glu	Ser	Pro	Ser	Glu	Glu	Arg	Leu	Glu	Pro	Lys	Arg	Glu	930	935	940	
Val	Lys	Glu	Pro	Arg	Lys	Glu	Gly	Glu	Glu	Glu	Val	Pro	Glu	Ile	Gln	945	950	955	960
Glu	Lys	Glu	Glu	Gln	Glu	Glu	Gly	Arg	Glu	Arg	Ser	Arg	Arg	Ala	Ala	965	970	975	
Ala	Val	Lys	Ala	Thr	Gln	Thr	Leu	Gln	Ala	Asn	Glu	Ser	Ala	Ser	Asp	980	985	990	
Ile	Leu	Ile	Leu	Arg	Ser	His	Glu	Ser	Asn	Ala	Pro	Gly	Ser	Ala	Gly	995	1000	1005	
Gly	Gln	Ala	Ser	Glu	Lys	Pro	Arg	Glu	Gly	Thr	Gly	Lys	Ser	Arg	Arg	1010	1015	1020	
Ala	Leu	Pro	Phe	Ser	Glu	Lys	Lys	Lys	Lys	Lys	Gln	Lys	Ala			1025	1030	1035	

(2) INFORMATION FOR SEQ ID NO:152:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 849 amino acids

(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:152:

Ile	Arg	His	Glu	Val	Ser	Phe	Leu	Trp	Asn	Thr	Glu	Ala	Ala	Cys	Pro	1	5	10	15
Ile	Gln	Thr	Thr	Thr	Asp	Thr	Asp	Gln	Ala	Cys	Ser	Ile	Arg	Asp	Pro	20	25	30	
Asn	Ser	Gly	Phe	Val	Phe	Asn	Leu	Asn	Pro	Leu	Asn	Ser	Ser	Gln	Gly	35	40	45	
Tyr	Asn	Val	Ser	Gly	Ile	Gly	Lys	Ile	Phe	Met	Phe	Asn	Val	Cys	Gly	50	55	60	
Thr	Met	Pro	Val	Cys	Gly	Thr	Ile	Leu	Gly	Lys	Pro	Ala	Ser	Gly	Cys	65	70	75	80
Glu	Ala	Glu	Thr	Gln	Thr	Glu	Glu	Leu	Lys	Asn	Trp	Lys	Pro	Ala	Arg	85	90	95	
Pro	Val	Gly	Ile	Glu	Lys	Ser	Leu	Gln	Leu	Ser	Thr	Glu	Gly	Phe	Ile	100	105	110	
Thr	Leu	Thr	Tyr	Lys	Gly	Pro	Leu	Ser	Ala	Lys	Gly	Thr	Ala	Asp	Ala	115	120	125	
Phe	Ile	Val	Arg	Phe	Val	Cys	Asn	Asp	Asp	Val	Tyr	Ser	Gly	Pro	Leu	130	135	140	
Lys	Phe	Leu	His	Gln	Asp	Ile	Asp	Ser	Gly	Gln	Gly	Ile	Arg	Asn	Thr	145	150	155	160
Tyr	Phe	Glu	Phe	Glu	Thr	Ala	Leu	Ala	Cys	Val	Pro	Ser	Pro	Val	Asp	165	170	175	
Cys	Gln	Val	Thr	Asp	Leu	Ala	Gly	Asn	Glu	Tyr	Asp	Leu	Thr	Gly	Leu	180	185	190	
Ser	Thr	Val	Arg	Lys	Pro	Trp	Thr	Ala	Val	Asp	Thr	Ser	Val	Asp	Gly	195	200	205	
Arg	Lys	Arg	Thr	Phe	Tyr	Leu	Ser	Val	Cys	Asn	Pro	Leu	Pro	Tyr	Ile	210	215	220	
Pro	Gly	Cys	Gln	Gly	Ser	Ala	Val	Gly	Ser	Cys	Leu	Val	Ser	Glu	Gly	225	230	235	240
Asn	Ser	Trp	Asn	Leu	Gly	Val	Val	Gln	Met	Ser	Pro	Gln	Ala	Ala	Ala	245	250	255	
Asn	Gly	Ser	Leu	Ser	Ile	Met	Tyr	Val	Asn	Gly	Asp	Lys	Cys	Gly	Asn	260	265	270	
Gln	Arg	Phe	Ser	Thr	Arg	Ile	Thr	Phe	Glu	Cys	Ala	Gln	Ile	Ser	Gly	275	280	285	
Ser	Pro	Ala	Phe	Gln	Leu	Gln	Asp	Gly	Cys	Glu	Tyr	Val	Phe	Ile	Trp	290	295	300	
Arg	Thr	Val	Glu	Ala	Cys	Pro	Val	Val	Arg	Val	Glu	Gly	Asp	Asn	Cys	305	310	315	320
Glu	Val	Lys	Asp	Pro	Arg	His	Gly	Asn	Leu	Tyr	Asp	Leu	Lys	Pro	Leu	325	330	335	
Gly	Leu	Asn	Asp	Thr	Ile	Val	Ser	Ala	Gly	Glu	Tyr	Thr	Tyr	Tyr	Phe	340	345	350	
Arg	Val	Cys	Gly	Lys	Leu	Ser	Ser	Asp	Val	Cys	Pro	Thr	Ser	Asp	Lys	355	360	365	
Ser	Lys	Val	Val	Ser	Ser	Cys	Gln	Glu	Lys	Arg	Glu	Pro	Gln	Gly	Phe	370	375	380	
His	Lys	Val	Ala	Gly	Leu	Leu	Thr	Gln	Lys	Leu	Thr	Tyr	Glu	Asn	Gly	385	390	395	400

[illegible]

(A) LENGTH: 852 amino acids

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

Met 1	Ala	Arg	Leu	Ser 5	Arg	Pro	Glu	Arg	Pro 10	Asp	Leu	Val	Phe	Glu 15	
Glu	Asp	Leu	Pro 20	Tyr	Glu	Glu	Glu	Ile 25	Met	Arg	Asn	Gln	Phe 30	Ser	Val
Lys	Cys	Trp 35	Leu	His	Tyr	Ile	Glu 40	Phe	Lys	Gln	Gly	Ala 45	Pro	Lys	Pro
Arg	Leu 50	Asn	Gln	Leu	Tyr	Glu 55	Arg	Ala	Leu	Lys	Leu 60	Leu	Pro	Cys	Ser
Tyr 65	Lys	Leu	Trp	Tyr	Arg 70	Tyr	Leu	Lys	Ala	Arg 75	Arg	Ala	Gln	Val	Lys 80
His	Arg	Cys	Val	Thr 85	Asp	Pro	Ala	Tyr	Glu 90	Asp	Val	Asn	Asn 95	Cys	His
Glu	Arg	Ala 100	Phe	Val	Phe	Met	His 105	Lys	Met	Pro	Arg	Leu 110	Trp	Leu	Asp
Tyr	Cys	Gln 115	Phe	Leu	Met	Asp	Gln 120	Gly	Arg	Val	Thr	His 125	Thr	Arg	Arg
Thr	Phe 130	Asp	Arg	Ala	Leu 135	Arg	Ala	Leu	Pro	Ile	Thr 140	Gln	His	Ser	Arg
Ile 145	Trp	Pro	Leu	Tyr 150	Leu	Arg	Phe	Leu	Arg	Ser 155	His	Pro	Leu	Pro	Glu 160
Thr	Ala	Val	Arg	Gly 165	Tyr	Arg	Arg	Phe	Leu 170	Lys	Leu	Ser	Pro	Glu 175	Ser
Ala	Glu	Glu 180	Tyr	Ile	Glu	Tyr	Leu 185	Lys	Ser	Ser	Asp	Arg 190	Leu	Asp	Glu
Ala	Ala 195	Gln	Arg	Leu	Ala	Thr	Val 200	Val	Asn	Asp	Glu 205	Arg	Phe	Val	Ser
Lys 210	Ala	Gly	Lys	Ser	Asn 215	Tyr	Gln	Leu	Trp	His 220	Glu	Leu	Cys	Asp	Leu
Ile 225	Ser	Gln	Asn	Pro 230	Asp	Lys	Val	Gln	Ser	Leu 235	Asn	Val	Asp	Ala	Ile 240
Ile	Arg	Gly	Gly 245	Leu	Thr	Arg	Phe	Thr	Asp 250	Gln	Leu	Gly	Lys	Leu 255	Trp
Cys	Ser	Leu 260	Ala	Asp	Tyr	Tyr	Ile 265	Arg	Ser	Gly	His 270	Phe	Glu	Lys	Ala
Arg	Asp 275	Val	Tyr	Glu	Glu	Ala 280	Ile	Arg	Thr	Val	Met 285	Thr	Val	Arg	Asp
Phe	Thr 290	Gln	Val	Phe	Asp 295	Ser	Tyr	Ala	Gln	Phe 300	Glu	Glu	Ser	Met	Ile
Ala 305	Ala	Lys	Met	Glu 310	Thr	Ala	Ser	Glu	Leu	Gly 315	Arg	Glu	Glu	Glu	Asp 320
Asp	Val	Asp	Leu 325	Glu	Leu	Arg	Leu 330	Ala	Arg	Phe 335	Glu	Gln	Leu	Ile	Ser
Arg	Arg	Pro 340	Leu	Leu	Asn 345	Ser	Val	Leu	Leu	Arg 350	Gln	Asn	Pro	His	

	1970	1971	1972	1973	1974	1975	1976	1977	1978	1979	1980	1981	1982	1983	1984	1985	1986	1987	1988	1989	1990	1991	1992	1993	1994	1995	1996	1997	1998	1999	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017	2018	2019	2020	2021	2022	2023	2024	2025	2026	2027	2028	2029	2030	2031	2032	2033	2034	2035	2036	2037	2038	2039	2040	2041	2042	2043	2044	2045	2046	2047	2048	2049	2050	2051	2052	2053	2054	2055	2056	2057	2058	2059	2060	2061	2062	2063	2064	2065	2066	2067	2068	2069	2070	2071	2072	2073	2074	2075	2076	2077	2078	2079	2080	2081	2082	2083	2084	2085	2086	2087	2088	2089	2090	2091	2092	2093	2094	2095	2096	2097	2098	2099	2100	2101	2102	2103	2104	2105	2106	2107	2108	2109	2110	2111	2112	2113	2114	2115	2116	2117	2118	2119	2120	2121	2122	2123	2124	2125	2126	2127	2128	2129	2130	2131	2132	2133	2134	2135	2136	2137	2138	2139	2140	2141	2142	2143	2144	2145	2146	2147	2148	2149	2150	2151	2152	2153	2154	2155	2156	2157	2158	2159	2160	2161	2162	2163	2164	2165	2166	2167	2168	2169	2170	2171	2172	2173	2174	2175	2176	2177	2178	2179	2180	2181	2182	2183	2184	2185	2186	2187	2188	2189	2190	2191	2192	2193	2194	2195	2196	2197	2198	2199	2200	2201	2202	2203	2204	2205	2206	2207	2208	2209	2210	2211	2212	2213	2214	2215	2216	2217	2218	2219	2220	2221	2222	2223	2224	2225	2226	2227	2228	2229	2230	2231	2232	2233	2234	2235	2236	2237	2238	2239	2240	2241	2242	2243	2244	2245	2246	2247	2248	2249	2250	2251	2252	2253	2254	2255	2256	2257	2258	2259	2260	2261	2262	2263	2264	2265	2266	2267	2268	2269	2270	2271	2272	2273	2274	2275	2276	2277	2278	2279	2280	2281	2282	2283	2284	2285	2286	2287	2288	2289	2290	2291	2292	2293	2294	2295	2296	2297	2298	2299	2300	2301	2302	2303	2304	2305	2306	2307	2308	2309	2310	2311	2312	2313	2314	2315	2316	2317	2318	2319	2320	2321	2322	2323	2324	2325	2326	2327	2328	2329	2330	2331	2332	2333	2334	2335	2336	2337	2338	2339	2340	2341	2342	2343	2344	2345	2346	2347	2348	2349	2350	2351	2352	2353	2354	2355	2356	2357	2358	2359	2360	2361	2362	2363	2364	2365	2366	2367	2368	2369	2370	2371	2372	2373	2374	2375	2376	2377	2378	2379	2380	2381	2382	2383	2384	2385	2386	2387	2388	2389	2390	2391	2392	2393	2394	2395	2396	2397	2398	2399	2400	2401	2402	2403	2404	2405	2406	2407	2408	2409	2410	2411	2412	2413	2414	2415	2416	2417	2418	2419	2420	2421	2422	2
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(i) SEQUENCE CHARACTERISTICS:

(B) TYPE: amino acid

(D) TOPOLOGY: linear

Met 1	Phe	Ser	Ala	Leu 5	Lys	Lys	Leu	Val 10	Gly	Ser	Asp	Gln	Ala	Pro 15	Gly
Arg	Asp	Lys	Asn 20	Ile	Pro	Ala	Gly	Leu 25	Gln	Ser	Met	Asn	Gln	Ala	Leu
Gln	Arg	Arg 35	Phe	Ala	Lys	Gly	Val 40	Gln	Tyr	Asn	Met	Lys 45	Ile	Val	Ile
Arg	Gly 50	Asp	Arg	Asn	Thr	Gly 55	Lys	Thr	Ala	Leu	Trp 60	His	Arg	Leu	Gln
Gly 65	Arg	Pro	Phe	Val 70	Glu	Glu	Tyr	Ile	Pro	Thr 75	Gln	Glu	Ile	Gln	Val
Thr	Ser	Ile	His 85	Trp	Ser	Tyr	Lys	Thr 90	Thr	Asp	Asp	Ile	Val	Lys 95	Val
Glu	Val	Trp	Asp 100	Val	Val	Asp	Lys	Gly 105	Lys	Cys	Lys	Lys	Arg 110	Gly	Asp
Gly	Leu 115	Lys	Met	Glu	Asn	Asp	Pro 120	Gln	Glu	Xaa	Glu 125	Ser	Glu	Met	Ala
Leu	Asp 130	Ala	Glu	Phe	Leu	Asp 135	Val	Tyr	Lys	Asn 140	Cys	Asn	Gly	Val	Val
Met 145	Met	Phe	Asp	Ile 150	Thr	Lys	Gln	Trp	Thr	Phe 155	Asn	Tyr	Ile	Leu	Arg
Glu	Leu	Pro	Lys 165	Val	Pro	Thr	His	Val 170	Pro	Val	Cys	Val	Leu	Gly 175	Asn
Tyr	Arg	Asp	Met 180	Gly	Glu	His	Arg	Val 185	Ile	Leu	Pro	Asp	Asp	Val	Arg
Asp	Phe 195	Ile	Asp	Asn	Leu	Asp	Arg 200	Pro	Pro	Gly	Ser 205	Ser	Tyr	Phe	Arg
Tyr	Ala 210	Glu	Ser	Ser	Met	Lys 215	Asn	Ser	Phe	Gly 220	Leu	Lys	Tyr	Leu	His
Lys 225	Phe	Phe	Asn	Ile 230	Pro	Phe	Leu	Gln	Leu	Gln 235	Arg	Glu	Thr	Leu	Leu
Arg	Gln	Leu	Glu 245	Thr	Asn	Gln	Leu	Asp	Met 250	Asp	Ala	Thr	Leu	Glu 255	Glu
Leu	Ser	Val	Gln 260	Gln	Glu	Thr	Glu	Asp 265	Gln	Asn	Tyr	Gly	Ile 270	Phe	Leu
Glu	Met 275	Met	Glu	Ala	Arg	Ser	Arg 280	Gly	His	Ala	Ser 285	Pro	Leu	Ala	Ala
Asn	Gly 290	Gln	Ser	Pro	Ser	Pro 295	Gly	Ser	Gln	Ser 300	Pro	Val	Leu	Pro	Ala
Pro	Ala	Val	Ser	Thr	Gly	Ser	Ser	Ser	Pro	Gly	Thr	Pro	Gln	Pro	Ala

